ICON: Improving Inter-Report Consistency in Radiology Report Generation via Lesion-aware Mixup Augmentation

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Abstract

Previous research on radiology report generation has made significant progress in terms of increasing the clinical accuracy of generated reports. In this paper, we emphasize another crucial quality that it should possess, i.e., interreport consistency, which refers to the capability of generating consistent reports for semantically equivalent radiographs. This quality is even of greater significance than the overall report accuracy in terms of ensuring the system's credibility, as a system prone to providing conflicting results would severely erode users' trust. Regrettably, existing approaches struggle to maintain inter-report consistency, exhibiting biases towards common patterns and susceptibility to lesion variants. To address this issue, we propose ICON, which Improves the inter-report **CON**sistency of radiology report generation. Aiming to enhance the system's ability to capture similarities in semantically equivalent lesions, our approach first involves extracting lesions from input images and examining their characteristics. Then, we introduce a lesion-aware mixup technique to ensure that the representations of the semantically equivalent lesions align with the same attributes, achieved through a linear combination during the training phase. Extensive experiments on three publicly available chest X-ray datasets verify the effectiveness of our approach, both in terms of improving the consistency and accuracy of the generated reports¹.

1 Introduction

Being part of the diagnostic process, radiology report generation (Shin et al., 2016; Zhang et al., 2017; Jing et al., 2018) has garnered significant attention within the research community, due to

¹Our codes and model checkpoints are available at https: //github.com/wjhou/ICon



Figure 1: Given two semantically equivalent cases (i.e., Case A and Case B), an example to illustrate the difference between three radiology report generation systems: a consistent and accurate system (i.e., System α) and a consistently inaccurate system (i.e., System β), and an inconsistent system (i.e., System γ).

its large potential to alleviate the heavy strain of radiologists. Recent research (Nishino et al., 2022; Tanida et al., 2023; Hou et al., 2023b) has made noteworthy progress in enhancing the clinical accuracy of the generated reports.

However, constructing a credible report generation system goes beyond the overall accuracy. There is another crucial quality for report generation systems that has been largely overlooked in the existing literature of medical report generation, which is, *inter-report consistency* (Elazar et al., 2021). To illustrate the disparity between accuracy and inter-report consistency, we exemplify two semantically equivalent cases as shown in Figure 1, where they share similar observations and reports. Specifically, System α demonstrates the ability to maintain both inter-report consistency and factual accuracy for two similar cases (i.e., "small bilateral pleural effusions" for positive Pleural Effu-

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sion), whereas other systems (i.e., β and γ) fail to meet these criteria. These systems might have overfitted to ordinary cases and could be vulnerable to noise or attack. In terms of enhancing the system's credibility, inter-report consistency might even hold greater significance than the overall accuracy, since a system prone to providing conflicting results would severely undermine users' trust (Qayyum et al., 2020; Asan et al., 2020). Regrettably, existing report generation systems struggle to maintain this important quality. They tend to exhibit biases towards common patterns, primarily describing normal observations and are susceptible to lesion variants and context noise (Chen et al., 2020; Qin and Song, 2022; Ma et al., 2021; Kaviani et al., 2022). We argue that this is largely due to their limited capability of capturing shared attributes of similar patterns, which arises from the data scarcity of distributed lesions and their semantically equivalent variants, rendering it challenging for neural models to accurately locate and describe abnormalities.

In this paper, we propose ICON, which aims to Improves inter-report <u>CON</u>sistency of radiology report generation. Our proposed method involves first extracting lesions from given input images, followed by examining the attributes of these lesions. Subsequently, both the radiographs and their associated attributes are utilized as inputs for report generation. To further enhance the inter-report consistency, we introduce a lesion-aware mixup technique by learning from linearly combined lesions and synthesized attributes that belong to the same observation. In summary, the contributions of this paper are as follows:

- To the best of our knowledge, we are the first to introduce *inter-report consistency* in radiology report generation. To this end, we devise two metrics (CON and R-CON) to measure such consistency.
- We propose ICON, which improves both the *consistency* and *accuracy* in radiology report generation by capturing abnormalities at the region level. ICON only requires coarse-grained labels (i.e., image labels) for training to extract lesions², in contrast to previous

methods that require fine-grained labels (i.e., bounding boxes).

• Extensive experiments are conducted on three publicly available datasets, and the results demonstrate the effectiveness of ICON in terms of improving both the consistency and accuracy of the generated reports.

2 Preliminaries

2.1 **Problem Formulation**

Given a set of radiographs $\mathcal{X} = \{X_1, \dots, X_L\}$ in one study, along with its historical records $\mathcal{X}^p = \{X^p_1, \dots, X^p_{|p|}\}$ (or $\mathcal{X}^p = \emptyset$ if no historical records are available), and its report \mathcal{Y} , the task of radiology report generation (RRG) is to generate the report \mathcal{Y} based on \mathcal{X} and \mathcal{X}^p . We elaborate on the justification for using historical records as context in Appendix A.6. Our proposed method, denoted as ICON, decomposes the RRG task into two stages: Lesion Extraction (Stage 1) and Report Generation (Stage 2). Specifically, given the input images \mathcal{X} , ICON first extracts M region candidates $\mathcal{R} = \{R_1, \ldots, R_M\}$ and then classifies regions as lesions $\mathcal{Z} = \{Z_1, \ldots, Z_{|O|}\}$, where O denotes the observations. Subsequently, in Stage 2, ICON generates a report based on the input images \mathcal{X} , historical records \mathcal{X}^p , and the extracted lesions \mathcal{Z} .

2.2 Observation and Attribute Annotation

Observations for Lesion Extraction. Lesion extraction requires report-level labels, and we adopt CheXbert (Smit et al., 2020) for this purpose. Specifically, CheXbert annotates a report with 14 observation categories $O = \{o_1, \ldots, o_{14}\}$, with data statistics provided in Appendix A.1. Each observation is assigned one of four statuses: *Present*, *Absent*, *Uncertain*, and *Blank*. During training and evaluation, *Present* and *Uncertain* are merged into the *Positive* category, which represents abnormal observations. Note that for the observation *No Finding*, only two statuses, *Present* or *Absent*, are applicable. Finally, observation information is utilized for lesion extraction as described in §3.2.

