



Syddansk Universitet

Estimating plant stem emerging points (PSEPs) of sugar of beets in early growth stages

Midtiby, Henrik Skov; Mosgaard Giselsson, Thomas; Jørgensen, Rasmus Nyholm

Published in:
Biosystems Engineering

DOI:
[10.1016/j.biosystemseng.2011.10.011](https://doi.org/10.1016/j.biosystemseng.2011.10.011)

Publication date:
2012

Document license
CC BY-NC-ND

Citation for pulished version (APA):
Midtiby, H., Mosgaard Giselsson, T., & Jørgensen, R. N. (2012). Estimating plant stem emerging points (PSEPs) of sugar of beets in early growth stages. *Biosystems Engineering*, 111(1), 83-90. DOI: [10.1016/j.biosystemseng.2011.10.011](https://doi.org/10.1016/j.biosystemseng.2011.10.011)

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from the public portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain
- You may freely distribute the URL identifying the publication in the public portal ?

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Estimating plant stem emerging point of beets in early growth stages

H. S. Midtiby^{a,*}, T. M. Giselsson^a, R. N. Jørgensen^a

^a*Institute of Chemical Engineering, Biotechnology and Environmental Technology,
University of Southern Denmark, Niels Bohrs Allé 1, 5230 M, Denmark*

Abstract

Successful intra-row mechanical weed control of sugar beet (*beta vulgaris*) in early growth stages requires precise knowledge about location of crop plants. A computer vision system for locating Plant Stem Emerging Point (PSEP) of sugar beet in early growth stages was developed and tested. The system is based on detection of individual leaves; each leaf location is then described by centre of mass and petiole location. After leaf detection were the true PSEP locations annotated manually and a multivariate normal distribution model of the PSEP relative to the located leaf was built. From testing the system, PSEP estimates based on a single leaf have an average error of $\sim 3mm$. When several leaves are detected the average error decreases to less than $2mm$.

Keywords: plant center, machine vision, leaf extraction

1. Introduction

Mechanical inter-row weeding between crop rows have been used for a long time. But mechanical intra-row weeding within rows between the single crop plants is relatively new. Physical intra-row methods can in general rely on three different strategies (Griepentrog and Dedouis, 2010): (1) soil coverage of weeds or (2) weed root/stem cutting or (3) uprooting of weeds (whole plant or partly). The first option is only relevant in some crop types like cereals and potatoes. Sugar beet (*beta vulgaris*) at dicotyledon stage does not belong to these groups (Melander, 2000; Kouwenhoven, 1997) and only strategy two and three may be used.

Several intra-row mechanical weed management methods need to know where the crop plants are located especially with concern to the Plant Stem Emerging Point (PSEP) which is defined as the point where the plant stem emerges from the soil surface. Computer vision was used by (Tillett et al., 2008) to locate transplanted cauliflower plants, before a cultivation disc is moved such that the crop plants are not harmed. RTK-GPS have been used to mark the position

*Corresponding author. E-mail address: hemi@kbm.sdu.dk (H. S. Midtiby)

17 of crop seeds during sowing (Griepentrog et al., 2005), but the PSEP is not
18 identical to the planted seeds position, as the orientation of the seed have not
19 been taken into account. (Nørremark et al., 2008) used the RTK GPS coor-
20 dinates to control a cycloid hoe doing intra-row weed control based on seed
21 positions. Uncertainty in seed orientation, PSEP, and GPS accuracy limits the
22 achievable precision to approx $30mm$. (Sun et al., 2010) used RTK-GPS for
23 mapping transplanted tomatoes, 95% of the plants were within $51mm$ from the
24 true plant position. Based on vision input the crop plant positions may be
25 determined with at higher accuracy and precision as (Åstrand and Baerveldt,
26 2002) indicated by guiding an autonomous weed robot with $20mm$ accuracy
27 along crop rows. Earlier work on extraction of individual leaves from images
28 include (Franz et al., 1991) which analysed boundary curvature by comparing
29 with a known leaf shape and (Neto et al., 2006) which detected individual leaves
30 in complex scenes based on Gustafson-Kessel clustering. This paper describes
31 and evaluates a vision based method which detects single crop leaves and predict
32 where the corresponding PSEP is located.

33 2. Materials and methods

34 The current work consists of three parts: (1) development of a leaf detector,
35 (2) building of a relative PSEP model, and (3) using the relative PSEP model
36 to predict true PSEP based on detected leaves. An example image of sugar beet
37 plants in early growth stages is shown in figure 2. The leaves can be described
38 as convex objects with a thin stem (petiole). Leaves are detected by locating
39 convex regions of the plant contour. The relative PSEP model is generated
40 by comparing manually marked PSEP locations (ground truth values) with the
41 detected leaves. Based on the relative PSEP location model and detected leaves,
42 estimates of the true PSEP locations are obtained automatically. Finally are
43 the methods for evaluating performance described.

