

Systematic Review **Drug-Induced Myoclonus: A Systematic Review**

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Abstract: *Background and Objectives*: Myoclonus is already associated with a wide variety of drugs and systemic conditions. As new components are discovered, more drugs are suspected of causing this disabling abnormal involuntary movement. This systematic review aims to assess the medications associated with drug-induced myoclonus (DIM). *Materials and Methods*: Two reviewers assessed the PubMed database using the search term "myoclonus", without language restriction, for articles published between 1955 and 2024. The medications found were divided into classes and sub-classes, and the subclasses were graded according to their level of evidence. *Results*: From 12,097 results, 1115 were found to be DIM. The subclasses of medications with level A evidence were intravenous anesthetics (etomidate), cephalosporins (ceftazidime, cefepime), fluoroquinolones (ciprofloxacin), selective serotonin reuptake inhibitors (citalopram, escitalopram, paroxetine, sertraline), tricyclic antidepressant (amitriptyline), glutamate antagonist (amantadine), atypical antipsychotics (clozapine, quetiapine), antiseizure medications (carbamazepine, oxcarbazepine, phenytoin, gabapentin, pregabalin, valproate), pure opioid agonist (fentanyl, morphine), bismuth salts, and mood stabilizers (lithium). The single medication with the highest number of reports was etomidate. Drug-induced asterixis is associated with a specific list of medications. The neurotransmitters likely involved in DIM are serotonin, dopamine, gamma-aminobutyric acid (GABA), and glutamate. *Conclusions*: DIM may be reversible with management that can include drug discontinuation, dose adjustment, and the prescription of a medication used to treat idiopathic myoclonus. Based on the main clinical constellation of symptoms and pathophysiological mechanisms found in this study, DIM can be categorized into three types: type 1 (serotonin syndrome), type 2 (non-serotonin syndrome), and type 3 (unknown).

Keywords: myoclonus; neurotoxicity; encephalopathy; drug-induced myoclonus; adverse effect; myoclonus/chemically induced; antidepressant-induced myoclonus; opioid-induced myoclonus; anti-seizure medication-induced myoclonus; antibiotic-induced myoclonus

1. Introduction

Myoclonus is a hyperkinetic movement disorder characterized by sudden, brief, shock-like involuntary movements [\[1\]](#page-21-0). When there is a sudden muscular contraction, it is called a positive myoclonus; on the other hand, when a cessation of the ongoing muscular contraction occurs, it is called a negative myoclonus or asterixis [\[2\]](#page-21-1). The term "myoclonus"

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is derived from "paramyoklonus multiplex", which was coined by Nikolaus Friedreich in a 50-year-old male patient while describing involuntary movements at rest [\[3\]](#page-21-2).

Many classifications have been developed to categorize myoclonus. For example, myoclonus can be classified based on its source, such as cortical, subcortical, cortical– subcortical, subcortical–nonsegmental, and spinal (segmental, propriospinal, and peripheral) [\[4\]](#page-21-3). Also, it can be categorized based on its clinical distribution into focal, segmental, multifocal, and generalized [\[5\]](#page-21-4). The etiological classification of myoclonus is crucial because it can assist in clinical decisions regarding the pharmacological management of this movement disorder [\[6\]](#page-21-5).

Myoclonus can be primary or secondary to infectious, metabolic, endocrine pathology, degenerative, inflammatory, toxic, genetic, and pharmacological causes [\[7\]](#page-21-6). Many examples of drug-induced myoclonus (DIM) are transient and occur in the setting of diffuse encephalopathy associated with drug toxicity, acute metabolic abnormalities, or infectious disorders [\[8\]](#page-21-7). In most cases, conducting a detailed investigation with neurophysiological studies is not practical.

The epidemiological data on DIM are scarce. A French pharmacovigilance database study revealed that the incidence of DIM is around 0.2% in the general population; the most frequent culprit drugs were antibiotics, antidepressants, anxiolytics, and opiate agents. Also, the authors reported that the median age of patients was 55 years, and 10% of these patients had a concomitant neurological disease [\[9\]](#page-21-8). Caviness et al. reported in Olmsted County, from 1976 to 1990, a lifetime prevalence of 8.6 cases per 100,000 people, while the annual incidence rate was 1.3 cases per 100,000 person-years [\[10\]](#page-21-9). Thwaites et al. showed no age and sex predilection for hydromorphone-induced myoclonus in terminally ill hospice patients [\[11\]](#page-21-10). Janssen et al. found that the most common classes of medications associated with myoclonus were opiates, antidepressants, antipsychotics, and antibiotics [\[5\]](#page-21-4).

More research on this subject is needed because myoclonus can impact patients and caregivers, causing increased morbidity and a higher economic burden for the healthcare system. In this systematic review, we will discuss myoclonus secondary to drugs, providing a list of drugs along with the level of evidence of their association with myoclonus. We highlight the importance of considering myoclonus as a potential side effect of various drugs, even at low doses, to avoid costly and unnecessary investigations, as well as excessive treatments.

2. Methodology

We searched the Medline/PubMed database to locate all existing reports on myoclonus secondary to medications published from January 1955 to June 2024 in electronic form. The search term was "myoclonus". The query was ("myoclonus"[MeSH Terms] OR "myoclonus"[All Fields]). For a complete list of all the publications concerning DIM in PubMed, consider reading Table S1. The Preferred Reporting Items for Systematic reviews and Meta-Analyses (PRISMA) statement published in 2020 was used for the literature search and methodology [\[12\]](#page-21-11).

