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# Hereditary spastic paraplegia and extensive leukoencephalopathy: a case report of a unique phenotype associated with a *GJB1*/Cx32 p.Pro174Ser variant

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# **Abstract**

**Background** Pathogenic variants in Gap junction protein beta 1 (*GJB1*), which encodes Connexin 32, are known to cause X-linked Charcot-Marie-Tooth disease (CMTX), the second most common form of CMT. CMTX presents with the following fve central nervous systems (CNS) phenotypes: subclinical electrophysiological abnormalities, mild fxed abnormalities on neurological examination and/or imaging, transient CNS dysfunction, cognitive impairment, and persistent CNS manifestations.

**Case presentation** A 40-year-old Japanese male showed CNS symptoms, including nystagmus, prominent spastic paraplegia, and mild cerebellar ataxia, accompanied by subclinical peripheral neuropathy. Brain magnetic resonance imaging revealed hyperintensities in difusion-weighted images of the white matter, particularly along the pyramidal tract, which had persisted since childhood. Nerve conduction assessment showed a mild decrease in motor conduction velocity, and auditory brainstem responses beyond wave II were absent. Peripheral and central conduction times in somatosensory evoked potentials elicited by stimulation of the median nerve were prolonged. Genetic analysis identifed a hemizygous *GJB1* variant, NM\_000166.6:c.520C>T p.Pro174Ser.

**Conclusions** The patient in the case described here, with a *GJB1* p.Pro174Ser variant, presented with a unique CNS-dominant phenotype, characterized by spastic paraplegia and persistent extensive leukoencephalopathy, rather than CMTX. Similar phenotypes have also been observed in patients with *GJC2* and *CLCN2* variants, likely because of the common function of these genes in regulating ion and water balance, which is essential for maintaining white matter function. CMTX should be considered within the spectrum of *GJB1*-related disorders, which can include patients with predominant CNS symptoms, some of which can potentially be classifed as a new type of spastic paraplegia.

**Keywords** X-linked Charcot-Marie-Tooth disease, *GJB1*, Spastic paraplegia, Leukoencephalopathy

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# **Background**

Pathogenic variants in Gap junction protein, beta 1 (*GJB1*), a gap junction family gene located at Xq13.1 cause X-linked Charcot-Marie-Tooth disease (CMTX) [[1\]](#page-6-0). Connexin 32 (Cx32), encoded by *GJB1*, is expressed in the myelinating Schwann cells of peripheral nerves, which are primarily afected in CMTX. Cx32 is also widely localized in outer oligodendrocyte membranes in the central nervous system (CNS) [\[2](#page-6-1)]. Refecting this, five CNS phenotypes are recognized in CMTX  $[3]$  $[3]$ : (1) subclinical abnormalities of visual- and auditory-evoked responses, (2) overt mild fxed abnormalities on neurological examination and/or CNS imaging that may or may not be accompanied by clinical manifestations, (3) severe transient CNS dysfunction accompanied by white matter changes observed by magnetic resonance imaging (MRI), (4) mild to severe cognitive impairment, and (5) persistent central nervous manifestations. The third phenotype, of severe, transient CNS symptoms, such as aphasia, dysarthria, ataxia, monoparesis, hemiparesis, paraparesis, or tetraparesis [[2,](#page-6-1) [4\]](#page-6-3), lasting from hours to weeks, is particularly well documented. The second and ffth phenotypes include persistent symptoms that are often associated with mild, persistent abnormalities on CNS imaging [[5–](#page-6-4)[7\]](#page-6-5).

Here, we report a Japanese patient with the *GJB1* variant, NM\_000166.6:c.520C>T p.Pro174Ser, characterized by spastic paraplegia associated with persistent and extensive leukoencephalopathy involving the pyramidal tracts. Clinically, this patient could potentially be diagnosed with hereditary spastic paraplegia rather than CMTX. Notably, this case features long-term MRI follow-up over 16 years and the frst electrophysiological results associated with this *GJB1* variant. We also present positive MRI and electrophysiological fndings in a female carrier of this variant. The phenotype observed in this patient is similar to those seen in patients with variants of *GJC2* and *CLCN2*, suggesting that these genes and *GJB1* contribute to a common phenotype through their role in regulating ion and water homeostasis in the brain [[8\]](#page-6-6).

