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(Article begins on next page)

1	CONVENTIONAL AND ULTRASOUND-ASSISTED EXTRACTION OF RICE
2	BRAN OIL WITH ISOPROPANOL AS SOLVENT
3	
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### 17 Abstract

After cereal harvesting, rice is subjected to several milling processes to remove hull, germ, and bran and produce the final white rice. The bran represents around 10% of total grain weight and is usually considered as waste material. One of the most common rice bran applications is the extraction of rice bran oil, rich in  $\gamma$ -oryzanol, which has shown many health benefits including antioxidant, anti-inflammatory, and antihypercholesterolemic properties. Rice bran oil is usually extracted by organic solvents, which are toxic for health and the environment. In this work, rice bran oil was extracted through isopropanol extraction, and the best-operating temperature and bran to solvent ratio have been identified. After that, an ultrasound-assisted extraction was conducted at room temperature and with the same rice bran to solvent ratio of the isopropanol extraction.

29 The kinetics evaluation through Peleg's model showed that the solvent extraction 30 reaches the steady-state after 15 minutes while the ultrasound-assisted extraction 31 reaches the steady-state after only 1 minute producing very similar yields in rice bran oil 32 and  $\gamma$ -oryzanol. Comparing these two green extraction techniques through a life cycle 33 assessment, it has emerged that with the same amount of rice bran oil produced, the 34 ultrasound-assisted extraction is the less environmentally impacting process. The room 35 temperature ultrasound-assisted extraction allows minimizing-the energy and time 36 consumption demonstrating to be a sustainable process in line with the principles of 37 green chemistry.

38

Keywords: Rice bran oil, green extraction, ultrasound-assisted extraction, life cycle
assessment.

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42

#### 43 **1. Introduction**

Rice is one of the most important food crops in the world, representing a huge contribution to the dietary need of the global population [1]. According to FAO, world rice production exceeded 755 million tons in 2019 [2]. After harvesting, the rice grain undergoes a milling process to remove all the external layers making the edible white rice kernel [3]. During these milling operations, around 40% of the total grain is lost due to discarding the byproducts, including the husk, the bran, the germ, and the broken

50 rice [4]. Usually, these are burned or used as animal feed, but they may represent an 51 excellent source of bioactive compounds, making them suitable for nutraceutical, 52 cosmetic and pharmaceutical applications [2]. In particular, rice bran, which represents 53 around 10% of the grain weight, contains proteins, fibers, and oil, this latter rich in 54 bioactive and antioxidant compounds [5]. This rice bran oil (RBO) presents a balanced 55 fatty acid composition and high levels of functional molecules, including phytosterols, 56 tocopherols, tocotrienols, and other nutrients. Thanks to its exceptional proprieties, 57 RBO is commonly used in Asian countries where it is considered as a "healthy oil". 58 Indeed, it has been demonstrated that RBO has antihypertensive, antidiabetic, anti-59 obesity, and anticarcinogenic properties due to its significant antioxidant and anti-60 inflammatory activity [6–8]. Several studies confirmed that one of the main responsible 61 for these beneficial effects is  $\gamma$ -oryzanol, an antioxidant mixture of ferulic acid esters of 62 phytosterols, present at high levels in RBO [9]. The conventional method used for the 63 commercial extraction of RBO is solvent extraction (SE) using hexane, a petroleum-64 derived, flammable, and toxic solvent, dangerous for human health and the 65 environment. The disadvantages of this type of extraction led most researchers to look 66 for alternative approaches, focusing on non-conventional and non-thermal extraction 67 techniques, for RBO extractions [10]. These innovative techniques employ less 68 dangerous solvents, often combined with one or more process intensification steps to 69 reduce time and energy waste, obtaining high-quality extracts devoid of toxic residues 70 [2, 11]. Some studies demonstrated that short-chain alcohols might represent an 71 excellent alternative to hexane as a solvent in RBO extraction. In particular, the use of 72 isopropanol allowed to obtain a high yield in oil and  $\gamma$ -oryzanol [12].

73	Moreover, in recent years, ultrasound-assisted extraction (UAE), thanks to the
74	phenomenon of "acoustic cavitation," has become an effective green method for oil
75	extraction. In their review, Mushtaq et al. [13] concluded that the UAE represents a
76	good alternative for extracting edible oil. This technique allows operating at low
77	temperatures and times, reducing solvent consumption, avoiding thermal damage and
78	preserving their structural and bioactive properties. Indeed, UAE was successfully
79	applied to extract bioactive compounds from several natural matrices, but only a few
80	studies on RBO extraction focus on this green technique [14].
81	The present work aims to show the suitability of isopropanol as non-conventional
82	solvent for RBO extraction and then demonstrate that the substitution of high
83	temperature with room-temperature ultrasound in the extraction with isopropanol as the
84	solvent allows minimizing the extraction time and energy consumption, obtaining
85	comparable yields.
86	Isopropanol SE and room temperature UAE were compared in terms of oil yield and $\gamma$ -
87	oryzanol content. The extraction kinetics were determined using Peleg's model to
88	identify the best extraction time. Moreover, a life cycle assessment (LCA) study was
89	performed to compare the environmental sustainability of-the two processes and and to
90	choose the most environmental-friendly extraction process.
91	
92	2. Materials and methods
93	2.1 Material and chemicals
94	Cryo-milled rice bran sample, with a particle diameter of 500 $\mu$ m, were supplied by

95 Agrindustria Tecco S.R.L. and stored at -20.0 °C until extraction. HPLC grade hexane,

96 methanol, acetonitrile, acid acetic and isopropanol used for extraction and high-

97 performance liquid chromatography (HPLC) analysis and γ-oryzanol standard for
98 quantification were purchased from Merck (Darmstadt, Germany).

#### 99 **2.2 Isopropanol extraction**

100 Rice bran was mixed with isopropanol in a Pyrex reaction flask connected with a Liebig

101 reflux condenser. The flask was put into a water bath with a magnetic stirrer. To choose

102 the best condition, 1:3, 1:6, and 1:9 solid-to-solvent ratio (w/v) was used at different

103 temperatures (30, 45, 60 °C). The extraction time was fixed for 1 h. After these

104 preliminary studies, the best solid-to-solvent ratio was chosen. The temperature range

105 was extended to determine the effect of temperature on the yield of extracted

106 components. The temperature investigated were 30, 60, 90, and 120 °C. At the best

107 temperature, a kinetic study was performed examining the yield in oil and γ-oryzanol

against time from 1 minute to 120 min.