The inclusion criteria covered case reports, case series, original articles, letters to the editor, bulletins, and poster presentations, published from January 1955 to June 2024, without language restriction, to ensure a thorough review. When the non-English literature was beyond the authors' proficiency (English, French, German, and Spanish) or the English abstract did not provide enough data, such as articles in Danish, Japanese, and Polish, the Google Translate service was used [\[13\]](#page-21-12).

The authors independently screened the titles and abstracts of all articles from the initial search. Disagreements between authors were solved through discussion. Cases not accessible by electronic methods, including after a formal request to the authors, were excluded. Cases with more than one factor contributing to the myoclonus were evaluated based on the probability of the event occurring, using the Naranjo algorithm. based on the probability of the event occurring, using the Naranjo algorithm.

From 12,097 results, 1115 were found to be DIM (Figure [1\)](#page-2-0). Data abstraction was From 12,097 results, 1115 were found to be DIM (Figure 1). Data abstraction was performed. The authors read the title and the abstract of all the articles found in the initial performed. The authors read the title and the abstract of all the articles found in the initial search. When provided, we extracted the PubMed identifier (PMID), the main cause of myoclonus, and the year of publication. The data were extracted by two independent myoclonus, and the year of publication. The data were extracted by two independent authors and double-checked to ensure that they matched. authors and double-checked to ensure that they matched.

Figure 1. Prisma flowchart of the screening process. Figure 1. PRISMA flowchart of the screening process.

to their PubMed identifier (PMID) number. No statistical analysis was performed. The articles are ordered in Table S1 according

The clinical definition of myoclonus was obtained from Caviness et al. [\[6\]](#page-21-5). The Naranjo algorithm was used to determine the likelihood of whether an adverse drug reaction was actually due to the drug rather than the result of other factors [\[14\]](#page-21-13). After the literature review, a grading system was developed based on the quantity of patients with DIM and the quality of the published manuscripts. An "A" level of evidence was defined when there were more than 20 individuals reported to have myoclonus caused by that specific class of medications. "B" was characterized by 5 and 20 individuals reported with DIM. "C" was defined by less than five individuals reported to have myoclonus. When there was no case report and only animal studies, the subclass of drugs was graded as "D". \mathbf{t} there was no case report and only animal studies, the subclass was graded as \mathbf{t}

3. Results

The classes of drugs were categorized as anesthetics, antibiotics, antidementia, antidepressants, antiemetics, antihemorrhagic, antihistamines, antineoplastic and immunosuppressive agents, antiparasitic, antiparkinsonian, antipsychotics, antiseizure, antiviral, anxiolytics, cardiovascular, opioids, and others. We also included a specific category regarding the drugs related to animal models of myoclonus. Table [1](#page-5-0) is a summary of the classes of medications already associated with myoclonus and their level of evidence.

Table 1. Drug-induced myoclonus and level of evidence.

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Table 1. *Cont.*

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* Grading level of the evidence: A, more than 20 individuals have been reported to have myoclonus; B, between 5 and 20 individuals have been reported to have myoclonus; C, less than 5 individuals have been reported to have myoclonus; D, only animal studies reporting myoclonus.

4. Anesthetic Agents

Enflurane [\[15\]](#page-21-14), etomidate [\[16\]](#page-21-15), nitrous oxide [\[17\]](#page-21-16), and propofol [\[18\]](#page-21-17) are some of the anesthetic agents associated with myoclonus. Deep anesthesia and muscle relaxants may attenuate myoclonus [\[19\]](#page-21-18). However, it is worth mentioning that an association between local, spinal anesthesia and myoclonus has already been reported [\[20\]](#page-21-19). For anesthetics associated with myoclonus in the literature, consider reading Table S2 [\[15](#page-21-14)[–18,](#page-21-17)[21–](#page-21-20)[30\]](#page-22-0).

In an experimental study, enflurane and isoflurane, compared to halothane, were more commonly associated with myoclonus in mild hypocapnic cats and provoked greater airway inflammation [\[31\]](#page-22-1). Ng et al. reported a case of enflurane-induced myoclonus involving multiple muscle groups, except those innervated by the cranial nerves [\[15\]](#page-21-14). Interestingly, nitrous oxide-induced myoclonus was already related to a propriospinal source due to subacute spinal cord degeneration associated with cobalamin deficiency [\[32\]](#page-22-2). A similar hypothesis can be assumed for cases of myoclonus induced by cobalamin supplementation [\[33\]](#page-22-3).

Propofol is a short-acting gamma-aminobutyric acid receptor (GABA-A) agonist that can safely be used as a general anesthetic. Propofol is already associated with causing [\[18\]](#page-21-17) and improving [\[34\]](#page-22-4) myoclonus. Walder et al. found that propofol-induced seizure-like phenomena are most common during induction, emergence, or the immediate postoperative periods [\[35\]](#page-22-5). Chao et al. proposed that propofol-induced myoclonus occurs due to cortical reflex myoclonus [\[36\]](#page-22-6).

Ketamine-induced myoclonus was rarely described in the literature. There are only studies with veterinary anesthesia in which dogs showed myoclonus [\[25\]](#page-22-7). However, no myoclonus was observed in humans, suggesting something specific to the metabolism of this medication in some dog breeds, especially the English Greyhound [\[37\]](#page-22-8).