Attributes for Lesion-Attribute Alignment. After extracting observations, we further extract entities that represent their characteristics. Specifically, we adopt the attributes released by Hou et al. (2023a)³, which are entities (with a relation *modify* or *located_at*) extracted from RadGraph (Jain

²In this context, the term "lesion" generally refers to a specific abnormality. It encompasses most observation categories, excluding *Support Devices*, *Cardiomegaly*, and *Enlarged Cardiomediastinum*. For simplicity, we consider all corresponding regions as lesions.

³The attributes are available at https://github.com/ wjhou/Recap.

et al., 2021) using PMI (Church and Hanks, 1990). We select the top 30 attributes for each observation and list some of them in Appendix A.2 for a better understanding. These attributes are then utilized for lesion-attribute alignment as described in §3.3.

2.3 Inter-Report Consistency Metrics

To assess the inter-report consistency of a model, we introduce two metrics, CON and R-CON, inspired by Elazar et al. (2021). Semantically equivalent samples should have high observation and entity similarity, which we calculate using the Overlap Coefficient (Simpson, 1943): $\operatorname{Overlap}(A, B) = \frac{|A \cap B|}{\min(|A|, |B|)}$. For a report Q_i and its relevant reports \mathcal{K}_i = $\{K_{i,1},\ldots,K_{i,N}\}$, when the observation similarity satisfies $\operatorname{Overlap}(O_{Q_i}, O_{K_{i,j}}) \geq 0.75$ and the entity similarity satisfies $Overlap(Q_i, K_{i,j}) \ge 0.5$, we regard them as semantically equivalent samples. We then collect the corresponding outputs of \mathcal{K}_i from a model, denoted as $\mathcal{K}_i = \{K_{i,1}, \ldots, K_{i,N}\}.$ The similarity between two outputs \widehat{Q}_i and $\widehat{K}_{i,j}$ is:

$$\texttt{Overlap}(\widehat{Q}_i, \widehat{K}_{i,j}) = \frac{|\widehat{e}_i \cap \widehat{e}_j|}{\min(|\widehat{e}_i|, |\widehat{e}_j|)},$$

where \hat{e}_i and \hat{e}_j are entities and attributes in \hat{Q}_i and $\hat{K}_{i,j}$, respectively. The inter-report consistency is then defined as:

$$\operatorname{CON}(\widehat{Q}_i,\widehat{\mathcal{K}}_i) = \frac{1}{N}\sum_{j=1}^{N}\operatorname{Overlap}(\widehat{Q}_i,\widehat{K}_{i,j})$$

Since CON only considers inter-report consistency without accounting for reference quality, we introduce R-CON, which considers both consistency and accuracy:

$$\mathbf{R}-\mathbf{CON}(\widehat{Q}_i,\widehat{\mathcal{K}}_i)=\tau_i\cdot\mathbf{CON}(\widehat{Q}_i,\widehat{\mathcal{K}}_i),$$

where $\tau_i = \texttt{Overlap}(\hat{Q}_i, Q_i)$ is the similarity between the hypothesis and its reference.

3 Methodology

3.1 Visual Encoding

Given an image X_l , an image processor is first utilized to split X_l into N patches. Then, a visual encoder f_{θ} , e.g., Swin Transformer (Liu et al., 2021d), is employed to extract visual representations X_l and the pooler output $P_l \in \mathbb{R}^h$:

$$[\boldsymbol{P}_l, \boldsymbol{X}_l] = f_{\theta}(X_l),$$

where $X_l = \{x_{l,i}, \ldots, x_{l,N}\}$ and $x_{l,i} \in \mathbb{R}^h$ is the *i*-th visual representation.

3.2 Stage 1: Extracting Lesions via Observation Classification (ZOOMER)

Observation Classification. A ZOOMER is a visual encoder parameterized by θ_Z and trained to classify a given input \mathcal{X} into abnormal observations as mentioned in §2.2:

$$p(o_i) = \text{ZOOMER}(\mathcal{X})$$

Specifically, ZOOMER first encodes images $\mathcal{X} = \{X_1, \ldots, X_L\}$ as outlined in §3.1, and then takes the averaged pooler output for classification, following these steps:

$$[\mathbf{P}_l, \mathbf{X}_l] = f_{\theta_z}(X_l),$$
$$\mathbf{P} = \frac{1}{L} \sum \mathbf{P}_l,$$
$$p(o_i) = \sigma(\mathbf{W}_i \mathbf{P} + b_i),$$

where $W_i \in \mathbb{R}^h$ is the weight for the *i*-th observation, $b_i \in \mathbb{R}$ is its bias, and σ is the Sigmoid function.

Zooming In for Lesion Extraction. Upon completing training ZOOMER, we can use it to extract lesions without the need for object detectors (Ren et al., 2015). It is worth noting that our method does not require fine-grained labels, such as bounding boxes (Tanida et al., 2023).

For an image X_l , a sliding window with a 0.375 ratio of X_l is applied to extract M region candidates $\mathcal{R}_l = \{R_{l,1}, \ldots, R_{l,M}\}$ from X_l , as shown in the left side of Figure 2. These regions are then sequentially fed into ZOOMER for classification. Further details on the extraction of these regions can be found in Appendix A.5. The probability of a region $R_{l,j}$ being classified as an abnormal observation o_i is:

$$p_{l,j}(o_i) = \operatorname{ZOOMER}(R_{l,j}).$$

For each study, all images in \mathcal{X} are iterated, and only the region with the highest $p_{l,j}(o_i)$ is chosen as a lesion Z_i corresponding to the observation o_i . Finally, the set of lesions is denoted as $\mathcal{Z} = \{Z_1, \ldots, Z_{|O|}\}$.

Training ZOOMER. ZOOMER is optimized using the binary cross-entropy (BCE) loss. To handle the class-imbalanced issue (refer to Appendix A.1 for details), a weight factor α_j is applied for each abnormal observation, and the loss function \mathcal{L}_{S1} is:

$$\begin{split} \mathsf{BCE}(p(o_j), o_j) &= -\frac{1}{|O|} \sum_j \left[\alpha_j \cdot o_j \cdot \log p(o_j) \right. \\ & \left. + (1 - o_j) \cdot \log(1 - p(o_j)) \right], \end{split}$$

where $o_j \in \{0,1\}$ is the label, $\alpha_j = 1 +$



Figure 2: Overview of the ICON framework, which first extracts lesions and then generates reports. Attributes are extracted from RadGraph (Jain et al., 2021).