44 2.1. Image acquisition and segmentation

45 Images from sugar beet fields were acquired by a bi-spectral line scanning
46 camera mounted on the Robovator (Poulsen, 2010) intra-row mechanical weed-
47 ing robot. The setup for image capturing is shown in Fig. 1. The imaged sugar
48 beet plants were part of field emergence trials conducted by Maribo Seed in 2009.
49 Precise plant placement is not required for field emergence trials which can be
50 seen directly in the acquired images where sugar beet plants are distributed
51 randomly over the captured region. The captured area was illuminated with
52 two $55W$ halogen lamps. Each line in the acquired image consists of 256 pixels
53 and a typical data file consist of approximately 13,000 scan lines. A single pixel
54 measured approximately $1.1mm \times 1.1mm$. A sample image can be seen in Fig.
55 2. For each pixel both a red and a near infrared value are available. Com-
56 bining red and near infrared values makes it possible to segment images into
57 plant material and soil which is done by calculating the NDVI value for each
58 pixel (Backes and Jacobi, 2006). After this operation a single channel image is

59 obtained with plant material having a high NDVI value compared to soil. This
60 image is segmented using a threshold of 0.2 to form a binary image, the thresh-
61 old was found by trial and error. These binary images are the basis for the data
62 material used in this paper. Before further analysis are connected components
63 located. It is assumed that a leaf will only contribute to one connected blob.
64 To remove noise only blobs with an area larger than 160 pixels are kept.

65 *2.2. Leaf extraction*

66 For detecting leaves the general leaf structure is exploited. Examples of
67 leaf shapes are shown in Fig. 2. The structure consists of a large mainly
68 convex region attached to the rest of the plant via a thin stem (petiole)(Meier,
69 2001). The leaf extraction method works in two steps. First convex regions are
70 located and marked as leaf tip candidates, this is described in section 2.3. From
71 the located leaf tip candidates a search for the corresponding petiole is then
72 initiated, the search process is described in section 2.4. If a petiole is located
73 a leaf is found. When a leaf is detected the leaf location and orientation is
74 described by petiole location \vec{S} and the leaf centre of mass \vec{C} .

75 *2.3. Leaf tip candidate location*

76 In this section a method for locating leaf tip candidates within the segmented
77 images is described. Leaf tip candidates are found at local curvature minima in
78 curvature of the plant boundary. At this stage is the plant boundary specified as
79 the list of coordinates \vec{z}_k where $k \in [1, \dots, n]$ and the boundary is followed clock
80 wise. The curvature is then defined as the angle between the line connecting
81 point $k - \Delta$ and k and the line connecting point k and $k + \Delta$. The sign of
82 the direction change indicates whether the current location of the boundary is
83 concave or convex. In this paper the parameter $\Delta = 12$ was used together with a
84 running average of the five nearest points. Plant boundary and curvature along
85 the boundary is visualized in Fig. 3. Local maxima corresponds to concave
86 regions, which are often located at leaf intersections or near the sugar beat
87 growth point, which is assumed to be vertically above PSEP where several
88 leaves are connected to a common area. Local minima corresponds to convex
89 regions such as leaf tips.

90 To locate a single leaf tip candidate for each leaf, the following steps are
91 used: (1) division of the boundary into concave and convex regions, (2) locate
92 the minima in each convex region and (3) thresholding of the located minima.
93 The purpose of the first step is to split the boundary into segments that at
94 most contain a single leaf tip. As splitting points are used locations where the
95 curvature changes from positive to negative or from negative to positive values.
96 The second step finds the most likely leaf tip location, which are the points
97 along the boundary where the boundary is convex and change of direction is
98 maximized. Step three removes possible leaf tip locations according to direction
99 change, if the direction change is too small (less than 1 radians) the candidate
100 is eliminated.

101 *2.4. Location of corresponding petiole*

102 From each of the candidate leaf tips a search for the corresponding petiole is
103 then initiated. Two walkers are placed at the leaf tip with the goal of following
104 the boundary in each direction, one clockwise and one counter clockwise. The
105 movement of the walkers is controlled such that they will reach the petiole
106 nearly simultaneous. Each walker is then moved forward until the next step
107 along the boundary will bring the euclidean distance between the walker and
108 the leaf tip point above a specified threshold distance l . Then the distance
109 between the walkers is measured. This process (walker movement and distance
110 measurement) is repeated with increasing values of l . In Fig. 4 the search
111 strategy is visualized. For each value of the distance threshold the corresponding
112 circle is drawn together with the two walker locations.