Etomidate was the most common medication reported to cause myoclonus in the literature. Several articles published were clinical trials assessing the possible management

Table 1. *Cont.*

of etomidate-induced myoclonus. Doenicke et al. reported that the incidence and intensity of myoclonus after induction with etomidate are dose-related, suppressed by pre-treatment, and unassociated with seizure-like EEG activity [\[38\]](#page-22-9). Etomidate-induced myoclonus likely involves neocortical glutamate accumulation and N-methyl-D-aspartate receptor (NMDAR) modulation activity. Myoclonus was correlated with the NMDAR-induced downregulation of potassium-chloride transporter member 5 (KCC2) protein expression [\[39\]](#page-22-10). Therefore, suppressing the astrogliosis in the neocortex and promoting extracellular glutamate uptake by regulating glutamate transporters in the motor cortex may be a therapeutic option for managing myoclonus associated with etomidate [\[40\]](#page-22-11).

Zhou et al. showed that the incidence of etomidate-induced myoclonus was significantly lower in midazolam-treated groups (RR = 0.34, 95% CI [0.26, 0.44], *p* < 0.05). The authors revealed that subgroups divided by the degree of myoclonus showed a significantly lower incidence of mild myoclonus (RR = 0.56, 95% CI [0.39, 0.80], *p* < 0.05), moderate myoclonus (RR = 0.20, 95% CI [0.10, 0.41], *p* < 0.05), and severe myoclonus (RR = 0.12, 95% CI [0.04, 0.39], *p* < 0.05) [\[41\]](#page-22-12). Nooraei et al. showed that using a priming dose of atracurium efficiently suppresses etomidate-induced myoclonus during the induction of anesthesia. The adjusted odds ratio in this model of myoclonus in the control group was 6.6 (95% Cl [1.5–9.7], $p < 0.05$] [\[42\]](#page-22-13). Collin et al. reported that alfentanil significantly reduced myoclonus associated with etomidate [\[43\]](#page-22-14). Du et al. found that etomidate-induced myoclonus in the dexmedetomidine-treated groups was significantly lower than that in the control groups (RR = 0.27, 95% CI [0.15, 0.47], *p* < 0.00001) [\[44\]](#page-22-15). Feng et al. reported that etomidate-induced myoclonus in the propofol-treated groups was significantly lower than that in the control groups (RR = 2.99, 95% CI [2.40, 3.71], *p* < 0.0001) [\[34\]](#page-22-4). Hua et al. reported that etomidate-induced myoclonus in the butorphanol-treated groups was significantly lower than that in the control groups (RR = 0.15, 95% CI [0.10, 0.22], *p* < 0.00001) [\[45\]](#page-22-16). Zhu et al. showed that the pre-injection of dezocine (opioid analgesic) can reduce the incidence of etomidate-induced myoclonus (RR = 0.25, 95% CI [0.13, 0.50], *p* < 0.0001) [\[46\]](#page-22-17). Finally, Greenwood et al. found an absolute reduction in risk with prophylactic medications, ranging from 47% to 81% for mild, 52% to 92% for moderate, and 61% to 96% for severe myoclonus. Also, the authors observed that opioids have a consistent and substantial effect on the reduction in myoclonus [\[47\]](#page-22-18). Further studies assessing all the therapeutic options already proposed for etomidate-induced myoclonus are mandatory. Studying this specific cause of myoclonus can be used to understand DIM pathophysiology and further improve current management strategies.

Spinal myoclonus following neuraxial anesthesia was rarely reported in the literature. Shiratori et al. found 23 cases of spinal myoclonus associated with local anesthetics. In total, 82.6% of the cases occurred following lumbar spinal anesthesia, and the rest following epidural anesthesia. Amide-type local anesthetics, such as dibucaine, lidocaine, bupivacaine, prilocaine, and levobupivacaine, were used in 95.7% of the cases, whereas an ester-type local anesthetic, such as tetracaine, was used in only one case [\[48\]](#page-23-0).

5. Antibiotics

Stimulus-sensitive myoclonus and encephalopathy have already been associated with cephalosporins [\[49\]](#page-23-1), carbenicillin [\[50\]](#page-23-2), imipenem [\[51\]](#page-23-3), quinolones [\[52\]](#page-23-4), penicillin [\[53\]](#page-23-5), piperacillin [\[54\]](#page-23-6), and ticarcillin [\[55\]](#page-23-7). For antibiotic medications associated with myoclonus in the literature, consider reading Table S3 [\[50,](#page-23-2)[54,](#page-23-6)[56](#page-23-8)[–86\]](#page-24-0).

5.1. Penicillins, Cephalosporins

Antibiotics like penicillin and cephalosporin have been shown to cause myoclonic jerks, which can be generalized, multifocal, or segmental [\[87\]](#page-24-1). Myoclonus is commonly

accompanied by other symptoms, such as an altered mental state, seizures, aphasia, chorea, and a skin rash [\[5\]](#page-21-4). Bhattacharyya et al. systematically reviewed encephalopathy secondary to antibiotics and found three clinical phenotypes. Type 1 was characterized by onset within days of antibiotic initiation, the occurrence of myoclonus or seizures, and resolution within days. Type 2 was marked by onset within days of antibiotic initiation, the frequent occurrence of psychosis, and the rare occurrence of seizures. Type 3, seen only with metronidazole, is characterized by the onset of encephalopathy within weeks after the initiation of metronidazole, the frequent occurrence of cerebellar dysfunction, and rare seizures [\[88\]](#page-24-2).