## **Case presentation**

The patient was a 40-year-old male (Fig.  $1A$  $1A$ ). His birth was unremarkable. He began rolling over at 4 months and was able to crawl by 8 months. By age 2, he had not yet started walking, and during a health check-up at that age, a pediatrician noted delayed motor development and gaze-evoked horizontal nystagmus. He experienced several febrile convulsions starting at age 1 and began anti-epileptic medications at age 4. At that time, a pediatric neurologist observed nystagmus, ataxia, and lower limb spasticity. Although he was able to walk unaided during his teenage years, his trunk balance remained unstable. His motor function progressively worsened, leading him to use a walking stick by age 28 and a wheelchair for outdoor activities. MRI revealed increased T2 signal in the white matter. At age 30, anti-epileptic medications were discontinued after 26 years of remission. At age 40, during a clinical examination at our hospital, the patient exhibited slight impairments in retrograde memory, visuospatial ability, motor programming, sensitivity to interference, and word retrieval, as indicated by his scores on several cognitive assessments: Montreal Cognitive Assessment  $[9]$  $[9]$  $[9]$  (21/30, with specific deficits in visuospatial ability: -3, attention: -2, language: -2. abstraction: -1, and retrograde memory: -1), Revised Hasegawa Dementia Scale [[10](#page-6-8)] (30/30), Addenbrooke's Cognitive Examination-Revised [[11](#page-6-9)] [96/100, with specific deficits in retrograde memory: -2 and visuospatial ability (clock drawing): -2], and Frontal Assessment Battery [[12\]](#page-6-10) (16/18, with specific deficits in motor programming: -1 and sensitivity to interference: -1). He displayed impaired smooth pursuit eye movement and gaze-evoked horizontal nystagmus. Muscle tone in the lower extremities showed marked spasticity. Although there was pes equinovarus in the lower extremities, muscle atrophy was minimal or very mild (Fig. [1](#page-2-0)B). Manual muscle test scores (right/ left) revealed a pyramidal pattern of weakness in the lower limbs: iliopsoas 4-/3+, quadriceps 5-/5-, hamstrings 3/3, tibialis anterior 2/2, gastrocnemius 2/2, toe extension 1/1, and toe flexion 4/4. Muscle strength in the upper limbs remained preserved. Tendon refexes in the lower limbs were markedly increased, with positive Babinski sign and Chaddock pathological refexes, while those in the upper limbs were normal. No abnormalities were noted in the sensory nervous system, including touch, pain, vibration, and position senses. The finger-to-nose and heel-to-knee test demonstrated mild cerebellar ataxia. Laboratory tests showed normal levels of creatinine kinase, lactate, pyruvic acid, and thyroid hormones, along with a negative result for anti-HTLV1 antibody. Anti-HIV antibody, fuoride, and very long-chain fatty acid levels were not tested. Cerebral spinal fuid analysis revealed a slightly elevated protein level (57 mg/dl) without pleocytosis. Brain MRI by T2 fuid-attenuated inversion recovery and difusion-weighted imaging showed difuse hyperintensity in the white matter, particularly along the pyramidal tract from the posterior limb of the internal capsule to the cerebral peduncle, with additional involvement in the occipital lobe and cerebellar peduncles, medial lemniscus, and corpus callosum (Fig. [1D](#page-2-0)). Mildly low apparent diffusion coefficient values were present in the same lesion (Fig. [1](#page-2-0)D), indicating myelin



<span id="page-2-0"></span>**Fig. 1 A** Family pedigree of the patients in the present case. **B** Pes equinovarus and minimal atrophy of the lower limbs. **C** Sanger sequencing of the *GJB1*:(c.520C>T, p.Pro174Ser) variant in family members. Arrows indicate the position of the variant. **D** Apparent difusion coefcient (ADC) map and difusion-weighted images (DWI) of the brain of the paatient acquired at age 40. **E** Temporal change of T2-weighted imaging (T2WI) in the patient from age 24 to 40

microvacuolation instead of hypomyelination [[13\]](#page-6-11). These signal abnormalities have remained stable from age 24 to 40 (Fig. [1](#page-2-0)E). Nerve conduction assessment indicated a moderate decrease in sensory nerve action potentials of the median, ulnar, and sural nerves, and a mild decrease in motor conduction velocities of the median, ulnar and tibial nerves (36–44 m/sec), indicating subclinical polyneuropathy (Table [1](#page-3-0)A). Electroencephalography yielded normal results, while brainstem auditory-evoked responses beyond wave II were absent. Prolonged peripheral and central conduction times were observed in somatosensory evoked potentials elicited by left median nerve stimulation (N9o-P13/14o latency: 6.4 ms; P13/14o-N20o latency: 9.4 ms).