### 109 **2.3 Ultrasound-assisted extraction**

110 For the UAE experiments, a VCX750 Ultrasonic Processors (Sonic and Materials Inc.),

111 with a frequency of 20 kHz and 40 % of amplitude, equipped with a 13 mm probe was

112 used. Rice bran (5 g) was mixed with 45 mL of isopropanol (1:9 w/v) in a Pyrex

113 reaction flask put into a water bath with a magnetic stirrer. The temperature was

114 maintained at room temperature (25 °C) and controlled with a thermocouple. A kinetic

study was performed investigating the yield in oil and  $\gamma$ -oryzanol against time from 10 s

116 to 30 min.

### 117 **2.4 Hexane extraction**

Isopropanol and UAE were compared with a conventional hexane extraction following
the method of Pengkumsri et al. [15]. Rice bran was mixed with hexane in a 1:10 (w/v)

ratio in a Pyrex reaction flask connected with a Liebig reflux condenser. The flask was
put into a water bath with a magnetic stirrer at 40 °C for 30 min.

# 122 **2.5 Determination of RBO Yield**

123 After the extractions, all the samples were filtered two times using a paper filter

124 Whatman grade 1 to separate the liquid phase from the exhausted rice bran. Then the

125 solvent was evaporated using a Heidolph Rotary Evaporator, Laborota 4000. The RBO

126 yield was calculated on the base of Eq. 1:

127

$$RBO Yield (\%) = \frac{RBO (g)}{Rice \, bran (g)} \times 100 \tag{1}$$

#### **2.6 Determination of γ-oryzanol content**

129 After the evaporation of the solvent and the determination of the oil yield, the RBO 130 samples were resuspended in 15 mL of isopropanol, and  $\gamma$ -oryzanol content was 131 determined by reversed-phase HPLC. The HPLC system (Shimadzu 20A Prominence) 132 was equipped with a Kinetex C18 column (5 µm, 150 x 4.6 mm) by Phenomenex and 133 photodiode array (PDA) detector using an isocratic elution. The mobile phase was 134 composed by methanol, acetonitrile and 0.03 % acid acetic at a ratio 52:45:3 (v/v/v) [16, 135 17]. The flow rate was maintained at 0.8 mL/min, and the column oven was 136 thermostated at 30 °C.  $\gamma$ -oryzanol content was determined through a calibration curve 137 prepared using 8 different concentrations of  $\gamma$ -oryzanol standard (0.01-0.8 mg/mL) in 138 isopropanol. The limit of detection (LOD) and the limit of quantification (LOQ) were 139 calculated with the following equation, where  $\sigma$  is the standard deviation of the response 140 and S is the slope of the calibration curve [18]. LOD and LOQ were respectively 0.01

- 141 and 0.04 mg/mL.
- 142
- 143

$$LOD = \frac{3.3 \times \sigma}{S} \tag{2}$$

$$LOQ = \frac{10 \times \sigma}{S} \tag{3}$$

144 The  $\gamma$ -oryzanol yield was calculated using the formula:

$$\gamma - oryzanol \ yield = \frac{\gamma - oryzanol \ (mg)}{Rice \ bran \ (g)}$$
 (4)

145

#### 146 **2.7 Statistical analysis**

- 147 All the extraction experiments were performed in triplicate and analyzed by one-way
- 148 ANOVA (analysis of variance) with a Tukey's posthoc test ( $P \le 0.05$ ), after the
- 149 assessment of the fundamental assumptions of ANOVA: the normality of distributions
- 150 (Shapiro-Wilk test, p-value N 0.05) and the homogeneity of the variances of the
- residuals (Levene's test with P(NF) N 0.05). The statistical software R (version 4.0.4 -
- 152 Feather Spray 2021) was used for all.

153

#### 155 **2.8 Extraction kinetics**

To describe the SE and UAE kinetics of RBO and γ-oryzanol from rice bran was used
the model proposed by Peleg [19], a two-parameters, non-exponential empirical
equation, originally proposed to describe sorption curves and adapted for the extraction
process in the form:

$$C(t) = C_0 + \frac{t}{K_1 + K_2 \times t}$$
(5)

160

161 where C(t) is the RBO or  $\gamma$ -oryzanol yield [(g RBO / g bran)·100] or [mg  $\gamma$ -oryzanol / g 162 bran], respectively, at time t; t is the extraction time [min], C<sub>0</sub> is the yield at time t = 0, 163 K<sub>1</sub> is Peleg's rate constant [min· (g bran / g RBO) ·100] or [min· (g bran / mg  $\gamma$ -164 oryzanol)], and K<sub>2</sub> is Peleg's capacity constant [g bran · 100 / g RBO] or [g bran / mg  $\gamma$ -165 oryzanol]. Since, C<sub>0</sub> is considered zero, and this term can be omitted from Peleg's 166 equation, the final form of the equation used is:

$$C(t) = \frac{t}{K_1 + K_2 \times t} \tag{6}$$

167

It should be noted that a lower K<sub>1</sub> value means a faster rate of extraction, and a lower K<sub>2</sub>
value suggests maximum yield [20]. The Peleg's rate constant K<sub>1</sub> relates to the

170 extraction rate ( $B_0$ ) at the start (t = t<sub>0</sub>).

$$B_0 = \frac{1}{K_1} \tag{7}$$

- 172 The Peleg capacity constant K<sub>2</sub> relates to a maximum of extraction yield, C<sub>e</sub> at
- 173 equilibrium when  $t = \infty$ .

$$C_{t\to\infty} = C_e = \frac{1}{K_2} \tag{8}$$

- 175 The accordance of experimental  $(\hat{y}_i)$  data and model-predicted results  $(y_i)$  were
- 176 established by correlation coefficient ( $R^2$ ), and root mean square error (RMSE) as
- 177 follows, where n represents the number of experiments.