113 To locate the petiole the distance between the walkers are investigated as
114 follows: (1) search for a narrow leaf region which initiates the region in which
115 the petiole can be located followed by a (2) search for a broadening of the leaf
116 width which ends the region in which the petiole can be found. This strategy
117 was implemented as a state machine. The state machine starts in the leaf-tip
118 state and remains there until the distance between the two walkers get below
119 half of the maximum distance between the walkers and the stage is changed to
120 the leaf-stem stage. While in leaf-stem stage the system keeps track of the min-
121 imum distance between the walkers and corresponding walker locations. When
122 the distance between the walkers exceed three times the minimum distance ob-
123 served in the leaf-stem stage the search is terminated. The leaf boundary cutoff
124 positions are given by the location of the walkers where the distance between the
125 walkers are minimized within the leaf-stage. The petiole location is set to the
126 midpoint of the two boundary cutoff positions. To avoid infinite loops petiole
127 search is terminated if one of the walkers reach a leaf tip candidate or the two
128 walkers pass each other.

129 *2.5. Manual marking of root / leaf relative locations*

130 After the automatic extraction of plant leaves as described in section 2.2,
131 real PSEP location were marked manually. A program showed each plant and
132 the user should then mark the pixel nearest the true PSEP. Fig. 5 illustrate a
133 sample image with PSEPs marked with red spots and detected leaves marked
134 by orange. To describe the marked PSEP location relative to the extracted leaf,
135 the leaf coordinate system is placed with origin located at the petiole \vec{S} and
136 direction of the x axis parallel to the vector $\vec{C} - \vec{S}$. An example is shown in Fig.
137 6.

138 The manual annotation of the location of the true PSEP locations is prone
139 to errors. PSEP locations were marked with a single pixel, so the average quan-
140 tization error will be $\sim 0.5mm$ along each dimension. The true PSEP locations
141 marked by a person will also have an uncertainty. To estimate size of the typical
142 error in this process the same image was annotated by two persons. Differences
143 in PSEP locations were calculated and mean distance between annotations were
144 determined.

145 *2.6. PSEP location model*

146 A multivariate normal distribution is used to model the PSEP location
 147 within the leaf coordinate system. The model is defined as:

$$p(\vec{x}) = \frac{1}{2\pi |\Sigma_{lc}|} \exp \left[-\frac{1}{2} (\vec{x} - \vec{x}_{lc})^T \Sigma_{lc}^{-1} (\vec{x} - \vec{x}_{lc}) \right] \quad (1)$$

148 where \vec{x}_{lc} is the centre of the true PSEP estimate and Σ_{lc} is the covariance
 149 matrix. Both \vec{x}_{lc} and Σ_{lc} are expressed in the leaf coordinate system. Ellipses
 150 are used to visualize the multivariate normal distribution, contours of certain
 151 values are drawn such that a given fraction of the probability is inside the
 152 ellipses. To calculate the ellipses the formula below is used:

$$(\vec{x} - \vec{x}_{lc})^T \Sigma_{lc}^{-1} (\vec{x} - \vec{x}_{lc}) = \chi_{2,\alpha}^2 \quad (2)$$

153 where $\chi_{2,\alpha}^2$ is the χ^2 distribution with 2 degrees of freedom and P value $1 - \alpha$.
 154 Typical fractions used for visualization are 68%, 95% and 99.7%. As the PSEP
 155 is defined relative to the leaf (Fig. 6) the x and y coordinate values translate to
 156 a displacement along the major leaf axis and displacement perpendicular to the
 157 same axis respectively. The PSEP is expected to lie in extension of the primary
 158 leaf axis (low y values) shifted to negative x values. For later analysis position
 159 and uncertainty parameters are converted to the global coordinate system using
 160 a coordinate transformation based on rotation and translation.

161 *2.7. Combination of relative PSEP location models*

162 In many cases it is possible to detect more than a single leaf, an example
 163 is shown in Fig. 7. In the figure 99.7% ellipses of the two estimates of the
 164 true PSEP share a common region and it is expected that the true PSEP is
 165 located within this region. To combine two PSEP models ($p_A(\vec{x})$ and $p_B(\vec{x})$)
 166 the probability densities are multiplied and normalized.

$$p_C(\vec{x}) \propto p_A(\vec{x}) \cdot p_B(\vec{x}) \quad (3)$$

167 If the PSEP models are defined by the parameters $\Sigma_A, \Sigma_B, \vec{x}_c^A$ and \vec{x}_c^B the
 168 parameters of the combined model can be expressed as (Gales and Airey, 2006)

$$\Sigma_C^{-1} = \Sigma_A^{-1} + \Sigma_B^{-1} \quad (4)$$

$$\vec{x}_c^C = \Sigma_C (\Sigma_A^{-1} \vec{x}_c^A + \Sigma_B^{-1} \vec{x}_c^B) \quad (5)$$

169 This combination of PSEP models is based on the same principle as least
 170 squares estimation in the Kalman filter.