Given the patient's predominant symptoms of spastic paraplegia, early onset, and gradual progression, exome sequencing was performed for both the patient and his parents. This analysis identified a hemizygous missense variant in *GJB1* (NM\_000166.6:c.520C>T p.Pro174Ser) in the patient and heterozygosity for this variant in his mother, which were confrmed by Sanger sequencing (Fig. [1](#page-2-0)C). In accordance with the guidelines from the



## <span id="page-3-0"></span>**Table 1** Nerve conduction studies of the patient (A) and the mother (B)

Nerve conduction studies were performed on the left side median, ulnar, tibial, fbular, and sural nerves

American College of Medical Genetics and Genomics [[14\]](#page-6-12), this variant was classified as pathogenic  $(PS1+PM2)$ +PP1+PP2+PP3+PP4).

His 73-year-old mother (Fig. [1](#page-2-0)A) also presented with subclinical neuropathy, as evidenced by a mild decrease in nerve conduction velocity, and mild abnormal intensity in corticospinal tract difusion-weighted images (Fig. [2](#page-4-0)A, Table [1](#page-3-0)B). She exhibited severe sensory aphasia with semantic jargon attributable to atrophy and decreased blood flow in the left temporal lobe, extending from the temporal pole to the temporoparietal region (Fig. [2](#page-4-0)B). On the Revised Hasegawa Dementia Scale, she was unable to understand the meaning of tasks and scored only 1/30, comprehending only her age. No known variants associated with semantic dementia were identifed in the mother by exome sequencing.

# **Discussion and conclusions**

The patient in our case, who carries a *GJB1* variant that is primarily recognized as a causative gene for CMTX, is notable for the predominance of progressive spastic paraplegia, minimal peripheral nervous system (PNS) involvement, and the presence of persistent extensive white matter abnormalities, including those affecting the corticospinal tract. Clinically, hereditary spastic paraplegia was strongly suspected over CMT, while radiologically, leukoencephalopathy was a notable consideration. Indeed, a previously reported case with the



<span id="page-4-0"></span>**Fig. 2 A** difusion-weighted images (DWI) of the patient's mother's brain. **B** 123I-N-isopropyl-p-iodoamphetamine single-photon emission computed tomography (SPECT) imaging of the mother. The Z-score maps displayed on an anatomically standardized MRI template are shown

*GJB1* p.Pro174Ser variant was described within the context of leukoencephalopathy, not CMT  $[8]$  $[8]$  $[8]$ . Their clinical features closely mirror those observed in our case (Table  $2$ ). The p.Pro174Ser variant may therefore represent a distinct phenotype characterized by spastic paraplegia and persistent extensive white matter abnormalities involving the pyramidal tract and middle cerebellar peduncles. The 16-year MRI follow-up of the patient provided clear evidence of the persistence of white matter abnormalities. Additionally, while a previous study had not observed MRI abnormalities in female

carriers of this gene variant  $[8]$  $[8]$ , the mother of the present patient exhibited notable fndings, including mild MRI changes in the pyramidal tract and reduced nerve conduction velocities. The differences in phenotypes among female carriers may be partially explained by the biased pattern of X-chromosome inactivation in individual myelinating glial cells [[15](#page-6-13)].

The reason for only certain *GJB1* variants resulting in persistent CNS symptoms that signifcantly deviate from the typical CMT phenotype remain unclear. An analysis of structural domains in Cx32 variants

<span id="page-4-1"></span>



indicates a genotype–phenotype correlation in CMTX, with variants in the intracellular cytoplasmic domain showing less severe phenotypes compared with variants in other domains [[16](#page-6-14)]. p.Pro174Ser is located in the second transmembrane domain, but variants associated with chronic corticospinal tract dysfunction, such as p.Ala39Val [[17](#page-6-15)], p.Thr55Ile [\[18](#page-6-16)], p.Met93Val [[19](#page-6-17)], p.Arg164Gln [\[18](#page-6-16)], p.Arg183His [\[20](#page-6-18)], p.Thr191 frameshift  $[21]$  $[21]$ , and p.Leu143Pro  $[22]$  $[22]$ , are not situated in the intracellular cytoplasmic domain and do not cluster in any specifc domain.