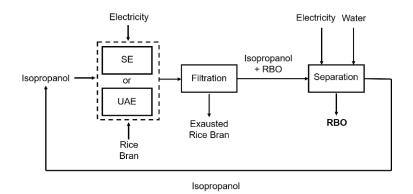
$$RMSE = \sqrt{\frac{\sum_{i=1}^{n} (\hat{y} - y)^2}{n}}$$
(9)

178

### 179 **2.9 Life cycle assessment**

- 180 LCA was performed with SimaPro 9.0.48 software, database Ecoinvent 3.0
- 181 **2.9.1 Goal and scope**
- 182 The goal of LCA was to compare SE vs. UAE of RBO, to choose the best extraction
- 183 process in terms of environmental sustainability.
- 184 The functional unit (FU) was 1 g of RBO produced.
- 185 The boundary conditions are shown in Fig. 1. Briefly, the entire process is divided in
- 186 three steps: extraction (SE or UAE), filtration and separation. In the extraction step,
- 187 the entered flows of matter and energy are isopropanol, rice bran and electricity. The
- 188 rice bran and the isopropanol come out of the extraction together and enter the second
- 189 step, where filtration occurs. Here the exhausted rice bran, which is discarded, and the

- 190 isopropanol containing the RBO are separated. Isopropanol and RBO enter the last step
- 191 where they are separated by a rotavapor which needs cooling water and electricity. In
- this step, the isopropanol is removed from the RBO and recirculated for a new
- 193 extraction.





- 195 **Fig. 1** LCA boundary conditions.
- 196

### 197 **2.9.2 Life cycle inventory**

- 198 The life cycle inventory (LCI) defined all inputs and outputs involved in the processes.
- 199 The primary data came from the present study, the produced emissions, the consumed
- 200 material, and the required energy were referred to this FU.
- 201 Expansion system methodology was applied to the recovery of isopropanol in the
- 202 separation step. The secondary data were taken from Ecoinvent 3.01 and reported in
- 203 Table 1.
- 204
- 205
- 206
- 207

209

210 **Table 1** Secondary data from Ecoinvent 3.01

Electricity	Electricity, medium voltage {Europe without Switzerland market}  Alloc
	Def Unit
Isopropanol	Isopropanol {GLO} market for  Alloc Def Unit
Rice bran	Rice bran from dry milling at plant   CN mass
Water	Tap Water from natural resource

211

212

# 213 **2.9.3 Life cycle impact assessment**

Life cycle impact assessment (LCIA) was performed with the ReCIPE Midpoint (H)

215 method. In the present study, the analyzed impact category were: Climate change (kg

216 CO<sub>2</sub> eq), Ozone depletion (kg CFC-11 eq), Human toxicity (kg 1,4-DB eq) and

217 Freshwater eutrophication (kg P eq).

218

# 219 **3. Results and discussion**

### 220 **3.1 Solvent extraction**

221 The solvent chosen for all the extractions experiments was isopropanol. Traditionally,

222 RBO was extracted using hexane as the solvent because it presents a low corrosiveness,

223 high stability, and a good capacity for dissolving oil and relevant compounds such as γ-

oryzanol [12, 21–23]. Despite these advantages, hexane presents health and

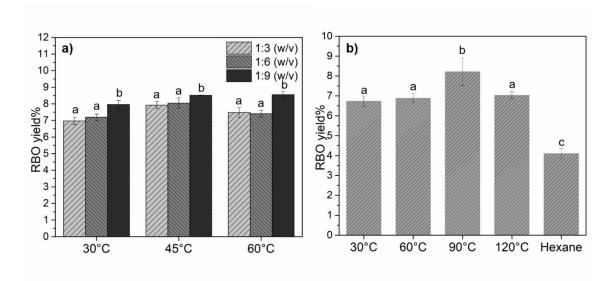
225 environmental risks, therefore researchers and oil industries are focusing on reliable 226 alternative solvents. Moreover, hexane derives from a non-renewable source and 227 presents high flammability and toxicity for the environment and human health [22]. A 228 few types of solvents have been tested to substitute hexane as an extractant of vegetable 229 oil. In particular, short-chain alcohols, including isopropanol, are particularly promising 230 due to their low toxicity [23]. Isopropanol is labeled as "recommended" or "preferred" 231 in several green-solvent selection guidelines, such as the GlaxoSmithKline (GSK), 232 Pfizer, Sanofi, AstraZeneca, and Green Chemistry Institute-Pharmaceutical Roundtable 233 (GCI-PR) [2]. Furthermore, all the components of  $\gamma$ -oryzanol present an alcohol group 234 deriving from the ferulic acid; this increases the polarity, making them more soluble in 235 polar solvents such as short-chain alcohols, including isopropanol [21]. 236 At the beginning of this work, a preliminary study on the effects of different solid-to-237 solvent ratios at various temperatures was performed. After that, using the best solvent-238 to-solid ratio condition, the temperature range was extended to determine the effect of 239 temperature on the yield of extracted components. At this step, all the extractions were 240 conducted for 1 hour. Figure 2 a) shows the oil yield using 1:3, 1:6, and 1:9 (w/v) solid-241 to-solvent ratios at three different temperatures (30, 45, and 60  $^{\circ}$ C). Remarkably, the 242 yield of RBO grows with the increase of the solvent volume at any temperatures tested. 243 This result is in line with Ruen-Ngam et al. [24], which investigated the use of different 244 solvents with an increasing solid-to-solvent ratio and stated that a low volume of solvent 245 does not allow its complete penetration into the rice bran material. The same trend was 246 noted by Hu et al. [12], which obtained a higher yield of RBO and  $\gamma$ -oryzanol, 247 increasing the solvent-to-bran ratio, using both hexane and isopropanol. Indeed the 248 solvent extraction of RBO is mainly considered as a mass transfer process between the

249 solid and liquid phases, where the oil passes from the bran powder to the solvent 250 through a diffusion mechanism [25]. The effect of the solid-to-solvent ratio on the 251 extraction yield is coherent with the mass transfer process. The concentration gradient 252 between the solid and the liquid represents the driving force of this phenomena [26], 253 and it is more significant when the solid is in contact with a large volume of solvent. 254 The high quantity of isopropanol reduces the saturation level of the solvent, increasing 255 oil extraction yields [26]. The same results have been found for the alcoholic SE of 256 antioxidants compounds from different natural sources such as grape pomace, olive 257 leaves or stonebreaker [27–29]. 258 Because no striking differences were noted between yields at the previously mentioned

temperatures, it was chosen to test the 1:9 solid-to-solvent ratio at 30, 60, 90, and 120
°C to understand if the high temperatures can affect the extraction. As reported in Fig.
2b, a temperature of 90 °C (p<0.05) gave the best results in terms of RBO extraction</li>
yield.