 $\log\left(\frac{|\mathcal{D}_{\text{train}}|-w_j}{w_j}\right)$, and $|\mathcal{D}_{\text{train}}|$ and w_j are the number of samples and the number of *j*-th observations in the training set, respectively.

3.3 Stage 2: Inspecting Lesions (INSPECTOR)

Inspecting Lesions with Attributes. Given that lesions of the same observation can exhibit different characteristics, it is crucial to inspect each lesion and match it with corresponding attributes (§2.2) to differentiate it from other variations. Specifically, an INSPECTOR is a visual encoder parameterized by θ_I , similar to §3.2. INSPECTOR(P^p , P, Z_j) takes prior and current visit chest X-rays as context, along with a lesion region as input:

$$egin{aligned} & [m{P}_{Z_j}, m{Z}_j] = f_{m{ heta}_I}(Z_j), \ & p_j(a_k) = \sigma(\texttt{MLP}(m{P}^p, m{P}, m{P}_{Z_j})). \end{aligned}$$

where MLP is a two-layer perceptron with nonlinear activation, and P^p , P, $P_{Z_j} \in \mathbb{R}^h$ are pooler outputs of prior images, current images, and the lesion, respectively. Concurrently, the lesion features $\mathcal{Z} = \{Z_1, \ldots, Z_{|O|}\}$ are collected for report generation. For image encoding, we use another visual encoder f_{θ_V} to encode \mathcal{X} into \mathcal{X} and \mathcal{X}^p into \mathcal{X}^p . By inspecting lesion-level features, ICON can capture fine-grained details which are beneficial for generating consistent outputs.

Lesion-aware Mixup. To further improve the consistency of the generated outputs, we adopt the mixup augmentation method (Zhang et al., 2018) and devise a lesion-aware mixup during the training phase. Specifically, for a lesion-attribute pair (Z_j, A_j) , we retrieve a similar pair (Z_k, A_k) with



Figure 3: Overview of our proposed lesion-aware mixup augmentation.

the same observation from the training data based on report similarity. These lesions are synthesized by a linear combination, as illustrated in Figure 3:

$$Z_j^* = \lambda Z_j + (1 - \lambda) Z_k,$$

where λ is set to 0.75. Note that during training, Z_j^* is used for both INSPECTOR and GENERATOR. **Training INSPECTOR.** Similar to §3.2, we adopt a linearly combined BCE loss to optimize INSPECTOR:

$$\mathcal{L}_{\mathbf{I}} = \lambda \mathsf{BCE}_j + (1 - \lambda) \mathsf{BCE}_k,$$

where BCE_j and BCE_k take A_j and A_k as their respective labels. Notably, only the attributes that are shared between Z_j and Z_k are fully optimized. Consequently, our lesion-aware mixup technique facilitates the improvement of output consistency for two semantically equivalent lesions.

| Datasat | Modol | | NLG Metrics | | | | | C | E Metrie | cs |
|---------|-----------------------------|--------------|-------------|-------|------------|-------|-------|--------------|----------|----------------|
| Dataset | WIGUEI | B-1 | B-2 | B-3 | B-4 | MTR | R-L | Р | R | \mathbf{F}_1 |
| | R2GEN | 0.290 | 0.157 | 0.093 | 0.061 | 0.105 | 0.208 | 0.266 | 0.320 | 0.272 |
| MIMIC | R2GENCMN | 0.264 | 0.140 | 0.085 | 0.056 | 0.098 | 0.212 | 0.290 | 0.319 | 0.280 |
| | ORGAN | 0.314 | 0.180 | 0.114 | 0.078 | 0.120 | 0.234 | 0.271 | 0.342 | 0.293 |
| -ADN | RECAP | <u>0.321</u> | 0.182 | 0.116 | 0.080 | 0.120 | 0.223 | <u>0.300</u> | 0.363 | 0.305 |
| | ICON (Ours) | 0.337 | 0.195 | 0.126 | 0.086 | 0.129 | 0.236 | 0.332 | 0.430 | 0.360 |
| | R2Gen | 0.353 | 0.218 | 0.145 | 0.103 | 0.142 | 0.270 | 0.333 | 0.273 | 0.276 |
| | R2GENCMN | 0.353 | 0.218 | 0.148 | 0.106 | 0.142 | 0.278 | 0.344 | 0.275 | 0.278 |
| | $\mathcal{M}^2 \mathrm{Tr}$ | 0.378 | 0.232 | 0.154 | 0.107 | 0.145 | 0.272 | 0.240 | 0.428 | 0.308 |
| | KNOWMAT | 0.363 | 0.228 | 0.156 | 0.115 | _ | 0.284 | 0.458 | 0.348 | 0.371 |
| | CMM-RL | 0.381 | 0.232 | 0.155 | 0.109 | 0.151 | 0.287 | 0.342 | 0.294 | 0.292 |
| MIMIC | CMCA | 0.360 | 0.227 | 0.156 | 0.117 | 0.148 | 0.287 | 0.444 | 0.297 | 0.356 |
| CVP | KiUT | 0.393 | 0.243 | 0.159 | 0.113 | 0.160 | 0.285 | 0.371 | 0.318 | 0.321 |
| -CAR | DCL | _ | _ | _ | 0.109 | 0.150 | 0.284 | 0.471 | 0.352 | 0.373 |
| | METrans | 0.386 | 0.250 | 0.169 | 0.124 | 0.152 | 0.291 | 0.364 | 0.309 | 0.311 |
| | RGRG | 0.373 | 0.249 | 0.175 | 0.126 | 0.168 | 0.264 | 0.380 | 0.319 | 0.305 |
| | ORGAN | 0.407 | 0.256 | 0.172 | 0.123 | 0.162 | 0.293 | 0.416 | 0.418 | 0.385 |
| | Recap | 0.429 | 0.267 | 0.177 | 0.125 | 0.168 | 0.288 | 0.389 | 0.443 | <u>0.393</u> |
| | ICON (Ours) | 0.429 | 0.266 | 0.178 | 0.126 | 0.170 | 0.287 | <u>0.445</u> | 0.505 | 0.464 |

Table 1: Experimental results of our model and baselines on the MIMIC-ABN and MIMIC-CXR datasets. The best results are in **boldface**, and the <u>underlined</u> are the second-best results.