Conway et al. reported five cases of endocarditis in which high doses of penicillin resulted in neurotoxicity in the form of drowsiness and myoclonus. The adverse effect was unrelated to the type of penicillin preparation (sodium or potassium) used [\[89\]](#page-24-3). Interestingly, other penicillin formulations or penicillin itself provoked the same neurotoxicity manifestations [\[90\]](#page-24-4). Also, Lerner et al. found higher concentrations of penicillin in the cerebrospinal fluid (CSF), which they attributed to the altered blood–brain barrier (BBB) permeability caused by uremia and the deterioration of renal function, increasing the toxic levels [\[90\]](#page-24-4). Penicillins are excreted by the kidneys into the urine in patients with normal renal function. Hence, the penicillin dose must be adjusted according to the renal function of the patient to prevent side effects [\[91\]](#page-24-5). Of note, some authors reported the occurrence of myoclonus with penicillins and the association between cephalosporins and renal clearance impairment [\[92\]](#page-24-6), but this should be evaluated with caution because end-stage renal disease is also associated with myoclonus [\[93\]](#page-24-7).

The cephalosporins most commonly associated with myoclonus were ceftazidime and cefepime. One of the mechanisms causing cephalosporin neurotoxicity is the induction of endotoxins and, possibly, glutaminergic mechanisms. Laboratory studies also show that cephalosporins have a high affinity for GABA-A receptors, which cause high penetrance through the BBB and are more neurotoxic [\[94\]](#page-24-8). Studies show that in patients with renal disease, the maintenance dose should be reduced, and the patients should be monitored for neurotoxicity. Care should be taken when using cefepime use and its toxicity should be kept in mind whenever a patient receiving it shows a change in mental status or myoclonus [\[95\]](#page-24-9). Chow et al. retrospectively studied 42 cases of cefepime and 12 cases of ceftazidime-induced neurotoxicity with myoclonus. In total, 33% of the individuals were already uremic, which led to a delay in diagnosis due to this confounding factor. The median time between the onset of symptoms and diagnosis for cefepime-induced encephalopathy compared to ceftazidime-induced neurotoxicity was 5 and 3 days, respectively $[p = 0.005]$ [\[96\]](#page-24-10).

5.2. Fluoroquinolones and Quinolones

Myoclonus is commonly associated with fluoroquinolones, especially ciprofloxacin. Some authors have called this association "ciproclonus" [\[74\]](#page-23-9). Ciprofloxacin is also associated with propriospinal myoclonus by antagonizing gamma-aminobutyric acid metabolism [\[73\]](#page-23-10). Another case reported that myoclonus is associated with delirium, and the authors highlighted the importance of ciprofloxacin as a cause of delirium and myoclonus in elderly patients [\[72\]](#page-23-11). Also, consideration should be given to prescribing reduced-dose ciprofloxacin to elderly patients with renal impairment. Rissardo et al. reported that the fluoroquinolone-induced myoclonus distribution is focal, multifocal, segmental, axial, and generalized. Myoclonus was the most common movement disorder associated with fluoroquinolones, and the authors found 25 cases in the literature. Finally, the authors described that previously reported fluoroquinolone-associated myoclonus was likely related to an increased glutamate concentration due to the neurotoxic effects of fluoroquinolones, including oxidative process, chelated cations, and disturbed gene expression [\[97\]](#page-24-11).

5.3. Other Classes of Antibiotics

Carbapenems are commonly associated with seizures, which can also explain the high number of reports of myoclonus. This can suggest a cortical source for the myoclonus secondary to carbapenems. Cannon et al. systematically reviewed the literature regarding carbapenems and their risk of seizure, and the authors found that imipenem was more epileptogenic than non-carbapenem antibiotics. But there was no statistically significant difference between the imipenem and the meropenem [\[98\]](#page-24-12). Noteworthy, after cephalosporins and fluoroquinolones, carbapenem and penicillins were the most common classes of antibiotic-induced myoclonus.

Macrolides such as erythromycin and azithromycin were rarely reported to be associated with myoclonus. It is worth mentioning that there are some rare cases of azithromycininduced myoclonus. Still, these cases can also be explained by encephalitis lethargica, commonly associated with some viral and bacterial pathogens [\[80\]](#page-24-13).

Linezolid-induced myoclonus was eventually described in the literature [\[86\]](#page-24-0). The role of linezolid as a weak, non-selective, reversible monoamine oxidase inhibitor can explain the reasonable number of articles about myoclonus. There have been post-marketing reports of serotonin syndrome when linezolid was given with or soon after the discontinuation of serotonergic drugs [\[99\]](#page-24-14).

Gentamicin-induced multifocal myoclonus was observed in elderly individuals with renal impairment, and a favorable prognosis on gentamicin discontinuation was noticed [\[81\]](#page-24-15). Also, myoclonus was reported with anthelmintics and antituberculosis drugs such as piperazine [\[100\]](#page-24-16) and isoniazid [\[101\]](#page-24-17), respectively. Other classes of antibiotics were rarely associated with myoclonus, such as lipopeptides [\[83\]](#page-24-18), glycopeptides [\[84\]](#page-24-19), and tetracyclines [\[85\]](#page-24-20).