171 *2.8. Generation of position predictions*

172 To test the developed method for PSEP estimation, the method was applied
173 to a test image. True plant locations were determined manually and compared
174 to six sets $D_{1,\dots,6}$ of predicted PSEP locations. These sets were used to measure
175 accuracy of the located PSEPs under different conditions, eg. different number
176 of detected leaves per plant.

177 From all the detected leaves were a PSEP generated (using only information
178 from this leaf). This is set D_1 . D_2 contains PSEPs calculated from two detected
179 leaves. All possible combinations were tested and leaf pairs was combined if
180 distance between centers of their PSEP models was less than $20mm$. D_3 and
181 D_4 are similar to D_2 except that 3 and 4 leaves are used for calculating the PSEP.
182 For a plant where n leaves was detected, the set D_k would contain $\binom{k}{n}$ elements
183 related to that plant. Not all plants had all four leaves detected, therefore will
184 D_4 not contain PSEPs associated to these plants so when the number of leaves
185 used to calculate PSEPs is increased, will the precision of the located PSEPs
186 increase, but a larger fraction will be missed. D_5 is a compromise between
187 large coverage and low placement error. The set is built on D_1 by merging
188 PSEP models with a distance between predicted plant centers of $20mm$ or less.
189 This merging scheme will generate combined PSEP models based on position
190 information from up to 4 leaves. In addition were a set, D_6 , generated by
191 manual annotation by a different person than the one who marked the reference
192 PSEPs. D_6 covered only one third of the test image and was used to estimate
193 uncertainty of the manually marked PSEPs.

194 *2.9. Performance evaluation*

195 Performance of the PSEP location model were judged according to the fol-
196 lowing values:

197 **False positives:** If a leaf is falsely found by the leaf separator method it con-
198 stitute a false positive. These cases are characterized by having a long
199 distance from the predicted PSEP to the nearest true PSEP. False pos-
200 itives are detected by setting a threshold on the allowed distance from
201 predicted leaf location to the nearest true PSEP.

202 **Missed PSEP locations:** If none of a plant's leaves have been detected a
203 PSEP is missed. It is characterized by having a long distance from the
204 true PSEP to the nearest predicted PSEP. Missed PSEPs are detected by
205 setting a threshold on the allowed distance.

206 **Predicted position error:** The error in the predicted PSEP location were av-
207 eraged for all predicted PSEP locations with an error less than a threshold
208 of $20mm$.

209 **3. Results**

210 *3.1. Leaf detector performance*

211 For evaluating performance of the leaf detector, the 805 leaves present in
212 the test images were counted manually. The leaf detector located 46.6% (395)
213 leaves, of those 2.4% (19) were false positives.

214 *3.2. Relative PSEP model*

215 The leaf detector were applied to three datasets. True PSEPs were marked
216 by hand in all three datasets. Additionally leaves were detected by the leaf
217 detector method and their location specific information recorded. Analyzing
218 leaves and PSEPs led to the generation of 223 data points. In the local leaf
219 coordinate system the multivariate normal distribution model is described by
220 the parameter values:

$$\vec{x}_{lc} = \begin{pmatrix} 5.40 \\ 0.24 \end{pmatrix} mm \quad \Sigma_{lc} = \begin{pmatrix} 12.65 & 1.28 \\ 1.28 & 2.35 \end{pmatrix} mm^2 \quad (6)$$

221 *3.3. Fraction of PSEP locations found*

222 The fraction of missed PSEPs is visualized as a function of the chosen thresh-
223 old in Fig. 8. All six PSEP prediction methods show the same trend. At first
224 the fraction of missed PSEPs decreases linearly until the curve flattens out. The
225 point where the curve flattens out indicates the maximum error of the position
226 estimate and the fraction of PSEPs that are not found. Note that humans are
227 good at locating a large fraction of the PSEPs. The fraction of roots not found
228 within 20mm are shown in the MR column in Tab 1. If a single leaf (D_1) is
229 used to predict PSEPs approximately 10% of the true PSEPs will be missed,
230 this number increases strongly when the number of leaves used in the prediction
231 is increased. $\sim 37\%$ of the true PSEPs are missed with estimates based on two
232 leaves, this number is increased to $\sim 89\%$ when four leaves are used to generate
233 estimates. This increase in fraction of missed PSEPs is only to be expected, as
234 the plants with one or two detected leaves are not present in D_3 and D_4 .