3.4 Stage 2: Generating Reports (GENERATOR)

Lesion-Attribute Alignment. To bridge the modality gap between lesion representations and attributes, we leverage a BART (Lewis et al., 2020) encoder to extract attribute representations. The attributes associated with each lesion are formulated as a prompt: $\langle s \rangle o_j \langle s \rangle A_j \langle s \rangle$, as depicted in Figure 2. Then, a cross-attention module (Vaswani et al., 2017) is inserted after every self-attention module. This module aligns the lesion representations with the attribute representations by querying visual representations using attribute representations, similar to Q-Former (Li et al., 2023a):

$$oldsymbol{H}_{j}^{a} = extsf{CrossAttention}(oldsymbol{H}_{j}^{s},oldsymbol{Z}_{j},oldsymbol{Z}_{j}),$$

where $H_j^a, H_j^s \in \mathbb{R}^h$ are the aligned attribute representation and the self-attended representation of A_j , respectively. All prompts are encoded, and the attribute representations of \mathcal{Z} are denoted as \mathcal{H}^a .

Report Generation. Given the input images \mathcal{X} , images of prior visits \mathcal{X}^p , the lesions \mathcal{Z} , and attribute \mathcal{H}^a , we utilize a BART decoder in conjunction with the Fusion-in-Decoder (FiD; (Izacard and Grave, 2021)) that simply concatenates multiple context sequences for report generation. Then, the probability of the *t*-th step is expressed as:

$$\begin{split} & \boldsymbol{h}_t = \texttt{FiD}([\boldsymbol{\mathcal{X}}; \boldsymbol{\mathcal{X}}^p; \boldsymbol{Z}; \boldsymbol{\mathcal{H}}^a], \boldsymbol{h}_{< t}), \\ & p(y_t | \boldsymbol{\mathcal{X}}, \boldsymbol{\mathcal{X}}^p, \boldsymbol{\mathcal{Z}}, \boldsymbol{\mathcal{Y}}_{< t}) = \texttt{Softmax}(\boldsymbol{W}_g \boldsymbol{h}_t + \boldsymbol{b}_g), \end{split}$$

where $h_t \in \mathbb{R}^h$ is the *t*-th hidden representation, $W_g \in \mathbb{R}^{|\mathcal{V}| \times h}$ is the weight matrix, $b_g \in \mathbb{R}^{|\mathcal{V}|}$ is the bias vector, and \mathcal{V} is the vocabulary.

Training GENERATOR. The generation process is optimized using the negative loglikelihood loss, given each token's probability $p(y_t|\mathcal{X}, \mathcal{X}^p, \mathcal{Z}, \mathcal{Y}_{< t})$:

$$\mathcal{L}_{\mathrm{G}} = -\sum_{t=1}^{T} \log p(y_t | \mathcal{X}, \mathcal{X}^p, \mathcal{Z}, \mathcal{Y}_{< t}).$$

The loss function of Stage 2 is: $\mathcal{L}_{S2} = \mathcal{L}_I + \mathcal{L}_G$.

4 Experiments

4.1 Datasets

Three public datasets are used to evaluate our models, i.e., IU X-RAY⁴ (Demner-Fushman et al., 2016), MIMIC-CXR⁵ (Johnson et al., 2019), and MIMIC-ABN (Ni et al., 2020). We follow previous research (Chen et al., 2020) to preprocess these datasets, and provide other details in Appendix A.3.

- IU X-RAY consists of 3,955 reports. We follow previous research (Chen et al., 2020) and split the dataset into train/validation/test sets with a ratio of 7:1:2.
- MIMIC-CXR consists of 377,110 chest Xray images and 227,827 reports.

⁴https://openi.nlm.nih.gov/

⁵https://physionet.org/content/mimic-cxr-jpg/
2.0.0/

| Dotocot | Model | NLG N | Aetrics | RadGraph | | | |
|---------|------------------------------|-------|---------|-----------------|------------------|----------------------|--|
| Dataset | WIUUCI | B-4 | R-L | RG _E | RG _{ER} | $RG_{\overline{ER}}$ | |
| | R2Gen | 0.120 | 0.298 | - | - | - | |
| IU | $\mathcal{M}^2 \mathrm{Tr}$ | 0.121 | 0.288 | - | _ | — | |
| X-ray | $\mathcal{T}_{\mathrm{NLL}}$ | 0.114 | _ | 0.230 | 0.202 | 0.153 | |
| | ICON | 0.098 | 0.320 | 0.342 | 0.312 | 0.246 | |
| | $\mathcal{T}_{\mathrm{NLL}}$ | 0.105 | 0.253 | 0.230 | 0.202 | 0.153 | |
| MIMIC | ORGAN | 0.123 | 0.293 | 0.303 | 0.275 | 0.199 | |
| -CXR | RECAP | 0.125 | 0.288 | 0.307 | 0.276 | 0.205 | |
| | ICON | 0.126 | 0.287 | 0.312 | 0.278 | 0.197 | |
| | | | | | | | |

Table 2: Radgraph evaluation results on the IU X-RAY and MIMIC-CXR datasets. Results of T_{NLL} are cited from Delbrouck et al. (2022).

 MIMIC-ABN is modified from the MIMIC-CXR dataset and its reports only contain abnormal part. We adopt the data-split as used in Hou et al. (2023a), and the data-split is 71,786/546/806 for train/validation/test sets.

Unlike previous research (Chen et al., 2020) which only used one view for report generation on MIMIC-CXR and MIMIC-ABN, we collect all views for each visit in experiments. The justification is provided in Appendix A.6.