263 Indeed, the temperature is one of the most significant factors influencing the mass 264 transfer and the RBO yield [25]. Capellini et al. [23] performed RBO extraction using 265 isopropanol and ethanol at different temperatures (50, 60, 70, and 80 °C). They found 266 that temperature increase resulted in a growth in oil yield, regardless of the solvent. The 267 same results had been found by, Imsanguan et al. [28], Xu and Godber [19], and Oliveira 268 et al. [20]. All these authors [19, 20, 28] agree that the diffusivity and the solubility of 269 the compounds to extract increase with the increase of temperature, and the extraction 270 output is enhanced [30]. At high temperatures, degradation of the sample matrix 271 structure occurs, making it more permeable to the solvent. Moreover, a temperature rise 272 produces a decrease in solvent viscosity, increasing its diffusivity. However, the high

273	temperature may cause the degradation of thermolabile compounds and the evaporation
274	of the solvent. This has been reported for different plant matrices such as tomatoes,
275	grape waste material and muitle [20, 31–33]. Furthermore, all these extractions have
276	been compared to hexane extraction. As can be noted from Fig. 2, the conventional
277	hexane extraction, proposed by Pengkumsri et al. [15], and used as the reference
278	method, produced only 4.1 $\pm$ 0.25 g of RBO/100 g of rice bran. This value is lower than
279	the yields obtained with isopropanol in any operating condition, which reaches the
280	maximum of 8.21 $\pm$ 0.69 g of RBO/100 g of rice bran at 90°C and bran to solvent ratio
281	of 1:9, demonstrating that isopropanol is a solvent suitable to replace hexane in RBO
282	extractions. These results are confirmed by Ruen-Ngam et al., Oliveira et al. and
283	Capellini et al. [22, 24, 34] which affirm that alcoholic solvents like isopropanol, can
284	extract higher amounts of oil than hexane due to their high polarity. In particular,
285	isopropanol can extract higher quantities of phospholipids and unsaponifiable material
286	from solid matrices and obtain RBO richer in $\gamma$ -oryzanol. Indeed, the scructure of $\gamma$ -
287	oryzanol presents both sides of polar and non-polar (ferulic acid and sterol) making this
288	group of molecules particularly soluble in short-chain alcohols which show dielectric
289	constant higher than hexane and lower than water [24].



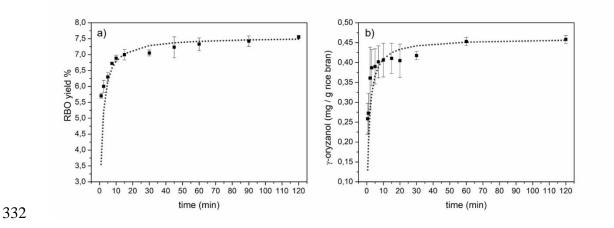
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Fig. 2 a) Effect of bran to solvent ratio at different temperatures on RBO yield (%). b)
effect of higher temperature on the RBO yield (%) with 1:9 (w/v).

### **3.2 SE kinetic**

296 The kinetic study was performed measuring the yields in RBO and  $\gamma$ -oryzanol against 297 the time, starting from 1 min up to 2 h. RBO yield % and  $\gamma$ -oryzanol were plotted in 298 function of time (Fig. 3). The shapes of the graphs indicate that the extraction yields are 299 significantly time-dependent. The yields in RBO and  $\gamma$ -oryzanol rise rapidly with time 300 initially, and then, after the knee at about 15 min, start to keep constant producing 6.89 301  $\pm$  0.089 g of RBO/ 100 g of rice bran and 0.41  $\pm$  0.038 mg of  $\gamma$ -oryzanol/g of rice bran. 302 These profiles fit with the typical SE curve composed of a fast extraction step called 303 "washing phase", followed by a slower extraction step called "diffusion step" [35]. 304 Indeed, the mass transfer rate into the solvent is exceptionally high at the beginning of 305 extraction, when the solvents penetrate the rice bran, thanks to the elevated 306 concentration gradients. Progressing with the extraction, the solutes diffuse from the 307 interior of the bran to the solvent. The mass transfer of solutes becomes more difficult

308 because the concentration gradient between the solid and the liquid phase decreases and 309 the extraction rate becomes slower [20, 35]. The obtained constants of the model (rate 310 constant K<sub>1</sub>, constant capacity K<sub>2</sub>) and the calculated parameters (initial extraction rate 311  $(B_0)$  and the maximum yield extraction  $(C_e)$ , regression coefficient  $(R^2)$ , and the root 312 mean square error (RMSE) are reported in Table 2. The model fitted well with the 313 experimental data, with reasonable accuracy, as evidenced by high  $R^2$  and low RMSE. 314 In particular, experimental data fit slightly worse in the case of  $\gamma$ -oryzanol yield, but this 315 could be due to the multiple treatments of the sample before the HPLC analysis, causing 316 uncertainty in measurement and high standard deviation. In both cases, there was 317 concordance between experimental and predicted yield values. Yields are lower than 318 those reported in the literature [5, 36, 37], but this is certainly due to different 319 experimental conditions and, above all, to the type of rice bran used for the extraction. 320 Indeed the composition of rice bran changes based on the rice variety, the growing 321 conditions, the milling system employed and the stabilization process [2]. Peleg's 322 equation is one of the most suitable models to describe the SE from plant matrix as 323 demonstrated by several studies. Karacabey et al. [38] compared the first-order kinetic 324 model, Peleg's model, two-site kinetic model and modified Gompertz equation to 325 describe solid–liquid extraction kinetics of trans-resveratrol from grape cane. Jurinjak 326 Tusek at al. [39] compared Peleg's, Page's, and Logarithmic model for total 327 polyphenols, antioxidants extraction yield from Asteraceae plants. Poojary and 328 Passamonti [20] the first-order kinetic model, the mass transfer model, and Peleg's 329 model for understanding the behavior of lycopene extraction from tomato processing 330 waste. All these authors agree that Peleg's model showed a better fit to the experimental 331 data than other models investigated in their studies.



**Fig. 3** Isopropanol extraction (SE) kinetics of a) RBO and b)  $\gamma$ -oryzanol. Black squares represent the experimental values. Each experiment was conducted in triplicate, and the error bars correspond to standard error. Dotted lines represent the modeled values by Peleg's equation.