235 *3.4. Fraction of false positives*

236 To gain insight in the accuracy of PSEP–location–estimates the fraction of
237 false positives is visualized as a function of threshold distance in Fig. 9. The
238 figure is divided into four regions, each representing a dataset. In dataset *One*
239 is the PSEP near which the leaf detector found a single leaf; in *Three* the leaf
240 detector located three leaves. From the green curve it is seen that $\sim 20\%$ of
241 the D_1 position–estimates have a distance (error) of more than 4mm to the
242 nearest true PSEP, for comparison is the corresponding distance for D_2 3mm.
243 The figure shows that when the number of leaves used to generate a PSEP–
244 location–estimate is increased the error in the estimate is reduced significantly.
245 The figure was divided into four underlying data sets such that each dataset
246 could be weighted appropriately. If all the data was shown in one plot it would

247 be difficult to interpret because each set of location estimates was based on a
248 unique dataset. The number of false positives and missed roots for each of the
249 estimate sets is given in table 1. The listed values are found using a threshold
250 distance of $20mm$. In addition the estimate error (distance from estimate to
251 nearest PSEP) is described using the average value and the 95% quantile (95%
252 of the predicted PSEP had an error of less than. . .).

253 4. Discussion

254 The leaf detector is not able to locate all leaves in the test images. This is
255 due to overlapping leaves, leaves with irregular shapes and to a certain extend
256 limitations in the implemented algorithm. Some typical cases are shown in Fig.
257 10. The petiole search is fragile and will fail if more than a single leaf tip
258 candidate is found in one leaf. In the used leaf definition (convex area with a
259 thin petiole) overlapping leaves can influence both criteria: the combined leaf
260 area is not guaranteed to be convex and the petiole region can be hidden or
261 widened. Rarely will the relative location of leaf tip estimate and petiole cause
262 the petiole search strategy to fail, this is the case when distance between petiole
263 and leaf tip estimate is less than the distance between leaf tip estimate and the
264 true leaf tip. To reduce the fraction of missed PSEPs the leaf detector must
265 be improved. If a PSEP is not located none of the associated leaves have been
266 detected.

267 Before evaluation of the implemented algorithms the uncertainty of the true
268 PSEP position should be investigated. This can be achieved by comparing
269 true PSEPs with PSEPs determined by a human being different from the one
270 who determined the true PSEPs initially. The difference between such two
271 manual annotations can be used as an estimate of the position uncertainty
272 of the true PSEPs. On average the difference was $1.37mm$ and in 95% of
273 the cases the difference between the two human annotations were less than
274 $3.58mm$. Two sources contribute to this difference (1) quantification error and
275 (2) uncertainty / unreliability of the human annotation. The quantification
276 error origins from the annotation program, which used integer coordinates for
277 describing PSEPs. A rough estimate of this error is $\pm 0.5mm$ along the two
278 coordinate axes. The human annotation unreliability origins from differences in
279 test image interpretation.

280 When the leaf detector has found two leaves of a single plant the correspond-
281 ing true PSEP will with a probability of 95% be within a distance of $5mm$ or less
282 from the guess. This and similar values are shown in table 1. (Sun et al., 2010)
283 positions 95% of the plants within $51mm$. The accuracy of the vision system
284 is thus one order of magnitude better than RTK-GPS seeding of plants. When
285 three or more leaves are used to predict PSEPs the accuracy is comparable to
286 the human annotation. One interpretation of this is that the developed method
287 can predict PSEPs with a higher accuracy than the reference predictions based
288 on manual annotation given that two or more leaves are detected for each PSEP.

289 **5. Conclusion**

290 A system for automated PSEP estimation of sugar beet plants (in growth
291 stages BBCH10-14) based on leaf detection has been developed and tested.
292 In a set of test images the system detected 46.7% of the present leaves. A
293 multivariate Gaussian PSEP model was built based on the detected leaves and
294 manual annotation of true PSEPs. Given centre of mass and attach point of a
295 single leaf the model states that the average true PSEP will be at a distance
296 of 6.2mm from the petiole attachment point and placed on the line connecting
297 the leaf attach point and the leaf centre of mass. 95% of the volume below the
298 multivariate Gaussian is contained within an ellipse with semi major and semi
299 minor axes of 12mm and 6mm respectively.

300 In the set of test images the detected leaves were used to predict the true
301 PSEPs. With PSEP prediction based on single leaves were 90% of the true
302 PSEPs located within 20mm of at least one predicted PSEP location. In this
303 case where the average distance from predicted location to true PSEP of 3.3mm.
304 When several leaves of the same plant are detected, the PSEP models can
305 be combined using least-squares estimation and thus produce an even better
306 estimate of the true root location. E.g. by combining two leaves the average
307 error is reduced to 1.9mm. Precise quantification of error in three and four leaf
308 based PSEP estimates is hindered as these methods perform on par with the
309 human annotation used as reference.

310 Backes, M., Jacobi, J., 2006. Classification of weed patches in quickbird images:
311 Verification by ground truth data. EARSeL European Association of Remote
312 Sensing Laboratories 5 (2), 172–179.
313 URL http://www.eproceedings.org/static/vol05_2/05_2_backes1.html

314 Franz, E., Gebhardt, M., Unklesbay, K., 1991. Shape description of completely
315 visible and partially occluded leaves for identifying plants in digital images.
316 Vol. 34. p. 673–681.