4.2 Evaluation Metrics and Baselines

NLG Metrics. To assess the quality of generated reports, we adopt several natural language generation (NLG) metrics for evaluation. BLEU (Papineni et al., 2002), METEOR (Banerjee and Lavie, 2005), and ROUGE (Lin, 2004) are selected as NLG Metrics, and we use the MS-COCO caption evaluation tool⁶ to compute the results.

CE Metrics. Following previous research (Chen et al., 2020, 2021), we adopt clinical efficacy (CE) metrics to evaluate the observation-level factual accuracy, and CheXbert (Smit et al., 2020) is used in this paper. To measure the entity-level factual accuracy, we leverage the RadGraph (Jain et al., 2021; Delbrouck et al., 2022) and temporal entity matching (TEM) scores proposed by Bannur et al. (2023) for evaluation.

Consistency Metrics. CON and R-CON (§2.3) are utilized to measure the inter-report consistency. Note that entities used in measuring consistency are adopted from RadGraph (Jain et al., 2021). A MAJORITY baseline which outputs the same report for all inputs, is included.

Baselines. We compare our models with the following state-of-the-art (SOTA) baselines: R2GEN (Chen et al., 2020), R2GENCMN (Chen et al., 2021), KNOWMAT (Yang et al., 2021), \mathcal{M}^2 TR (Nooralahzadeh et al., 2021), CMM-RL (Qin and

| | ⁶ https:/ | //github.com | /tylin/ | <pre>/coco-caption</pre> |
|--|----------------------|--------------|---------|--------------------------|
|--|----------------------|--------------|---------|--------------------------|

| Model | MIMI | C-ABN | MIMIC-CXR | | |
|----------------------|-------|-------|-----------|-------|--|
| WIGUEI | CON | R-CON | CON | R-CON | |
| MAJORITY | 1.000 | _ | 1.000 | - | |
| R2Gen | 0.280 | 0.072 | 0.137 | 0.042 | |
| R2GENCMN | 0.302 | 0.091 | 0.155 | 0.049 | |
| ORGAN | 0.338 | 0.127 | 0.345 | 0.126 | |
| RECAP | 0.311 | 0.108 | 0.345 | 0.114 | |
| ICON (Ours) | 0.316 | 0.140 | 0.351 | 0.163 | |
| ICON <i>w/o</i> ZOOM | 0.183 | 0.073 | 0.175 | 0.066 | |
| ICON w/o INSPECT | 0.253 | 0.100 | 0.245 | 0.090 | |
| ICON w/o MIXUP | 0.286 | 0.119 | 0.334 | 0.156 | |

Table 3: The CON score and the R-CON score. MAJOR-ITY: outputs the same report for all inputs.

Song, 2022), CMCA (Song et al., 2022), CXR-RePaiR-Sel/2 (Endo et al., 2021), BioViL-T (Bannur et al., 2023), DCL (Li et al., 2023b), METrans (Wang et al., 2023c), KiUT (Huang et al., 2023), RGRG (Tanida et al., 2023), ORGAN (Hou et al., 2023b), and RECAP (Hou et al., 2023a).

4.3 Implementation Details

The small and tiny versions of Swin Transformer V2 (Liu et al., 2022) are used as the visual backbone for ZOOMER and INSPECTOR, respectively. The GENERATOR is initialized with the base version of BART (Lewis et al., 2020) pretrained on biomedical corpus (Yuan et al., 2022). Other parameters are randomly initialized. For Stage 2 training, the learning rate is 5e - 5 with linear decay, the batch size is 32, and the models are trained for 20 and 5 epochs on MIMIC-ABN and MIMIC-CXR with early stopping, respectively. Since the number of samples in IU X-RAY is too small to train a multimodal model, we only provide results produced by models trained on MIMIC-CXR as a reference, similar to (Delbrouck et al., 2022). For other training details, and the resources used in this paper, we list them in Appendix A.4.

5 Results

5.1 Quantitative Analysis

Inter-Report Consistency Analysis. Table 3 provides CON and R-CON scores of baselines, our model, and its ablated variants. ICON achieves the highest R-CON on both datasets, indicating the best inter-report consistency. In terms of the CON score, ICON demonstrates competitive performance when compared with the best baseline, i.e., ORGAN. We also observe that introducing mixup augmentation leads to a moderate improvement in CON, demonstrating the effectiveness of lesion-aware mixup.

NLG and Temporal Modeling Results. The NLG

| Detect | Model | Components | | NLG Metrics | | | | | CE Metrics | | | | |
|---------------|------------------|--------------|--------------|--------------|-------|-------|-------|-------|------------|-------|-------|-------|----------------|
| Dataset | WIGUEI | ZOOM | INSPECT | MIXUP | B-1 | B-2 | B-3 | B-4 | MTR | R-L | Р | R | \mathbf{F}_1 |
| | ICON | \checkmark | \checkmark | \checkmark | 0.337 | 0.195 | 0.126 | 0.086 | 0.129 | 0.236 | 0.332 | 0.430 | 0.360 |
| MIMIC | ICON w/o ZOOM | × | × | × | 0.310 | 0.181 | 0.119 | 0.084 | 0.120 | 0.243 | 0.306 | 0.353 | 0.306 |
| -ABN | ICON w/o INSPECT | \checkmark | × | × | 0.315 | 0.182 | 0.117 | 0.081 | 0.121 | 0.236 | 0.338 | 0.401 | 0.352 |
| | ICON w/o MIXUP | \checkmark | \checkmark | × | 0.335 | 0.192 | 0.124 | 0.085 | 0.129 | 0.239 | 0.332 | 0.413 | 0.356 |
| | ICON | \checkmark | \checkmark | \checkmark | 0.429 | 0.266 | 0.178 | 0.126 | 0.170 | 0.287 | 0.445 | 0.505 | 0.464 |
| MIMIC -CXR | ICON w/o ZOOM | × | × | × | 0.377 | 0.237 | 0.162 | 0.119 | 0.149 | 0.288 | 0.363 | 0.280 | 0.278 |
| | ICON w/o INSPECT | \checkmark | × | × | 0.399 | 0.248 | 0.168 | 0.122 | 0.157 | 0.287 | 0.444 | 0.447 | 0.423 |
| | ICON w/o MIXUP | \checkmark | \checkmark | × | 0.427 | 0.264 | 0.176 | 0.124 | 0.169 | 0.285 | 0.444 | 0.502 | 0.462 |