The spastic paraplegia and white matter abnormalities seen in our patient are primary characteristics of several disorders, making diferential diagnosis crucial [[23](#page-6-21)]. One such disorder is caused by certain variants of *GJC2*, which encodes Cx47, a gap junction protein family member primarily expressed in oligodendrocytes  $[24]$  $[24]$  $[24]$ . The symptoms and imaging findings associated with this variant are similar to those observed in our case [[24\]](#page-6-22). While most pathogenic *GJC2* variants lead to Pelizaeus-Merzbacher-like disease type 1 (PMLD1) or hypomyelinating leukodystrophy 2 (HLD2) [[25\]](#page-6-23), a rare type characterized by prominent spastic paraplegia is classifed as autosomal recessive spastic paraplegia type 44 (SPG44) [\[26](#page-6-24)]. Despite the presence of white matter abnormalities, severe cognitive impairment is rare in SPG44 [\[24\]](#page-6-22), and was not observed in a patient with the Cx32 p.Pro174Ser variant  $[8]$ . Similarly, cognitive function was generally preserved in our patient. The mother of our patient showed severe cognitive decline consistent with semantic dementia, yet her white matter abnormalities were milder than those observed in the patient, indicating no relationship with the *GJB1* p.Pro174Ser variant.

Another example is that loss-of-function *CLCN2C* variants cause similar phenotypes to those observed in our patient [\[8](#page-6-6)], suggesting that *GJB1*, *GJC2*, and *CLCN2C* may have related functions. The white matter of the brain is primarily composed of axons with myelin sheaths, and its most crucial physiological function, impulse conduction, depends on the movement of ions and water. A loss-of-function variant of *CLCN2*, which encodes the ClC-2 chloride channel involved in ion and water homeostasis in the brain, can cause leukoencephalopathy  $[8]$  $[8]$ . Similarly, Cx32 forms channels between opposing membranes of adjacent cells to create gap junctions between axons and myelinating Schwann cells or oligodendrocytes, which facilitate the movement of small molecules and ions between myelin and axons [[19\]](#page-6-17). Given the channel function of Cx32 and CIC-2, it is reasonable to assume that loss of channel function leads to demyelination, resulting in a common CNS phenotype of spastic paralysis and white matter abnormalities. Similarly, abnormalities in Cx47, which, like Cx32, belongs to the connexin family are thought to produce comparable efects.

By contrast, patients with total *GJB1* deletion show typical CMT1-like symptoms without CNS involvement [[27\]](#page-6-25). In this scenario, the impact of Cx32 loss of function is limited to the PNS, while compensatory mechanisms by other oligodendrocyte gap junction proteins, including Cx47, may operate in the CNS [[27](#page-6-25)]. Considering this, the Cx32 p.Pro174Ser variant is speculated to act in a dominant-negative manner in the CNS to reduce the function of not only Cx32 but all oligodendrocyte gap junction proteins. Conversely, in the PNS, the loss-offunction efect of this variant on Cx32 is estimated to be relatively mild.

Furthermore, although the genetics and pathomechanisms difer, X-linked adrenoleukodystrophy also shows high signal intensity along white matter tracts on MRI and causes demyelination in both the CNS and PNS [[28](#page-7-0), [29\]](#page-7-1). Therefore, in hereditary diseases that present with demyelinating lesions, such as in the patient case, it is important to focus on both the CNS and PNS during clinical evaluation.

In some patients with *GJB1* variants, including the case presented here, the clinical diagnosis may be hereditary spastic paraplegia or leukoencephalopathy rather than CMT. Therefore, just as PMLD1/HLD2 and SPG44 are regarded as part of the spectrum of *GJC2*-related neurological disorders [\[24](#page-6-22)], CMTX should be recognized as part of the spectrum of *GJB1*-related disorders. This spectrum may include patients with predominant CNS phenotypes, as well as those that could potentially be classifed as a novel type of spastic paraplegia.