317 Gales, M., Airey, S., Jan. 2006. Product of gaussians for speech recognition.
318 Computer Speech & Language 20 (1), 22–40.
319 URL <http://www.sciencedirect.com/science/article/B6WCW-4FBHW7F-1/2/b1001c29af3057ecce30bf>

320 Griepentrog, H. W., Dedousis, A. P., 2010. Mechanical Weed Control. Vol. 20.
321 Springer Berlin Heidelberg, Ch. 11, pp. 171–179.

322 Griepentrog, H. W., Nørremark, M., Nielsen, H., Blackmore, B. S., 2005. Seed
323 mapping of sugar beet. Precision Agriculture 6, 157–165, 10.1007/s11119-005-
324 1032-5.
325 URL <http://dx.doi.org/10.1007/s11119-005-1032-5>

326 Kouwenhoven, J. K., 1997. Intra-row mechanical weed control—possibilities and
327 problems. Soil and Tillage Research 41 (1-2), 87 – 104.
328 URL <http://www.sciencedirect.com/science/article/B6TC6-3RGTBJD-7/2/3ce3ed554667f0adfed53e>

- 329 Meier, U., 2001. Growth stages of mono-and dicotyledonous plants.
330 URL <http://www.bba.de/veroeff/bbch/bbcheng.pdf>
- 331 Melander, B., 2000. Mechanical weed control in transplanted sugar beet. In: 4th
332 EWRS Workshop on Physical Weed Control.
333 URL http://orgprints.org/1542/1/Abstract_Elspeet1.pdf
- 334 Neto, J. C., Meyer, G. E., Jones, D. D., Apr. 2006. Individual leaf extractions
335 from young canopy images using gustafson-kessel clustering and a genetic
336 algorithm. *Computers and Electronics in Agriculture* 51 (1-2), 66–85.
337 URL <http://www.sciencedirect.com/science/article/B6T5M-4J2M44F-1/2/b947a39d2c204692a56e4a>
- 338 Nørremark, M., Griepentrog, H., Nielsen, J., Søgaaard, H., 2008. The develop-
339 ment and assessment of the accuracy of an autonomous gps-based system
340 for intra-row mechanical weed control in row crops. *Biosystems Engineering*
341 101 (4), 396 – 410.
342 URL <http://www.sciencedirect.com/science/article/B6WXV-4TX6W5S-1/2/8aede62984cd66ed1ef62a>
- 343 Poulsen, F., 2010. Measuring emerging plants using machine vi-
344 sion. [http://www.visionweeding.com/Products/Plant-Counting/Plant-](http://www.visionweeding.com/Products/Plant-Counting/Plant-Counting.htm)
345 [Counting.htm](http://www.visionweeding.com/Products/Plant-Counting/Plant-Counting.htm).
346 URL <http://www.visionweeding.com/Products/Plant-Counting/Plant-Counting.htm>
- 347 Sun, H., Slaughter, D. C., Ruiz, M. P., Gliever, C., Upadhyaya, S. K., Smith,
348 R. F., Apr. 2010. Rtk gps mapping of transplanted row crops. *Computers and*
349 *Electronics In Agriculture* 71 (1), 32–37.
- 350 Tillett, N., Hague, T., Grundy, A., Dedousis, A., 2008. Mechanical within-row
351 weed control for transplanted crops using computer vision. *Biosystems*
352 *Engineering* 99 (2), 171 – 178.
353 URL <http://www.sciencedirect.com/science/article/B6WXV-4R5H1V8-2/2/4487816230cc4673f885d6>
- 354 Åstrand, B., Baerveldt, A.-J., Jul. 2002. An agricultural mobile robot with
355 vision-based perception for mechanical weed control. *Autonomous Robots*
356 13 (1), 21–35.
357 URL <http://dx.doi.org/10.1023/A:1015674004201>



Figure 1: The camera unit consisted of camera combined with halogen lamp. During image acquisition were eight such units mounted in front of a tractor.

Set	# leaves	Count	FP	MR	Avg	95%
D_1	1	395	4.8%(19)	10.0%	3.29 ± 0.14	15.76
D_2	2	313	1.6%(5)	37.3%	1.88 ± 0.07	4.62
D_3	3	132	0.8%(1)	70.1%	1.42 ± 0.09	3.02
D_4	4	29	0.0%(0)	89.1%	1.22 ± 0.20	2.39
D_5	1-4	188	8.0%(15)	10.4%	2.66 ± 0.21	49.51
D_6	na	71	0.0%(0)	2.7%	1.37 ± 0.26	3.58

Table 1: Count: Number of position estimates. FP: False positives, percentage of predicted plant positions with a distance to the nearest true plant location larger than 20mm. MR: Missed roots, percentage of true PSEPs within 20mm of a predicted PSEP location. Avg: Average estimate error in mm. 95%: 95% quantile of estimate errors in mm.