Table 4: Ablation results of our model and its variants on the MIMIC-ABN and MIMIC-CXR datasets. A \checkmark indicates that the component is included, while an \checkmark denotes that it is removed.

| Model | B-4 | R-L | \mathbf{CE} - \mathbf{F}_1 | TEM |
|--------------|-------|-------|--------------------------------|-------|
| CXR-RePaiR-2 | 0.021 | 0.143 | 0.281 | 0.125 |
| BioViL-NN | 0.037 | 0.200 | 0.283 | 0.111 |
| BioViL-T-NN | 0.045 | 0.205 | 0.290 | 0.130 |
| BioViL-AR | 0.075 | 0.279 | 0.293 | 0.138 |
| BioViL-T-AR | 0.092 | 0.296 | 0.317 | 0.175 |
| Recap | 0.118 | 0.279 | 0.400 | 0.304 |
| ICON (Ours) | 0.120 | 0.279 | 0.468 | 0.335 |

Table 5: Progression modeling results on the MIMIC-CXR dataset. Results of BioViL-* are cited from Bannur et al. (2023).

results are presented in Table 1 and the temporal modeling results are listed in Table 5. Among all models, ICON achieves SOTA performance on the NLG and temporal metrics. As shown in Table 1, our model demonstrates significant improvements on the MIMIC-ABN dataset and achieves competitive performance on the MIMIC-CXR dataset. Additionally, we provide experimental results on the IU X-RAY dataset as a reference in Table 2. Regarding temporal modeling, ICON exhibits significant improvements over other baselines in terms of BLEU score, clinical accuracy, and TEM score while maintaining competitive performance on ROUGE, indicating its enhanced capacity to utilize historical records effectively.

Clinical Efficacy Results. In the right section of Table 1, we observe that ICON achieves SOTA clinical efficacy, increasing the macro CE F_1 score from 0.393 to 0.464 on the MIMIC-CXR dataset and rising by 5.5% on the MIMIC-ABN dataset. These results indicate that our model is capable of generating accurate radiology reports. Furthermore, Table 2 presents the RadGraph F_1 score on both the IU X-RAY and MIMIC-CXR datasets. Our model achieves competitive performance compared with the non-RL-optimized baselines. We also provide per-observation CE results in Table 8, example-based CE results in Table 9, and the performance of ZOOMER in Table 10 for reference.

Ablation Results. The ablation results for MIMIC-ABN and MIMIC-CXR are listed in Table 3 and Table 4. We study three variants: (1) w/o ZOOM, where all components are removed, (2) w/o IN-SPECT, where both the INSPECTOR and MIXUP are removed, and (3) w/o MIXUP, where only MIXUP is removed. The performance of the variant w/o ZOOM drops significantly for both datasets, while the variant w/o INSPECT achieves competitive results in terms of CE scores. This suggests that the ZOOMER effectively extracts lesions and provides relevant abnormal information for report generation. In addition, the variant w/o MIXUP further improves performance, demonstrating the effectiveness of the INSPECTOR in transforming concise lesion information into precise reports. Moreover, introducing lesion-aware mixup augmentation strengthens the consistency of generated outputs, indicating the overall effectiveness of ICON.

5.2 Qualitative Analysis

Case Study. Figure 4 showcases two semantically equivalent cases, i.e., Case A and Case B, extracted from the test set of MIMIC-CXR. In both instances, ICON successfully identifies abnormal observations (e.g., *Cardiomegaly*, *Pleural Effusion*, and *Atelectasis*) and generates consistent phrases including "*pulmonary vascular congestion*", "*bilateral pleural effusions*", and "*compressive atelectasis*." Conversely, the variant *w/o* ZOOM fails to produce these descriptions in Case A. This demonstrates that ZOOMER plays a crucial role in identifying lesions and highlights the ability of the mixup augmentation to ensure the alignment of lesions with their corresponding attributes.

Error Analysis. Figure 5 presents an error case produced by ICON. Although ZOOMER successfully identifies *Pneumonia* in the given radiographs, the GENERATOR fails to realize it into descriptions like "*multifocal pneumonia*" (i.e., a false negative



Figure 4: A case study of ICON on two semantically equivalent cases (i.e., Case A and Case B), given their radiographs and lesions. Spans with the same color (*Cardiomegaly*, *Pleural Effusion*, *Atelectasis*, and *Edema*) represent the same positive observation. Consistent and accurate outputs are highlighted with <u>underline</u>.



Figure 5: An error case produced by ICON. The span and the span denote false negative and false positive observations, respectively.

observation). We notice that the region of this observation is inaccurately identified. Additionally, ZOOMER outputs a false positive observation *Lung Opacity*, leading to an inaccurate phrase "*increased opacity*". To mitigate this issue, a better ZOOMER trained with larger datasets could be beneficial.

6 Related Works

Radiology report generation (Jing et al., 2018; Li et al., 2018; Jing et al., 2019) has gained much attention. Prior research has either devised various memory mechanisms to record key information (Chen et al., 2020, 2021; Qin and Song, 2022; Wang et al., 2023c; Zhao et al., 2023) or proposed different learning methods to enhance performance (Liu et al., 2021c,a,b). In addition, Yang et al. (2021); Li et al. (2023b); Huang et al. (2023); Yan et al. (2023) proposed utilizing knowledge graphs

for report generation. Liu et al. (2019); Lovelace and Mortazavi (2020); Miura et al. (2021); Nishino et al. (2022); Delbrouck et al. (2022) designed various rewards for reinforcement learning to improve clinical accuracy. Tanida et al. (2023) proposed an explainable framework for report generation. Hou et al. (2023b) introduced observations to improve factual accuracy. Kale et al. (2023) proposed a template-based approach to improve the quality and accuracy of radiology reports. Additionally, Ramesh et al. (2022); Bannur et al. (2023); Hou et al. (2023a); Dalla Serra et al. (2023) focused on exploring the temporal structure. Wang et al. (2023b,a) utilized CLIP (Radford et al., 2021) to bridge the modality gap. Mixup is also closely related to this research (Zhang et al., 2018), and this method has been extensively adopted in NLP research (Sun et al., 2020; Yoon et al., 2021; Yang et al., 2022). Although consistency has been studied in many domains (Thimm, 2013; Ribeiro et al., 2019; Camburu et al., 2019; Elazar et al., 2021), it remains unexplored in medical report generation.