### **338 3.3 UAE kinetic**

After determining the best isopropanol extraction conditions in terms of temperature
and bran to solvent ratio, an intensification process step was performed to work at room
temperature and reduce the reaction time. The extraction was conducted with the help of
ultrasound as described in Paragraph 2.3, maintaining the bran to solvent ratio of 1:9, as
in the previous experiments. The extraction yields in RBO and γ-oryzanol in the

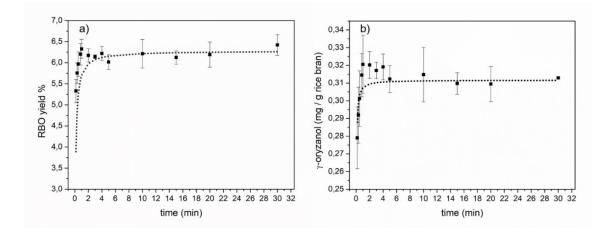
344 function of time were plotted to find the best extraction time and to understand the

345 process kinetic (Fig. 4). These two yields were initially measured every 10 s up to 1 min

and then less frequently up to 30 min. As shown in Fig.4 the UAE reaches the steady-

- 347 state after only 1 minute producing  $6.33 \pm 0.22$  g of RBO/100 g of rice bran and  $0.32 \pm$
- $0.016 \text{ mg of } \gamma$ -oryzanol/g of rice bran. Peleg's model was adapted to experimental
- 349 conditions and used for data approximation (see paragraph 3.2) and the results are

350 reported in Tab. 2. The initial extraction rate  $(B_0)$  is much higher than the  $B_0$  found for 351 the isopropanol extraction. Indeed, the knee of the curve occurs at about 1 min, where 352 yields reach the maximum values and then stabilize. The maximum yield extraction (C<sub>e</sub>) is slightly lower than SE and the  $R^2$ , but this may be due to the difficulty of manually 353 354 measuring the yield every 10 s. This difficulty caused a high uncertainty and an elevated 355 standard deviation producing the worst fit with the model. Until now, to the best of the 356 authors' knowledge, previous studies on the combination of isopropanol and UAE do 357 not exist. Cravotto et al. [40] and Khoei et al. [41] studied the RBO extraction using 358 UAE and water as the solvent, demonstrating that the ultrasound is suitable for aqueous 359 extracting rice bran oil. Other authors studied the effect of UAE of RBO combined with 360 short-chain alcohols like methanol and ethanol [42, 43], demonstrating the feasibility of 361 this kind of extraction. Still, no one managed to complete the extraction in such a short 362 time. Kumari et al. [41] and Galvan et al. [42] showed in their works that Peleg's model 363 efficiently describes the kinetic of UAE for other plant matrixes such as potato peels 364 and black chokeberry wastes.



365

Fig. 4 UAE kinetics of a) RBO and b) γ-oryzanol. Black squares represent the
experimental values. Each experiment was conducted in triplicate, and the error bars

368 correspond to standard error. Dotted lines represent the modeled values by Peleg's369 equation.

#### 370 3.4 SE and UAE kinetics comparison

371 The obtained constants of the model (rate constant K1, constant capacity K2) and the 372 calculated parameters, initial extraction rate  $(B_0)$  and the maximum yield extraction 373  $(C_e)$ , regression coefficient  $(R^2)$ , and the root mean square error (RMSE), for the two 374 kinds of extractions, are reported in Table 2. As can be noted, the initial extraction rate 375 is higher for the UAE than SE, demonstrating that UAE can reach a steady state in a 376 shorter time. Assuming infinite extraction time, the maximum yield obtained is 7.55 g 377 RBO/100 g bran and 0.46 mg  $\gamma$ -oryzanol/g bran for the SE and 6.34 g RBO/100 g bran 378 and 0.31 mg  $\gamma$ -oryzanol/g bran for the UAE. SE allows obtaining slightly higher 379 quantities of RBO and γ-oryzanol at 90°C, but UAE allows to reach the maximum yield 380 in just one minute and therefore in much shorter times, operating at room temperature. 381 In their work, Mohammed Danlami at al. [43] and Zhang et al. [44] compared 382 traditional SE with that of other extraction techniques to extract valuable components 383 from plants. They affirmed that ultrasound facilitates the extraction of thermally 384 sensitive compounds enhancing the extraction rate and reducing the extraction 385 temperature. Khoei and Chekin [44], in their works, extracted RBO using aqueous 386 extraction and compared the conventional SE with UAE. The two extraction techniques 387 allowed to obtain very similar RBO yield and they demonstrated that the application of 388 ultrasound permitted to work at room temperature in a shortened extraction time. The 389 global production of RBO exceeds 1.7 million tons per year [11]. Although the use of 390 ultrasound at room temperature leads to a slightly lower oil yield, on such a large annual 391 production this decrease may not be relevant. Using UAE instead of SE would allow to

392 increase the global production of RBO as the extraction has a shorter duration and it is 393 therefore possible to increase the number of annual extractions. The results showed in 394 the present work are in accordance with the literature studies cited, the yields obtained 395 with the two extraction techniques are very similar, but the UAE seems to be the most 396 promising as it allows to reduce the time and energy costs derived from the use of high 397 temperatures. An LCA study will be described in the next paragraph to verify if the 398 UAE is the most sustainable extraction, even environmentally. 399 Table 2 Peleg's parameters for SE and UAE. Rate constant K<sub>1</sub>, constant capacity K<sub>2</sub>,

- 400 initial extraction rate ( $B_0$ ), maximum yield extraction ( $C_e$ ), regression coefficient ( $R^2$ ),
- 401 and the root mean square error (RMSE).

		$K_1$ [min $\cdot$ (g bran	$K_2$ [g bran $\cdot$	$B_0$ [g RBO $\cdot$ 100 /	C <sub>e</sub> [(g RBO /	g	
		/ g RBO) · 100 or	100 / g RBO or	min g bran or mg	bran) · 100	or R <sup>2</sup>	RMSD
		min · ( g bran /	g bran / mg γ-	γ-oryzanol / min g	mg γ-oryzar	ol	
		mg γ-oryzanol)]	oryzanol]	bran]	/gbran]		
SE	RBO	0,1506	0,1324	6,6401	7,5529	0,9302	0,0534
	γ-oryzanol	2,7668	2,1712	0,3614	0,4606	0,8919	0,7076
	RBO	0,0175	0,1577	57,1429	6,3412	0,7743	0,6070
UAE	γ-oryzanol	0,0442	3,2091	22,6244	0,3116	0,7947	0,0070

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### 403 **3.5 Life cycle assessment**

404 After evaluating the technical feasibility of oil extraction from rice bran, the

405 environmental sustainability of the different approaches was analyzed through LCA.