#### **Abbreviations**

- CMTX X-linked Charcot-Marie-Tooth disease *GJB1* Gap Junction Protein, Beta 1
- CNS Central nervous systems
- Cx32 Connexin 32
- MRI Magnetic resonance imaging
- PNS Peripheral nervous system
- PMLD1 Pelizaeus-Merzbacher-like disease type 1
- HLD2 Hypomyelinating leukodystrophy 2
- SPG44 Spastic paraplegia type 44

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#### **Authors' contributions**

H.N., H.D. and F.T drafted the manuscript; E.T. and Y.M. performed electrophysical analysis; H.F., T.W., A.F., and N.M. identifed the GJB1 variant; M.T., Y.H., Y.N., K.K., M.H. and H.K. conducted clinical evaluations. The fnal manuscript was read and approved by all authors.

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#### **Data availability**

No datasets were generated or analysed during the current study.

#### **Availability of data and materials**

The DNA sequencing data analyzed during the current study is available in the DNA Data Bank of Japan [Accession number LC822818].

#### **Declarations**

#### **Ethics approval and consent to participate**

Research protocols were approved by the Institutional Review Board of Yokohama City University School of Medicine (A130530002), and written informed consent was obtained from all patients.

#### **Consent for publication**

Written informed consent for publication was obtained from all patients.

#### **Competing interests**

The authors declare no competing interests.