7 Conclusion and Future Works

In this paper, we propose ICON, comprising three components to improve both accuracy and interreport consistency. ICON first extracts lesions and then matches fine-grained attributes for report generation. A lesion-aware mixup method is devised for attribute alignment. Experimental results on three datasets demonstrate the effectiveness of ICON. In the future, we plan to explore incorporating large language models (LLMs) into our framework, given their advanced capabilities in planning and generation, to further enhance the performance of radiology report generation. Leveraging the strengths of LLMs could provide more refined signals to enhance the performance of ICON.

Limitations

Although ICON can improve the consistency of radiology report generation, it still exhibits some limitations. Since our lesion extraction method is based on image labels, training such a model requires annotations for images. However, obtaining these annotations can be challenging in some medical settings. Recent advances in foundation vision models (Kirillov et al., 2023) and open-set learning (Zara et al., 2023) could be a potential direction to address this issue. Additionally, image labels are coarse-grained, so the overall accuracy is likely to be lower than when using fine-grained labels (e.g., bounding boxes). Moreover, since our framework consists of two stages, prediction errors can propagate through the pipeline, making the final performance of our framework largely dependent on Stage 1. Reinforcement learning (Nishino et al., 2022) that takes factual improvement as a reward could be a solution to optimize the framework in an end-to-end manner.

Ethics Statement

The IU X-RAY (Demner-Fushman et al., 2016), MIMIC-ABN (Ni et al., 2020), and MIMIC-CXR (Johnson et al., 2019) datasets are publicly available and have been automatically de-identified to protect patient privacy. Our goal is to enhance the inter-report consistency of radiology report generation systems. Despite the substantial improvement of our framework over SOTA baselines, the performance still lags behind the requirements for real-world deployment and could lead to unexpected failures in untested environments. Thus, we urge readers of this paper and potential users of this system to cautiously check the generated outputs and seek expert advice when using it.

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A Appendix

A.1 Abnormal Observation Statistics

The abnormal observation statistics of MIMIC-ABN, MIMIC-CXR, and IU X-RAY are listed in Table 6.

| #Observation | MIMIC-ABN | MIMIC-CXR | IU X-RAY |
|------------------|----------------|------------------|-------------|
| No Finding | 5002/32/22 | 64,677/514/229 | 744/108/318 |
| Cardiomegaly | 16,312/118/244 | 70,561/514/1,602 | 244/38/61 |
| Pleural Effusion | 10,502/80/186 | 56,972/477/1,379 | 60/13/15 |
| Pneumothorax | 1,452/24/4 | 8,707/62/106 | 9/2/5 |
| Enlarged Card. | 5,202/40/90 | 49,806/413/1,140 | 159/29/28 |
| Consolidation | 4,104/36/96 | 14,449/119/384 | 17/1/3 |
| Lung Opacity | 22,598/166/356 | 67,714/497/1,448 | 295/35/57 |
| Fracture | 4,458/32/76 | 11,070/59/232 | 84/6/15 |
| Lung Lesion | 5,612/54/112 | 11,717/123/300 | 85/14/17 |
| Edema | 8,704/76/168 | 33,034/257/899 | 28/2/7 |
| Atelectasis | 19,132/134/220 | 68,273/515/1,210 | 143/15/37 |
| Support Devices | 9,886/58/196 | 60,455/450/1,358 | 89/20/16 |
| Pneumonia | 17,826/138/260 | 23,945/184/503 | 20/2/1 |
| Pleural Other | 2,850/30/62 | 7,296/70/184 | 32/4/7 |

Table 6: Observation distribution in train/valid/test split of three datasets. *Enlarged Card.* refers to *Enlarged Cardiomediastinum*.

A.2 Attributes of Observations

We list top-5 attributes for each observation for a better understanding in Table 7.

A.3 Other Preprocessing Details

We adopt the same preprocessing setup used in Chen et al. (2020), and the minimum count of each token is set to 3/3/10 for IU X-RAY/MIMIC-ABN/MIMIC-CXR, respectively. Other tokens are replaced with a special token <unk>.

| Observation | Top-5 Attributes |
|------------------|--|
| Cardiomegaly | cardiomegaly, borderline, moderately, severely, mildly |
| Pleural Effusion | layering, subpulmonic, thoracentesis, trace, small |
| Pneumothorax | hydropneumothorax, apical, tiny, tension, component |
| Enlarged Card. | mediastinum, widening, contour, widened, lymphadenopathy |
| Consolidation | consolidative, collapse, underlying, developing, consolidations |
| Lung Opacity | opacification, opacifications, patchy, heterogeneous, scarring |
| Fracture | healed, fractured, healing, nondisplaced, posterolateral |
| Lung Lesion | nodular, nodule, mass, nodules, mm |
| Edema | indistinctness, asymmetrical, haziness, asymmetric, interstitial |
| Atelectasis | atelectatic, atelectasis, collapsed, subsegmental, collapse |
| Support Devices | sidehole, carina, coiled, tunneled, duodenum |
| Pneumonia | infectious, infection, atypical, supervening, developing |
| Pleural Other | fibrosis, thickening, biapical, blunting, scarring |

Table 7: Top-5 attributes for each observation.