6. Antidementia

Myoclonus secondary to memantine was observed in patients with dementia [\[102\]](#page-24-21). Pei et al. found five cases in the literature of memantine-induced myoclonus. The authors reported that the onset of myoclonus after memantine exposure ranged from 6 days to 2 months, with the complete resolution of myoclonus upon the cessation of memantine [\[103\]](#page-24-22). The mechanism underlying memantine-induced myoclonus remains unclear but might involve altered dopamine, serotonin, and glutamate levels. Noteworthy, memantine is an amino-adamantane chemically similar to amantadine [\[104\]](#page-24-23).

Rissardo et al. reported a case of action myoclonus secondary to donepezil. The authors also found six other reports of myoclonus secondary to donepezil/galantamine, but no report of rivastigmine-induced myoclonus was identified. Also, they observed that the most frequent presentation was multifocal myoclonus [\[105\]](#page-25-0). In experimental studies, reducing acetylcholine function improved picrotoxin-induced myoclonus [\[106\]](#page-25-1). Therefore, high levels of acetylcholine may be associated with the frequency of involuntary twitching. Noteworthy, myoclonus is common in Alzheimer's disease and vascular dementia and can also be present in Lewy body dementia, although typically in the later stages of the disease [\[107\]](#page-25-2).

7. Antidepressants

During the last decade, 13.2% of adults have used at least one antidepressant in the past 30 days, which is higher among females (17.7%) than males (8.4%) [\[108\]](#page-25-3). Also, the prescription of antidepressants, especially SSRIs and SNRIs, has increased over the last few years due to the COVID-19 pandemic [\[109\]](#page-25-4). In 2006, The US Food and Drug Administration (FDA) issued an advisory about the risk of serotonin syndrome (which has myoclonus as a component) associated with the concomitant use of drugs from two

widely prescribed medication classes of this family, namely selective serotonin reuptake inhibitor (SSRI) and selective norepinephrine reuptake inhibitor (SNRI). Interestingly, among the antidepressants, SSRIs [\[110\]](#page-25-5) and tricyclic antidepressants (TCA) [\[111\]](#page-25-6) were the most common classes associated with myoclonus. For antidepressant medications associated with myoclonus in the literature, consider reading Table S4 [\[111](#page-25-6)[–135\]](#page-26-0).

A 62-year-old woman with a history of congestive heart failure and coronary artery disease, mitral valve stenosis, and chronic kidney disease received buspirone and, within a day, developed dramatic myoclonus, which did not improve with intramuscular diphenhydramine. However, the myoclonus completely improved after clonazepam administration [\[136\]](#page-26-1). Riaz et al. found only eight cases of bupropion-induced myoclonus, in which the most common association with myoclonus in younger individuals was overdose and intoxication. Bupropion has higher rates of seizures compared to other antidepressants, which can occur even in the presence of benzodiazepines. The authors hypothesized that the cause of seizures could be related to the antagonization of nicotinic acetylcholine receptors [\[122\]](#page-25-7). Also, there are reports of myoclonus related to monoamine oxidase inhibitors [\[137\]](#page-26-2) and trazodone [\[126\]](#page-25-8).

Janssen et al. reported that the distribution of myoclonus associated with serotonin reuptake inhibitors is mainly multifocal and generalized [\[5\]](#page-21-4). On the other hand, TCAs are related to focal (especially jaw), multifocal, and generalized myoclonus. Garvey et al. reported a prevalence of 30% of myoclonus in individuals using TCA, but only 9% was clinically significant [\[138\]](#page-26-3). Rissardo et al. found 26 cases of myoclonus associated with amitriptyline. They correlated the occurrence of myoclonus with the effect of amitriptyline on the serotonin receptors, and a dose-dependent effect was observed [\[111\]](#page-25-6).

The mechanism of antidepressant-induced myoclonus is unclear but may be related to increased serotonergic transmission. A study showed EEG and evoked potential abnormalities in TCA-induced myoclonus [\[92\]](#page-24-6). In another post-marketing pharmacovigilance database study, phenelzine was associated with the highest reported adjusted odds ratio for antidepressant-induced movement disorders, followed by clomipramine [\[139\]](#page-26-4). SSRIs increase serotonin levels in the synaptic cleft, and TCAs increase serotonin activity. Interestingly, a combination of TCA and lithium appears to have a compound effect more likely to cause myoclonus than these drugs administered in an isolated fashion [\[140\]](#page-26-5).

8. Antiemetics

Dopamine and 5HT3 antagonists have already been associated with myoclonus. Due to their dopamine receptor blocker properties, most antiemetics, such as prochlorperazine, promethazine, and metoclopramide, may be associated with some tardive disorders [\[141\]](#page-26-6).

Metoclopramide was commonly associated with myoclonus, but this medication alone was never related to myoclonus [\[142\]](#page-26-7). Metoclopramide is believed to have a 5-HT3 receptor-blocking effect; although this effect is weak, it may influence the development of myoclonus [\[143\]](#page-26-8). Hyser et al. reported a case of metoclopramide-induced multifocal myoclonic jerking in the setting of chronic kidney disease, which ceased after the discontinuation of metoclopramide [\[144\]](#page-26-9). Harada et al. reported a 40-year-old patient who was observed to be unresponsive, with occasional myoclonus in the legs following an intramuscular injection of metoclopramide [\[143\]](#page-26-8).