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#### **References**

- <span id="page-6-0"></span>Bergoffen J, Scherer SS, Wang S, Scott MO, Bone LJ, Paul DL, et al. Connexin mutations in X-linked Charcot-Marie-Tooth disease. Science (New York, NY). 1993;262(5142):2039–42. [https://doi.org/10.1126/science.](https://doi.org/10.1126/science.8266101) [8266101](https://doi.org/10.1126/science.8266101).
- <span id="page-6-1"></span>2. Paulson HL, Garbern JY, Hoban TF, Krajewski KM, Lewis RA, Fischbeck KH, et al. Transient central nervous system white matter abnormality in X-linked Charcot-Marie-Tooth disease. Ann Neurol. 2002;52(4):429–34. [https://doi.org/10.1002/ana.10305.](https://doi.org/10.1002/ana.10305)
- <span id="page-6-2"></span>3. Wang Y, Yin F. A Review of X-linked Charcot-Marie-Tooth Disease. J Child Neurol. 2016;31(6):761–72. <https://doi.org/10.1177/0883073815604227>.
- <span id="page-6-3"></span>4. Hanemann CO, Bergmann C, Senderek J, Zerres K, Sperfeld AD. Transient, recurrent, white matter lesions in X-linked Charcot-Marie-Tooth disease with novel connexin 32 mutation. Arch Neurol. 2003;60(4):605–9. [https://](https://doi.org/10.1001/archneur.60.4.605) [doi.org/10.1001/archneur.60.4.605.](https://doi.org/10.1001/archneur.60.4.605)
- <span id="page-6-4"></span>5. Abrams CK, Freidin M. GJB1-associated X-linked Charcot-Marie-Tooth disease, a disorder afecting the central and peripheral nervous systems. Cell Tissue Res. 2015;360(3):659–73. [https://doi.org/10.1007/](https://doi.org/10.1007/s00441-014-2014-6) [s00441-014-2014-6.](https://doi.org/10.1007/s00441-014-2014-6)
- 6. Kassubek J, Bretschneider V, Sperfeld AD. Corticospinal tract MRI hyperintensity in X-linked Charcot-Marie-Tooth Disease. Journal of clinical neuroscience : official journal of the Neurosurgical Society of Australasia. 2005;12(5):588–9. [https://doi.org/10.1016/j.jocn.2004.07.020.](https://doi.org/10.1016/j.jocn.2004.07.020)
- <span id="page-6-5"></span>7. Siskind C, Feely SM, Bernes S, Shy ME, Garbern JY. Persistent CNS dysfunction in a boy with CMT1X. J Neurol Sci. 2009;279(1–2):109–13. [https://doi.](https://doi.org/10.1016/j.jns.2008.12.031) [org/10.1016/j.jns.2008.12.031](https://doi.org/10.1016/j.jns.2008.12.031).
- <span id="page-6-6"></span>8. Depienne C, Bugiani M, Dupuits C, Galanaud D, Touitou V, Postma N, et al. Brain white matter oedema due to ClC-2 chloride channel defciency: an observational analytical study. The Lancet Neurology. 2013;12(7):659–68. [https://doi.org/10.1016/S1474-4422\(13\)70053-X](https://doi.org/10.1016/S1474-4422(13)70053-X).
- <span id="page-6-7"></span>9. Nasreddine ZS, Phillips NA, Bedirian V, Charbonneau S, Whitehead V, Collin I, et al. The Montreal Cognitive Assessment, MoCA: a brief screening tool for mild cognitive impairment. J Am Geriatr Soc. 2005;53(4):695–9. [https://doi.org/10.1111/j.1532-5415.2005.53221.x.](https://doi.org/10.1111/j.1532-5415.2005.53221.x)
- <span id="page-6-8"></span>10. Kim KW, Lee DY, Jhoo JH, Youn JC, Suh YJ, Jun YH, et al. Diagnostic accuracy of mini-mental status examination and revised hasegawa dementia scale for Alzheimer's disease. Dement Geriatr Cogn Disord. 2005;19(5– 6):324–30. <https://doi.org/10.1159/000084558>.
- <span id="page-6-9"></span>11. Larner AJ, Mitchell AJ. A meta-analysis of the accuracy of the Addenbrooke's Cognitive Examination (ACE) and the Addenbrooke's Cognitive Examination-Revised (ACE-R) in the detection of dementia. Int Psychogeriatr. 2014;26(4):555–63.<https://doi.org/10.1017/s1041610213002329>.
- <span id="page-6-10"></span>12. Dubois B, Slachevsky A, Litvan I, Pillon B. The FAB: a Frontal Assessment Battery at bedside. Neurology. 2000;55(11):1621–6. [https://doi.org/10.](https://doi.org/10.1212/wnl.55.11.1621) [1212/wnl.55.11.1621](https://doi.org/10.1212/wnl.55.11.1621).
- <span id="page-6-11"></span>13. Patay Z. Difusion-weighted MR imaging in leukodystrophies. Eur Radiol. 2005;15(11):2284–303. [https://doi.org/10.1007/s00330-005-2846-2.](https://doi.org/10.1007/s00330-005-2846-2)
- <span id="page-6-12"></span>14. Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J, et al. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. 2015;17(5):405– 24. <https://doi.org/10.1038/gim.2015.30>.
- <span id="page-6-13"></span>15. Siskind CE, Murphy SM, Ovens R, Polke J, Reilly MM, Shy ME. Phenotype expression in women with CMT1X. Journal of the peripheral nervous system : JPNS. 2011;16(2):102–7. [https://doi.org/10.1111/j.1529-8027.](https://doi.org/10.1111/j.1529-8027.2011.00332.x) [2011.00332.x](https://doi.org/10.1111/j.1529-8027.2011.00332.x).