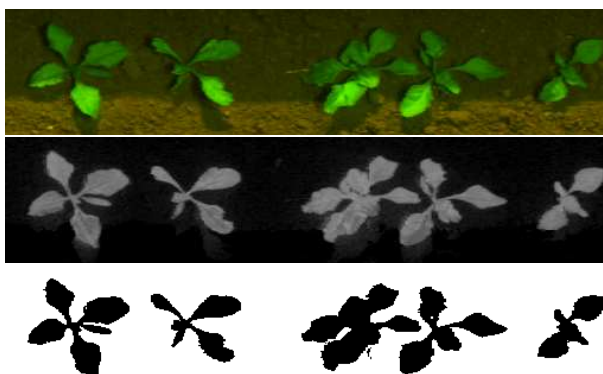


Figure 2: Plant segmentation was done in two steps. First were NDVI values calculated for each pixel, then was the image thresholded. The shown images are (a) pseudo RGB image of raw data (red is shown as red and NIR is shown as green while the blue channel is set to zero) (b) NDVI image before thresholding and (c) after thresholding.

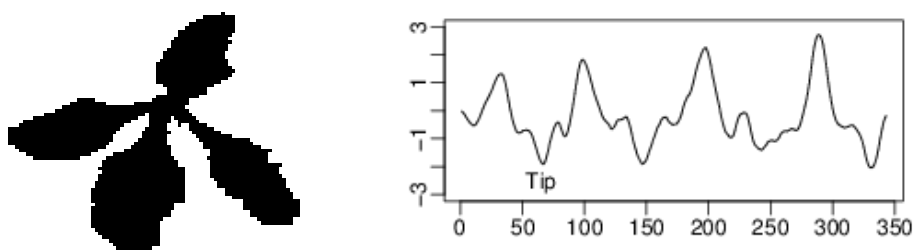


Figure 3: Example of plant boundary and the calculated curvature along the boundary. The boundary is followed clockwise. Leaf tips are local minima and locations near the PSEP corresponds to peaks.

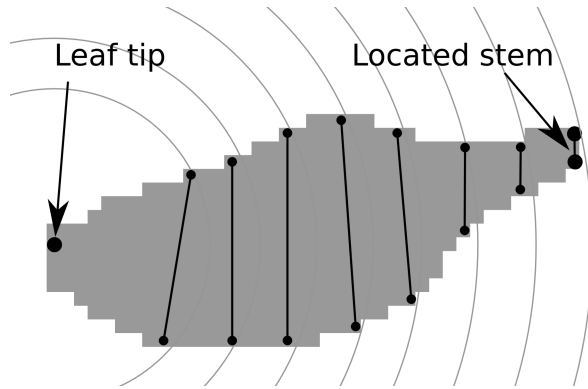


Figure 4: Visualization of the search strategy. The boundary is followed from the leaf tip until the euclidean distance between the current location and the leaf tip exceeds a specified threshold. This is done in both directions and distance between the located points is measured. The procedure is repeated with increasing distance thresholds illustrated by concentric circles. When distance between located points is minimized the leaf cut-off location is found.

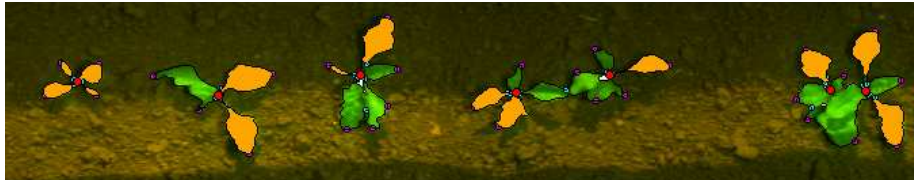


Figure 5: Manually marking of PSEPs. The orange leaves were detected by the leaf detector. PSEPs are marked with a red spot.

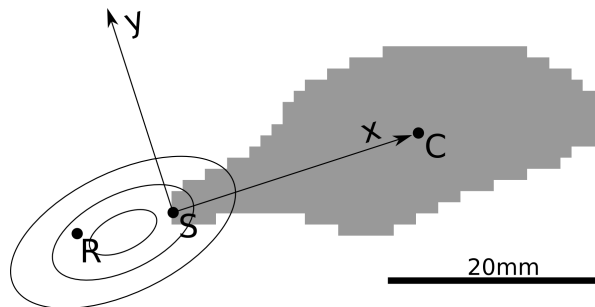


Figure 6: PSEP location as specified in the leaf coordinate system. The following points are marked: centre of mass C , stem attach point S and PSEP location R . The PSEP location model is indicated by the three concentric ellipses. According to the PSEP location model, will 68% of the true PSEP locations be placed within the central ellipse, the two other ellipses will contain 95% and 99.7% respectively.