The dopaminergic, cholinergic, and GABA-ergic systems may be involved in promethazine-induced movement disorders [\[145\]](#page-26-10). Dy et al. reported a 63-year-old patient experiencing inducible myoclonus after receiving dextromethorphan–promethazine cough syrup for an upper respiratory infection [\[146\]](#page-26-11).

Other antiemetics associated with myoclonus were ondansetron [\[147\]](#page-26-12) and palonosetron [\[148\]](#page-26-13). Interestingly, palonosetron has allosteric interactions and positive

cooperativity with 5-HT3 receptors, while ondansetron has simple bimolecular competitive binding. Also, palonosetron may trigger 5-HT3 receptor internalization and the degradation of the internalized receptor, which reduces receptor density at the cell surface and leads to the prolonged inhibition of 5-HT3 receptor function [\[149\]](#page-26-14).

9. Antihistamines

Cimetidine-induced myoclonus is usually associated with dose-dependent encephalopathy and is more frequently observed in renal and hepatic dysfunction in elderly individuals. Other symptoms, besides myoclonus, include mild disorientation and psychosis; the symptoms fully improve after the discontinuation of cimetidine [\[150\]](#page-26-15). Oxatomide, a second-generation antihistamine, is prescribed for allergies in Europe and Japan, and has already been reported with myoclonus. The authors hypothesized that minor toxic– metabolic disturbance caused by antihistamine use might have driven their elderly subject, a myoclonus-prone patient, into a transient myoclonic state [\[151\]](#page-26-16). Other antihistamines associated with myoclonus were triprolidine [\[152\]](#page-26-17) and tripelennamine [\[153\]](#page-26-18).

10. Antineoplastic

Among the antineoplastic and immunosuppressive agents, alkylating agents were the most common class associated with myoclonus. Chlorambucil-induced myoclonus was associated with therapeutic doses and intoxication levels [\[154\]](#page-26-19). The electroencephalographic findings were generalized slowing or paroxysms of high-amplitude spike–wave activity [\[155\]](#page-26-20). The discontinuation of this nitrogen mustard agent usually improves the myoclonus [\[156\]](#page-26-21). Other alkylating agents associated with myoclonus were cyclophosphamide [\[157\]](#page-26-22), ifosfamide [\[158\]](#page-26-23), and busulfan [\[159\]](#page-26-24).

Monoclonal antibodies like ipilimumab [\[160\]](#page-26-25), nivolumab [\[161\]](#page-26-26), and pembrolizumab [\[162\]](#page-27-0) have already been reported to be associated with myoclonus. Ipilimumab, a fully human IgG1 monoclonal antibody against cytotoxic T-lymphocyte-associated protein 4 (CTLA-4), is an effective treatment for melanoma. Nivolumab, a fully human IgG4 monoclonal antibody against programmed cell death protein 1 (PD-1), is an effective treatment for non-small cell lung cancer and melanoma, among other cancers [\[163\]](#page-27-1). Also, the nitrogen mustard derivative prednimustine [\[164\]](#page-27-2) and some nucleoside analogs, such as 5-fluorouracil [\[165\]](#page-27-3), floxuridine [\[166\]](#page-27-4), and pentostatin [\[167\]](#page-27-5), were associated with myoclonus.

Tacrolimus-induced severe neurotoxicity in the form of myoclonus, seizures, and leukoencephalopathy is uncommon and occasionally reported in kidney transplant recipients. Tacrolimus has been reported to cause generalized myoclonus, myoclonus of both lower limbs, and segmental myoclonus of the abdominal wall [\[168\]](#page-27-6). Cyclosporine is a lipophilic, cyclic oligopeptide that inhibits calcineurin and modulates the immune system by suppressing T-cell proliferation. Tremor is the most common neurological complication of cyclosporine [\[169\]](#page-27-7). Kang et al. reported a case of opsoclonus–myoclonus secondary to cyclosporine therapy. The authors believe that the highly lipophilic nature of cyclosporine allows it to cross the BBB and provoke changes in neurotransmission through altered dopamine receptor function [\[170\]](#page-27-8).

Topoisomerase drugs were rarely associated with myoclonus. There are isolated reports associating myoclonus with irinotecan and pyrazoloacridine [\[171\]](#page-27-9). Shun et al. suggested that benzodiazepine should be considered the drug of choice in managing irinotecan-induced myoclonus [\[172\]](#page-27-10).

11. Antiparkinsonian

The antiparkinsonian medications most commonly associated with myoclonus were levodopa and amantadine. Rissardo et al. found 22 individuals with amantadine-associated

myoclonus reported in the literature [\[173\]](#page-27-11). The exact mechanism causing this side effect is unknown. However, it is thought that it could act by increasing stimulatory neurotransmitters like dopamine, norepinephrine, and sigma-1 receptors in the cortico-striatopallido-thalamo-cortical loop in the brain and antagonizing inhibitory neurotransmitters like acetylcholine and glutamate [\[174\]](#page-27-12). Interestingly, amantadine has been shown to cause craniofacial myoclonus [\[175\]](#page-27-13).