- <span id="page-6-14"></span>16. Record CJ, Skorupinska M, Laura M, Rossor AM, Pareyson D, Pisciotta C, et al. Genetic analysis and natural history of Charcot-Marie-Tooth disease CMTX1 due to GJB1 variants. Brain : a journal of neurology. 2023;146(10):4336–49. [https://doi.org/10.1093/brain/awad187.](https://doi.org/10.1093/brain/awad187)
- <span id="page-6-15"></span>17. Marques W Jr, Sweeney JG, Wood NW, Wroe SJ, Marques W. Central nervous system involvement in a novel connexin 32 mutation afecting identical twins. J Neurol Neurosurg Psychiatry. 1999;66(6):803–4. [https://](https://doi.org/10.1136/jnnp.66.6.803) [doi.org/10.1136/jnnp.66.6.803.](https://doi.org/10.1136/jnnp.66.6.803)
- <span id="page-6-16"></span>18. Panas M, Karadimas C, Avramopoulos D, Vassilopoulos D. Central nervous system involvement in four patients with Charcot-Marie-Tooth disease with connexin 32 extracellular mutations. J Neurol Neurosurg Psychiatry. 1998;65(6):947–8.<https://doi.org/10.1136/jnnp.65.6.947a>.
- <span id="page-6-17"></span>19. Kleopa KA, Abrams CK, Scherer SS. How do mutations in GJB1 cause X-linked Charcot-Marie-Tooth disease? Brain Res. 2012;1487:198–205. <https://doi.org/10.1016/j.brainres.2012.03.068>.
- <span id="page-6-18"></span>20. Bort S, Nelis E, Timmerman V, Sevilla T, Cruz-Martínez A, Martínez F, et al. Mutational analysis of the MPZ, PMP22 and Cx32 genes in patients of Spanish ancestry with Charcot-Marie-Tooth disease and hereditary neuropathy with liability to pressure palsies. Hum Genet. 1997;99(6):746–54. <https://doi.org/10.1007/s004390050442>.
- <span id="page-6-19"></span>21. Lee MJ, Nelson I, Houlden H, Sweeney MG, Hilton-Jones D, Blake J, et al. Six novel connexin32 (GJB1) mutations in X-linked Charcot-Marie-Tooth disease. J Neurol Neurosurg Psychiatry. 2002;73(3):304–6. [https://doi.org/](https://doi.org/10.1136/jnnp.73.3.304) [10.1136/jnnp.73.3.304](https://doi.org/10.1136/jnnp.73.3.304).
- <span id="page-6-20"></span>22. Kleopa KA, Zamba-Papanicolaou E, Alevra X, Nicolaou P, Georgiou DM, Hadjisavvas A, et al. Phenotypic and cellular expression of two novel connexin32 mutations causing CMT1X. Neurology. 2006;66(3):396–402. <https://doi.org/10.1212/01.wnl.0000196479.93722.59>.
- <span id="page-6-21"></span>23. Pavone P, Falsaperla R, Polizzi A. A Hundred Faces for a Unique Disorder: Hereditary Spastic Paraplegia. J Integr Neurosci. 2024;23(6):115. [https://](https://doi.org/10.31083/j.jin2306115) [doi.org/10.31083/j.jin2306115.](https://doi.org/10.31083/j.jin2306115)
- <span id="page-6-22"></span>24. Ghasemi A, Tavasoli AR, Khojasteh M, Rohani M, Alavi A. Description of Phenotypic Heterogeneity in a GJC2-Related Family and Literature Review. Molecular syndromology. 2023;14(5):405–15. [https://doi.org/10.](https://doi.org/10.1159/000529678) [1159/000529678.](https://doi.org/10.1159/000529678)
- <span id="page-6-23"></span>25. Henneke M, Combes P, Diekmann S, Bertini E, Brockmann K, Burlina AP, et al. GJA12 mutations are a rare cause of Pelizaeus-Merzbacher-like disease. Neurology. 2008;70(10):748–54. [https://doi.org/10.1212/01.wnl.](https://doi.org/10.1212/01.wnl.0000284828.84464.35) [0000284828.84464.35.](https://doi.org/10.1212/01.wnl.0000284828.84464.35)
- <span id="page-6-24"></span>26. Orthmann-Murphy JL, Salsano E, Abrams CK, Bizzi A, Uziel G, Freidin MM, et al. Hereditary spastic paraplegia is a novel phenotype for GJA12/GJC2 mutations. Brain : a journal of neurology. 2009;132(Pt 2):426–38. [https://](https://doi.org/10.1093/brain/awn328) [doi.org/10.1093/brain/awn328](https://doi.org/10.1093/brain/awn328).
- <span id="page-6-25"></span>27. Takashima H, Nakagawa M, Umehara F, Hirata K, Suehara M, Mayumi H, et al. Gap junction protein beta 1 (GJB1) mutations and central nervous

system symptoms in X-linked Charcot-Marie-Tooth disease. Acta Neurol Scand. 2003;107(1):31–7. [https://doi.org/10.1034/j.1600-0404.2003.](https://doi.org/10.1034/j.1600-0404.2003.01317.x) [01317.x.](https://doi.org/10.1034/j.1600-0404.2003.01317.x)

- <span id="page-7-0"></span>28. Kemp S, Berger J, Aubourg P. X-linked adrenoleukodystrophy: clinical, metabolic, genetic and pathophysiological aspects. Biochem Biophys Acta. 2012;1822(9):1465–74. [https://doi.org/10.1016/j.bbadis.2012.03.012.](https://doi.org/10.1016/j.bbadis.2012.03.012)
- <span id="page-7-1"></span>29. Lynch DS, Wade C, Paiva ARBd, John N, Kinsella JA, Merwick A, et al. Practical approach to the diagnosis of adult-onset leukodystrophies: an updated guide in the genomic era. J Neurol Neurosurg Psychiatry. 2019;90(5):543–55. [https://doi.org/10.1136/jnnp-2018-319481.](https://doi.org/10.1136/jnnp-2018-319481)

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