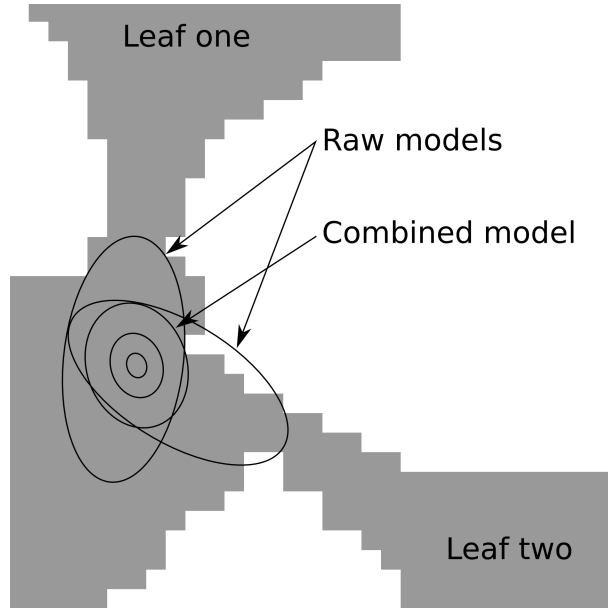


Figure 7: Combination of two PSEP location models. The ellipses contains are similar to those shown in figure 6. For the raw models are the ellipse for 99.7% shown and for the combined model: 68%, 95% and 99.7%.

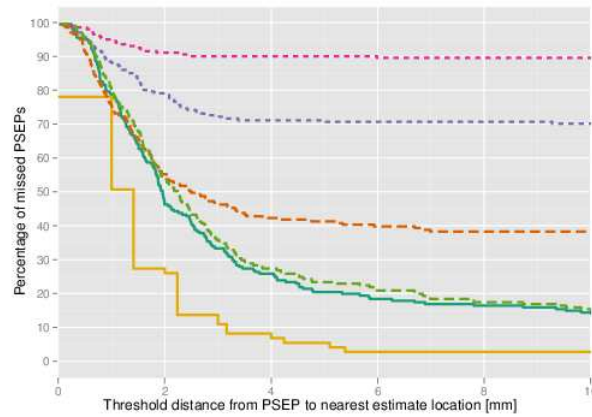


Figure 8: Fraction of missed PSEPs as a function of the threshold distance. When the number of leaves used to estimate true PSEPs are increased the fraction of missed PSEPs also increases. The following color encoding is used: ■ D1, ■ D2, ■ D3, ■ D4, ■ D5 and ■ D6

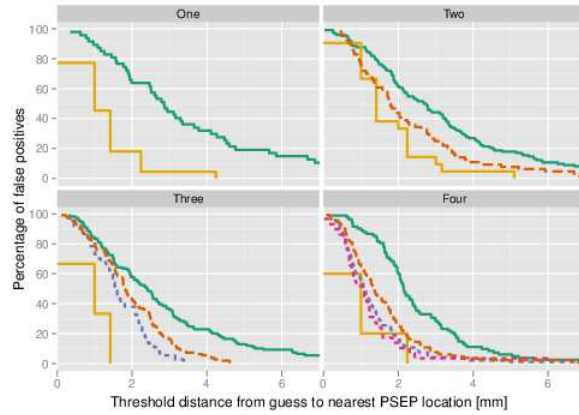


Figure 9: Fraction of false positives as a function of the threshold distance. Error of PSEP-location-estimates is seen to decrease when the number of leaves used to make the estimate is increased. Color encodings as in figure 8.

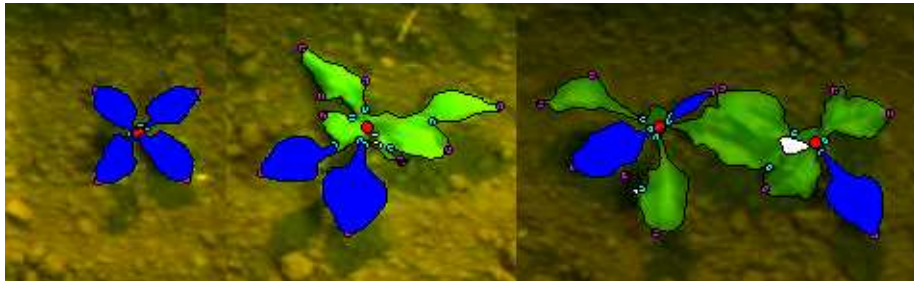


Figure 10: Easy and difficult cases for the leaf detector. Leaf tip candidates are marked by purple squares. Cyan indicates concave locations. Detected leaves are marked in blue.