Dopamine precursors, especially levodopa, were among the most common classes of medication associated with myoclonus. In 1969, Cotzias et al. reported the first cases of myoclonus associated with levodopa [\[176\]](#page-27-14). In 1975, Klawans et al. described 12 individuals who developed levodopa-induced myoclonus after at least one year of levodopa. They described that the myoclonus was bilateral and was not interrupted by the sleep cycle; also, seven individuals developed choreiform dyskinesias [\[177\]](#page-27-15). Nausieda et al. found a correlation between the prevalence and severity of myoclonus and the duration of levodopa therapy [\[178\]](#page-27-16). Luquin et al. reported an incidence of 3.5% of myoclonus in individuals with PD with levodopa therapy [\[179\]](#page-27-17). Also, apparently, PD progression influences the incidence of myoclonus. Marconi et al. revealed an incidence of 66% in individuals with at least ten years of levodopa therapy [\[180\]](#page-27-18). Cases of negative myoclonus and seizures were also associated with levodopa therapy [\[181\]](#page-27-19). Interestingly, in old literature, myoclonus secondary to levodopa was included in the spectrum of levodopa-induced dyskinesia [\[182\]](#page-27-20), which can be a confusing term due to the current understanding of myoclonus pathophysiology with electrodiagnostic studies.

Vardi et al. reported six cases of myoclonus associated with bromocriptine [\[183\]](#page-27-21). Tandberg et al. noticed that individuals recently diagnosed with PD without therapy have an increased incidence of myoclonus compared to healthy individuals [\[184\]](#page-27-22). Other antiparkinsonian drugs associated with myoclonus were trihexyphenidyl [\[185\]](#page-27-23), entacapone [\[186\]](#page-27-24), pramipexole [\[187\]](#page-27-25), and selegiline [\[9\]](#page-21-8).

12. Antipsychotics

There are reports of myoclonus associated with typical and atypical antipsychotics. Historically, typical antipsychotics were more frequently reported with myoclonus, but recent studies showed that atypical antipsychotics are more commonly associated. Haddad et al. proposed that atypical antipsychotics lower the seizure threshold to a more significant degree than typical antipsychotics [\[188\]](#page-27-26). The incidence of clinical seizures with antipsychotics is 0.5–1.2% in patients without a history of epilepsy, and only EEG seizures are observed in around 7% of the individuals [\[189\]](#page-27-27). However, the low rate of recognition may be due to the complexity of psychiatric illness symptoms and the atypicality of psychomotor seizures. Also, the number of prescriptions for atypical compared to typical antipsychotics has significantly increased in the last few decades [\[190\]](#page-27-28). Janssen et al. reported that the myoclonus distribution was multifocal for typical and multifocal to generalized for atypical [\[5\]](#page-21-4). For antipsychotic medications associated with myoclonus in the literature, consider reading Table S5 [\[191–](#page-28-0)[200\]](#page-28-1).

Chlorpromazine and clozapine, both antipsychotics, have a relatively high potential to induce seizures [\[201\]](#page-28-2). Clozapine is already associated with all types of seizures with and without impaired awareness. Although generalized tonic–clonic seizures are frequently observed, myoclonic seizures may occur and may be encountered in daily practice [\[202\]](#page-28-3). Also, among the antipsychotics, clozapine stands out for its heightened seizure risks, especially during titration and at high doses, necessitating close monitoring and individualized approaches [\[203\]](#page-28-4).

Altıparmak studied antipsychotic-induced myoclonus in an inpatient psychiatric hospital. Six of the ten patients in the study who developed myoclonus received clozapine, two received olanzapine, one received amisulpride, and one received quetiapine. The mean age was 24.2 years, and the mean duration of the psychiatric disorder was 33.5 months. Valproic acid was prescribed to eight individuals to control these myoclonic seizures, while lorazepam and clonazepam were used in others [\[192\]](#page-28-5). Of note, levetiracetam can also be used for the management of myoclonus but is not commonly prescribed in clinical practice in psychiatric wards due to dose-independent aggression and psychotic side effects.

Tominaga et al. proposed the term "tardive myoclonus", which was defined as a postural myoclonus associated with the long-term use of antipsychotic therapy [\[204\]](#page-28-6). Fukuzako et al. reported that 38% of the individuals in a psychiatry hospital had tardive myoclonus; they also observed that the antipsychotic dose was significantly higher in these individuals compared to those who did not develop tardive myoclonus [\[205\]](#page-28-7). Ortí-Pareja et al. reported an incidence of only 1% of tardive myoclonus, but the population assessed was those referred to the neurology service [\[206\]](#page-28-8). Interestingly, Little et al. reported a case of myoclonus after five months of antipsychotic withdrawal [\[207\]](#page-28-9). Also, Staedt et al. described nocturnal myoclonus in all individuals with schizophrenia and long-term antipsychotic therapy [\[208\]](#page-28-10).

Myoclonus was observed with subtherapeutic, therapeutic, and higher doses of quetiapine. Uvais et al. reported a case of quetiapine-induced myoclonus that was sensitive to posture (more in lying down position) in a 64-year-old male diagnosed with mild depression and insomnia, even with a single low dose of 12.5 mg per night [\[209\]](#page-28-11). Aggarwal et al. presented two cases of probable quetiapine-induced myoclonus at high doses (400–800 mg). The first individual was a 19-year-old female with mania who was started on 400 mg/day quetiapine and developed right upper limb myoclonic jerks, which were resolved by reducing the dose to 200 mg/day. The second case was a 17-year-old female with schizophrenia who developed myoclonus on clozapine 250 mg and then, after an adequate washout period, again developed myoclonus while on 600 mg/day quetiapine. Both cases fully recovered after reducing the dose of quetiapine [\[197\]](#page-28-12). In addition, Baysal Kirac et al. reported a case of a patient with dementia and a positive family history of juvenile myoclonic epilepsy who was given quetiapine and then developed myoclonic status epilepticus within one month [\[210\]](#page-28-13). Velayudhan et al. also reported a 64-year-old man with schizophrenia who developed myoclonus after quetiapine 800 mg/day was introduced; the myoclonus resolved after reducing the dose to 400 mg/day [\[211\]](#page-28-14).