A.4 Additional Implementation Details

For Stage 1, all three datasets use the same hyperparameters for training ZOOMER, with a learning rate of 1e - 4, batch size of 128, and dropout rate of 0.1, and the number of training epochs is adjusted accordingly. We train ZOOMER for 5, 10, and 15 epochs on MIMIC-CXR, MIMIC-ABN, and IU X-RAY, respectively. During training, several data augmentation methods are applied. The input resolution of Swin Transformer is 256×256 , and we first resize an image to 288×288 , and then randomly crop it to 256×256 with random horizontal flip. All experiments are conducted using one NVIDIA-3090 GTX GPU. For Stage 2, no data augmentation is applied, and we conduct experiments on MIMIC-ABN and IU X-RAY using two NVIDIA-3090 GTX GPUs, and on MIMIC-CXR using four NVIDIA-V100 GPUs, both with half precision. Our model has 328.38M trainable parameters, and the implementations are based on the HuggingFace's Transformers (Wolf et al., 2020). Here are the pretrained models we used:

- Small version of Swin Transformer V2: https://huggingface.co/microsoft/ swinv2-small-patch4-window8-256
- Tiny version of Swin Transformer V2: https://huggingface.co/microsoft/ swinv2-tiny-patch4-window8-256
- Base Version of Biomedical BART: https://huggingface.co/GanjinZero/ biobart-v2-base

A.5 Lesion Extraction

There are two steps in extraction lesions: candidate generation and candidate classification. Given an image with a resolution of 1024×1024 , padding if needed, we apply a sliding window of 384×384 , with a step size of 128 to extract candidates for classification. This operation results in 36 regions. Then, each region is fed into the ZOOMER for class

| Observation | Image | Classifi | cation | Report Classification | | | |
|------------------|-------|----------|----------------|-----------------------|-------|----------------|--|
| Observation | Р | R | \mathbf{F}_1 | Р | R | \mathbf{F}_1 | |
| Enlarged Card. | 0.426 | 0.540 | 0.476 | 0.442 | 0.525 | 0.428 | |
| Cardiomegaly | 0.635 | 0.838 | 0.722 | 0.630 | 0.822 | 0.714 | |
| Lung Opacity | 0.535 | 0.725 | 0.616 | 0.542 | 0.563 | 0.552 | |
| Lung Lesion | 0.318 | 0.187 | 0.235 | 0.321 | 0.177 | 0.228 | |
| Edema | 0.471 | 0.851 | 0.607 | 0.464 | 0.784 | 0.583 | |
| Consolidation | 0.283 | 0.227 | 0.251 | 0.275 | 0.162 | 0.204 | |
| Pneumonia | 0.367 | 0.396 | 0.381 | 0.341 | 0.350 | 0.345 | |
| Atelectasis | 0.541 | 0.660 | 0.595 | 0.539 | 0.620 | 0.577 | |
| Pneumothorax | 0.392 | 0.481 | 0.432 | 0.400 | 0.444 | 0.421 | |
| Pleural Effusion | 0.719 | 0.842 | 0.776 | 0.721 | 0.827 | 0.770 | |
| Pleural Other | 0.289 | 0.440 | 0.349 | 0.295 | 0.315 | 0.304 | |
| Fracture | 0.266 | 0.198 | 0.227 | 0.225 | 0.164 | 0.190 | |
| Support Devices | 0.747 | 0.850 | 0.795 | 0.785 | 0.784 | 0.785 | |
| No Finding | 0.366 | 0.459 | 0.407 | 0.263 | 0.535 | 0.352 | |
| Macro Average | 0.454 | 0.550 | 0.491 | 0.445 | 0.505 | 0.464 | |

Table 8: Experimental results of each observation onthe MIMIC-CXR dataset.

| Madal | M | MIC-A | BN | MIMIC-CXR | | | |
|------------------|-------|-------|----------------|-----------|-------|----------------|--|
| Model | Р | R | \mathbf{F}_1 | Р | R | \mathbf{F}_1 | |
| R2GEN | 0.340 | 0.413 | 0.348 | 0.390 | 0.336 | 0.337 | |
| R2GENCMN | 0.360 | 0.363 | 0.336 | 0.358 | 0.276 | 0.290 | |
| RGRG | - | _ | _ | 0.461 | 0.475 | 0.447 | |
| ORGAN | 0.418 | 0.471 | 0.412 | 0.493 | 0.560 | 0.493 | |
| RECAP | 0.366 | 0.468 | 0.382 | 0.447 | 0.558 | 0.464 | |
| ICON | 0.512 | 0.428 | 0.436 | 0.513 | 0.597 | 0.522 | |
| ICON w/o ZOOM | 0.397 | 0.406 | 0.372 | 0.440 | 0.362 | 0.373 | |
| ICON w/o INSPECT | 0.430 | 0.479 | 0.424 | 0.506 | 0.553 | 0.500 | |
| ICON w/o MIX-UP | 0.433 | 0.509 | 0.438 | 0.507 | 0.590 | 0.517 | |

Table 9: Example-based CE results on the MIMIC-ABN and MIMIC-CXR datasets.

sification, and only the top-1 region is selected for each observation. Note that before extracting lesions, each input case is first assigned with their observations by ZOOMER, and as a result, the number of lesions corresponds to the number of observations.

The *No Finding* observation is excluded for lesion extraction, as it estimates the overall conditions of a patient, which makes it difficult to locate at specific regions.

A.6 Justifications for Additional Data Processing

Justification for Using Historical Records. As stated in Bannur et al. (2023); Hou et al. (2023a), without historical information, it is unreasonable to generate reports with comparisons between two consecutive visits and will lead to hallucinations (Ramesh et al., 2022). As a result, we include historical records as context information for report generation.

Justification for Using All Views. Prior research (Chen et al., 2020, 2021; Hou et al., 2023b,a) treated different views of radiographs in one visit as different samples. However, this is unreasonable

| Dataset | P | R | \mathbf{F}_1 |
|-----------|-------|-------|----------------|
| IU X-ray | 0.223 | 0.243 | 0.225 |
| MIMIC-ABN | 0.379 | 0.472 | 0.411 |
| MIMIC-CXR | 0.454 | 0.550 | 0.491 |

Table 10: Abnormal observation prediction results of ZOOMER at Stage 1. Results on the IU X-RAY dataset are only provided for reference.

to generate a report with only one view position, since different diseases could be observed from different view positions. For example, most of the devices can not be observed from a Lateral view. Given a lateral view radiograph, writing a sentence of "*A right chest tube is in unchanged position*." is unreasonable.

In addition, some reports describe how many views are provided at the beginning, e.g., "PA and lateral views are provided." Above all, we have justified reasons to use all the views in one visit of a patient to generate the target report. Note that previous work treated each image as a sample and their settings have more samples than ours. For a fair comparison, each generated output of a study with L images is duplicated L times so that the number of samples in evaluation is consistent with previous research.