406 This analysis aimed to understand if to produce one gram of RBO, it is more

407 environmentally friendly to heat at 90°C for 15 minutes or to generate ultrasounds

408 (20kHz) for one minute at room temperature.

409 Fig. 5 shows the results of comparative LCA between SE and UAE in terms of four

410 impact categories: Climate change (kg CO<sub>2</sub> eq), Ozone depletion (kg CFC-11 eq),

411 Human toxicity (kg 1,4-DB eq), and Freshwater eutrophication (kg P eq). In each

412 impact category, the single contribution of the filtration steps, electricity, and

413 isopropanol to the emission and the total emission of all the two extraction procedures

414 are shown.

415 Regarding the Climate change impact category, to produce 1 gram of RBO, the SE and

416 UAE produce 0.206 and 0.156 kg of CO<sub>2</sub> eq, respectively; hence, the application of

417 UAE allows a reduction of the total impact of 25 %. In this impact category, the most

418 impacting step for both processes is filtration, which produces a considerable amount of

419 exhausted rice bran as waste material.

The SE process emits to the atmosphere  $1.95 \cdot 10^{-8}$  kg CFC-11 eq while the UAE process 420 emits only 1.46 · 10<sup>-8</sup> kg CFC-11 eq reducing the contribution to the Ozone depletion of 421 422 25 %. To evaluate if the difference among the two adopted techniques was statistically 423 significant a test of student was carried out considering p < 0.05. In this case, the most 424 significant contribution is due to electricity consumption, and the value is almost the 425 same in the two treatments. SE and UAE differ in the emissions in the filtration step 426 because the amount of RBO + isopropanol is higher in the second one, which produces 427 a minor quantity of exhausted rice bran.

428 Concerning the human toxicity impact category, no significant difference between SE

429 and UAE has been found, because the solvent employed was the same in both the430 extractions.

431 Regarding the freshwater eutrophication impact category, SE and UAE produce 7.13

 $432 cdot 10^{-5}$  and  $1.31 \cdot 10^{-5}$  kg P eq, respectively. Therefore, there is a massive reduction of the

433 impact of applying ultrasound instead of conventional extraction.

434 In all impact categories, the recycling of isopropanol originates avoided emissions

435 represented in Fig. 5 by negative bars.

436 For the best of author's knowledge, in the scientific literature LCA studies on oil

437 extraction from rice bran with SE and UAE techniques were not available.

438 To discuss the achieved results, the comparisons with other studies were performed

439 considering studies with cradle to gate approach, the product extracted as a functional

440 unit and midpoint as method to analyse the data coming from life cycle inventory.

441 Papadaki et al. [45] carried out an LCA study comparing SE, micro-waves, and

442 ultrasounds to recover the bioactive compounds from microalgae. The authors

443 demonstrated that among the three extraction techniques, ultrasound was the most

444 suitable one, since it reached the highest yielding, the lowest economic cost and

445 medium environmental impacts. Such results agree with the outputs of the present work.

446 Castro-Puyana et al., [46] performed bio-compounds extraction from rosemary plant by

447 means of green solvent and pressurized hot water extraction. The SE impacts obtained

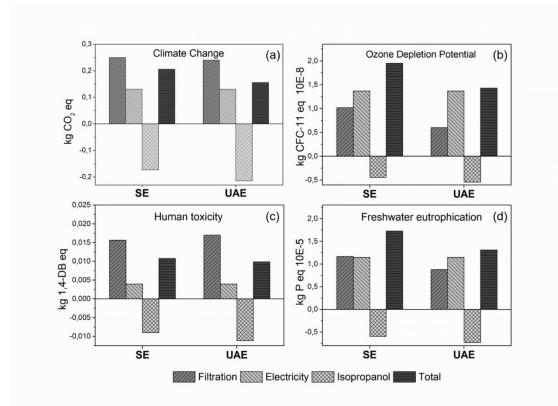
448 by the authors were in line with the impacts of SE technique achieved in the present

449 work.

450 Amiri et al. [47] carried out an LCA about alkaloids extracted from the Atropa

451 belladonna by methanol. The global warming potential (GWP) and ozone depletion

452 (OD) reported by the authors were equal to 0.899 kg CO<sub>2</sub> eq and to 0.00015 kg CFC-11 453 eq, respectively, whereas in the present work the GWP was 0.206 kg CO<sub>2</sub> eq and the OD was  $1.43 \cdot 10^{-8}$  kg CFC-11 eq. Hence, the present study reached potential impacts 454 455 lower than Amiri et al. [45] ones in a range between 77.00%-99.98 %. 456 Barjovenu et al. [48][48] performed an LCA study on polyphenols extraction from 457 spruce bark, by means of SE using ethanol and UAE. The difference between SE using 458 ethanol and UAE was 70.00%, whereas in the present study was 24%. The main 459 difference with the present work is due to the different solvent used. However, the UAE 460 technique resulted in an environmental impact lower than SE. Thus, the present work 461 proved that UAE technique is both technically more efficient and environmentally more 462 sustainable than SE technique. The present study proved the technical feasibility of the 463 two proposed techniques SE and UAE and the feasible scale up, furthermore the LCA 464 study, performed considering the data at laboratory scale, underscored the bottleneck of 465 the processes, which are the filtration steps and the energy consume for both the 466 techniques. Hence, the recommendation and future prospective are the minimisation of 467 waste production at filtration step improving the technique and the optimisation of the 468 energy consume by doing a proper design of the plant.



470 Fig. 5 Comparative LCA results between SE and UAE. Four main impact categories are 471 illustrated: Climate change, Ozone depletion, Human toxicity and Freshwater 472 eutrophication. Each graph reports the result for SE (on the left) and for UAE (on the 473 right); for each extraction process, the contribution of filtration, electricity, isopropanol 474 and their sum (total) are reported.

475

### 476 **4. Conclusions**

477 In the present work, the isopropanol SE of RBO was optimized in terms of temperature

478 and bran to solvent ratio. The best RBO yield was obtained at 90°C and 1:9 bran to

479 solvent ratio. The results were compared with the RBO yield obtained in a standard

480 hexane extraction, demonstrating that isopropanol is suitable to RBO extraction, making

481 the substitution of organic and toxic solvents possible. The kinetics of isopropanol SE at

482 the best-operating conditions was evaluated and compared with a room temperature 483 UAE using a 1:9 bran to solvent ratio. The two extraction techniques produced similar 484 yields in terms of RBO and  $\gamma$ -oryzanol, but UAE reduced remarkably the extraction 485 time. A comparative LCA between the two extraction techniques showed that UAE allows lower the emission contribution to climate change, ozone depletion, and 486 487 freshwater eutrophication compared to SE, to produce 1 gram of RBO generating high 488 yield, operating at room temperature in a very short time, in line with the principles of 489 green chemistry. To the best of the authors' knowledge, this paper shows for the first 490 time a comparison between the extraction of RBO with isopropanol at 90°C and with 491 isopropanol at room temperature assisted by ultrasounds both from a technical and 492 environmental point of view. The evaluation carried out and the results obtained can be 493 the basis for new experimental campaigns or to design a scale-up RBO production 494 plant.

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#### 496 **5. References**

TZ 1

497	1.	Kraehmer H, Thomas C, Vidotto F (2017) Rice production in Europe
498	2.	Fraterrigo Garofalo S, Tommasi T, Fino D (2020) A short review of green
499		extraction technologies for rice bran oil. Biomass Convers Biorefinery.
500		https://doi.org/10.1007/s13399-020-00846-3
501	3.	García A, De Lucas A, Rincón J, et al (1996) Supercritical carbon dioxide
502		extraction of fatty and waxy material from rice bran. JAOCS, J Am Oil Chem
503		Soc 73:1127–1131. https://doi.org/10.1007/BF02523373

O X' I + F (0017) D'

...

504 4. Dhankhar P (2014) Rice Milling. IOSR J Eng 4:34–42.

https://doi.org/10.9790/3021-04543442

- 506 5. Sharif MK, Butt MS, Anjum FM, Khan SH (2014) Rice Bran: A Novel
- 507 Functional Ingredient. Crit Rev Food Sci Nutr 54:807–816.
- 508 https://doi.org/10.1080/10408398.2011.608586
- 509 6. Rohman A (2014) Rice Bran Oil's Role in Health and Cooking. Elsevier
- 510 7. Lai O-M, Jacoby JJ, Leong W-F, Lai W-T (2019) Nutritional Studies of Rice
  511 Bran Oil
- 5128.Wang Y (2019) Applications of Rice Bran Oil. Rice Bran Rice Bran Oil 159–
- 513 168. https://doi.org/10.1016/b978-0-12-812828-2.00006-8
- 514 9. B. Ghatak S, J. Panchal S (2011) Gamma-Oryzanol A Multi-Purpose Steryl
  515 Ferulate. Curr Nutr Food Sci 7:10–20.
- 516 https://doi.org/10.2174/157340111794941120
- 517 10. Zia S, Khan MR, Shabbir MA, et al (2020) An Inclusive Overview of Advanced
- 518 Thermal and Nonthermal Extraction Techniques for Bioactive Compounds in
- 519 Food and Food-related Matrices. Food Rev Int 00:1–31.
- 520 https://doi.org/10.1080/87559129.2020.1772283
- 521 11. Garba U, Singanusong R, Jiamyangyeun S, Thongsook T (2019) Extraction and
- 522 utilisation of rice bran oil. A review. Riv Ital delle Sostanze Grasse 96:161–170
- 523 12. Hu W, Wells JH, Shin TS, Godber JS (1996) Comparison of isopropanol and
- 524 hexane for extraction of vitamin E and oryzanols from stabilized rice bran.
- 525 JAOCS, J Am Oil Chem Soc 73:1653–1656.
- 526 https://doi.org/10.1007/BF02517967

527	13.	Mushtaq A, Roobab U, Denoya GI, et al (2020) Advances in green processing of
528		seed oils using ultrasound-assisted extraction: A review. J Food Process Preserv
529		44:. https://doi.org/10.1111/jfpp.14740
530	14.	Tiwari BK (2015) Ultrasound: A clean, green extraction technology. TrAC -
531		Trends Anal Chem 71:100–109. https://doi.org/10.1016/j.trac.2015.04.013
532	15.	Pengkumsri N, Chaiyasut C, Sivamaruthi BS, et al (2015) The influence of
533		extraction methods on composition and antioxidant properties of rice bran oil.
534		Food Sci Technol 35:493-501. https://doi.org/10.1590/1678-457X.6730
535	16.	Yoshie A, Kanda A, Nakamura T, et al (2009) Comparison of $\gamma$ -oryzanol
536		contents in crude rice bran oils from different sources by various determination
537		methods. J Oleo Sci 58:511-518. https://doi.org/10.5650/jos.58.511
538	17.	Hung CC, Weng YM, Yu ZR, Wang BJ (2019) Optimal selectivity of $\gamma$ -oryzanol
539		and total phenolic compounds from rice bran using supercritical carbon dioxide
540		fractionation technique. Int Food Res J 26:639-647
541	18.	Guy RC (2014) International Conference on Harmonisation. Encycl Toxicol
542		Third Ed 2:1070–1072. https://doi.org/10.1016/B978-0-12-386454-3.00861-7
543	19.	Peleg M (1988) An Empirical Model for the Prediction. J Food Sci 53:1216–
544		1217
545	20.	Poojary MM, Passamonti P (2015) Extraction of lycopene from tomato
546		processing waste: Kinetics and modelling. Food Chem 173:943–950.
547		https://doi.org/10.1016/j.foodchem.2014.10.127
548	21.	Xu Z, Godber JS (2000) Comparison of supercritical fluid and solvent extraction

549		methods in extracting $\gamma$ -oryzanol from rice bran. JAOCS, J Am Oil Chem Soc
550		77:547-551. https://doi.org/10.1007/s11746-000-0087-4
551	22.	Oliveira R, Oliveira V, Aracava KK, Rodrigues CEDC (2012) Effects of the
552		extraction conditions on the yield and composition of rice bran oil extracted with
553		ethanol - A response surface approach. Food Bioprod Process 90:22-31.
554		https://doi.org/10.1016/j.fbp.2011.01.004
555	23.	Capellini MC, Giacomini V, Cuevas MS, Rodrigues CEC (2017) Rice bran oil
556		extraction using alcoholic solvents: Physicochemical characterization of oil and
557		protein fraction functionality. Ind Crops Prod 104:133-143.
558		https://doi.org/10.1016/j.indcrop.2017.04.017
559	24.	Ruen-Ngam D, Thawai C, Nokkoul R, Sukonthamut S (2014) Gamma-Oryzanol
560		Extraction from Upland Rice Bran. Int J Biosci Biochem Bioinforma 4:252–255.
561		https://doi.org/10.7763/ijbbb.2014.v4.350
562	25.	Aliwarga L (2019) Investigating Mass Transfer Phenomena in Batch Solvent
563		Extraction of Rice Bran Oil. Reaktor 19:1–10.
564		https://doi.org/10.14710/reaktor.19.1.1-10
565	26.	Mas F, Bangngalino H, Indriati S, et al (2019) oil
566	27.	Wong BY, Tan CP, Ho CW (2013) Effect of solid-to-solvent ratio on phenolic
567		content and antioxidant capacities of "Dukung Anak" (Phyllanthus niruri). Int
568		Food Res J 20:325–330
569	28.	Pinelo M, Rubilar M, Jerez M, et al (2005) Effect of solvent, temperature, and
570		solvent-to-solid ratio on the total phenolic content and antiradical activity of

571		extracts from different components of grape pomace. J Agric Food Chem
572		53:2111–2117. https://doi.org/10.1021/jf0488110
573	29.	Elboughdiri N (2018) Effect of Time, Solvent-Solid Ratio, Ethanol Concentration
574		and Temperature on Extraction Yield of Phenolic Compounds From Olive
575		Leaves. Eng Technol Appl Sci Res 8:2805–2808.
576		https://doi.org/10.48084/etasr.1983
577	30.	Chanioti S, Liadakis G, Tzia C (2014) Solid–Liquid Extraction. 253–286.
578		https://doi.org/10.1201/b17803-7
579	31.	García-Márquez E, Román-Guerrero A, Pérez-Alonso C, et al (2012) Effect of
580		solvent-temperature extraction conditions on the initial antioxidant activity and
581		total phenolic content of muitle extracts and their decay upon storage at different
582		pH. Rev Mex Ing Quim 11:1–10
583	32.	Spigno G, Tramelli L, De Faveri DM (2007) Effects of extraction time,
584		temperature and solvent on concentration and antioxidant activity of grape marc
585		phenolics. J Food Eng 81:200–208.
586		https://doi.org/10.1016/j.jfoodeng.2006.10.021
587	33.	Bucić-Kojić A, Planinić M, Tomas S, et al (2007) Study of solid-liquid extraction
588		kinetics of total polyphenols from grape seeds. J Food Eng 81:236–242.
589		https://doi.org/10.1016/j.jfoodeng.2006.10.027
590	34.	Capellini MC, Giacomini V, Cuevas MS, Rodrigues CEC (2017) Rice bran oil
591		extraction using alcoholic solvents: Physicochemical characterization of oil and
592		protein fraction functionality. Ind Crops Prod 104:133-143.
593		https://doi.org/10.1016/j.indcrop.2017.04.017

594	35.	Chan CH, Yusoff R, Ngoh GC (2014) Modeling and kinetics study of
595		conventional and assisted batch solvent extraction. Chem Eng Res Des 92:1169-
596		1186. https://doi.org/10.1016/j.cherd.2013.10.001
597	36.	Tan BL, Norhaizan ME (2020) Rice By-products: Phytochemicals and Food
598		Products Application
599	37.	Shukla HS, Pratap A (2017) Comparative studies between conventional and
600		microwave assisted extraction for rice bran oil. J Oleo Sci 66:973–979.
601		https://doi.org/10.5650/jos.ess17067
602	38.	Karacabey E, Bayindirli L, Artik N, Mazza G (2013) Modeling solid-liquid
603		extraction kinetics of trans-resveratrol and trans- $\epsilon$ -viniferin from grape cane. J
604		Food Process Eng 36:103-112. https://doi.org/10.1111/j.1745-
605		4530.2011.00660.x
606	39.	Jurinjak Tušek A, Benković M, Belščak Cvitanović A, et al (2016) Kinetics and
607		thermodynamics of the solid-liquid extraction process of total polyphenols,
608		antioxidants and extraction yield from Asteraceae plants. Ind Crops Prod 91:205-
609		214. https://doi.org/10.1016/j.indcrop.2016.07.015
610	40.	Cravotto G, Binello A, Merizzi G, Avogadro M (2004) Improving solvent-free
611		extraction of policosanol from rice bran by high-intensity ultrasound treatment.
612		Eur J Lipid Sci Technol 106:147–151. https://doi.org/10.1002/ejlt.200300914
613	41.	Khoei M, Chekin F (2016) The ultrasound-assisted aqueous extraction of rice
614		bran oil. Food Chem 194:503–507.
615		https://doi.org/10.1016/j.foodchem.2015.08.068

616	42.	Tabaraki R, Nateghi A (2011) Optimization of ultrasonic-assisted extraction of
617		natural antioxidants from rice bran using response surface methodology. Ultrason
618		Sonochem 18:1279–1286. https://doi.org/10.1016/j.ultsonch.2011.05.004
619	43.	Kumar P, Yadav D, Kumar P, et al (2016) Comparative study on conventional,
620		ultrasonication and microwave assisted extraction of $\gamma$ -oryzanol from rice bran. J
621		Food Sci Technol 53:2047–2053. https://doi.org/10.1007/s13197-016-2175-2
622	44.	Khoei M, Chekin F (2016) The ultrasound-assisted aqueous extraction of rice
623		bran oil. FOOD Chem 194:503–507.
624		https://doi.org/10.1016/j.foodchem.2015.08.068
625	45.	Papadaki SG, Kyriakopoulou KE, Krokida MK (2016) Life cycle analysis of
626		microalgae extraction techniques. Chem Eng Trans 52:1039–1044.
627		https://doi.org/10.3303/CET1652174
628	46.	Castro-puyana M, Mendiola J a, Rodríguez-meizoso I, et al (2009) Comparative
629		Life Cycle Assessment Study of Green Extraction Processes to Obtain
630		Antioxidants from Rosemary Leaves . J Supercrit Fluids 72:13-17
631	47.	Amiri M, Arabhosseini A, Kianmehr MH, et al (2017) Environmental impact
632		assessment of total alkaloid extracted from the Atropa belladonna L . using LCA
633		. Geol Ecol Landscapes 1:257–263.
634		https://doi.org/10.1080/24749508.2017.1389502
635	48.	Barjoveanu G, Pătrăuțanu OA, Teodosiu C, Volf I (2020) Life cycle assessment
636		of polyphenols extraction processes from waste biomass. Sci Rep 10:1-12.
637		https://doi.org/10.1038/s41598-020-70587-w

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