The pathophysiology of antipsychotic-induced myoclonus is not fully understood. However, the action of quetiapine on serotonergic, dopaminergic, and gamma-aminobutyric acid (GABA)-ergic mechanisms can potentially cause myoclonic jerks [\[5\]](#page-21-4).

13. Antiseizure Medications

Magaudda et al. reported a 31-year-old male with idiosyncratic epilepsy taking carbamazepine (CBZ) 800 mg/day, who developed a subcortical myoclonus involving the right thumb and shoulder [\[212\]](#page-28-15). The myoclonic jerks improved with carbamazepine withdrawal, and they returned with the carbamazepine rechallenge. Similar observations were already described in adults and pediatric individuals. Interestingly, the plasma carbamazepine levels and EEG were normal [\[213\]](#page-28-16). Dhuna et al. presented a child who developed subcortical, multifocal myoclonus, which resolved within 24 h after carbamazepine was discontinued. This exacerbation occurred with therapeutic carbamazepine serum levels and was thought to be related to the toxic levels of carbamazepine-10,11-epoxide (CBZE) metabolite [\[214\]](#page-28-17). Holtmann et al. stated that carbamazepine-induced myoclonus is idiosyncratic rather than dose-related [\[215\]](#page-28-18). Parmeggiani et al. proposed that increased cortical inhibition could be the electrophysiological basis of carbamazepine-induced asterixis. They also proposed that the presence of spike–wave (rather than sharp wave) discharges in children with benign

epilepsy with centro-temporal spikes (BECTS) might be used as an electrophysiological predictor of an abnormal response to CBZ [\[216\]](#page-28-19). For antiseizure medications associated with myoclonus in the literature, consider reading Table S6 [\[217–](#page-28-20)[232\]](#page-29-0).

14. Antiviral

Ganciclovir, acyclovir, valacyclovir, and foscarnet are all known to cause neurotoxic side effects, albeit rarely. Use may lead to tremors, myoclonus, dysarthria, ataxia, delirium, hallucinations, and lethargy [\[233\]](#page-29-1). Acyclovir-induced myoclonus was uncommonly reported in the literature, but the first trials of this medication revealed that myoclonus occurred in 18% of the individuals, likely associated with intoxication [\[234\]](#page-29-2). Haefeli et al. found that acyclovir caused tremor/myoclonus in more than half of the subjects [\[235\]](#page-29-3). In other studies, a rare reversible encephalopathy occurring in less than one percent of the individuals treated with conventional doses of acyclovir was reported [\[236\]](#page-29-4). Other symptoms of acyclovir encephalopathy were disorientation (58%), decreased consciousness (38%), hallucinations (36%), agitation (27%), and dysarthria (19%) [\[234\]](#page-29-2).

Vidarabine, another antiviral related to purines, was also associated with myoclonus [\[237\]](#page-29-5). However, no antiviral medication pyrimidine analog was found to be associated with myoclonus in the literature, including animal studies.

15. Anxiolytics

Myoclonus was associated with the initiation of some anxiolytic drugs and also with their withdrawal; some examples of this association are buspirone [\[238\]](#page-29-6), carisoprodol [\[239\]](#page-29-7), lorazepam [\[240\]](#page-29-8), and midazolam [\[23\]](#page-22-19).

Carisoprodol-induced myoclonus was associated with overdose in the literature. In 2012, carisoprodol was placed on Schedule IV by the Drug Enforcement Administration [\[241\]](#page-29-9). Carisoprodol is not detected on all toxicology tests, which may delay the diagnosis of overdose. A cortical myoclonus source was observed in individuals taking carisoprodol [\[239\]](#page-29-7).

In experimental studies, the Guinea baboon (Papio papio) can present two different types of myoclonus. One type, induced by photic stimulation (intermittent luminous stimulation) preceded by paroxysmal discharges, can be blocked by benzodiazepines. On the other hand, type two of myoclonus may be facilitated by lorazepam and diazepam, lowering the seizure threshold observed on EEG [\[242\]](#page-29-10). Based on these observations, we can assume that the origin of benzodiazepine-induced myoclonus is related to a cortical source, which was interestingly noticed with other anxiolytics [\[243\]](#page-29-11).

16. Cardiovascular

There are some isolated case reports of calcium channel blockers associated with myoclonus. Nifedipine was associated with myoclonus and dysarthria [\[244\]](#page-29-12). Verapamil was associated with myoclonic dystonia [\[245\]](#page-29-13) and multifocal myoclonus [\[246\]](#page-29-14). Amlodipineinduced myoclonus was reported in the setting of stable chronic renal failure [\[247\]](#page-29-15). Diltiazem, in therapeutic doses and in combination with citalopram, was considered responsible for myoclonus while recumbent and in response to startling [\[248\]](#page-29-16).

Carvedilol, a nonselective beta-adrenergic blocker, was reported to cause multifocal myoclonus without other clinical signs [\[249\]](#page-29-17). Other antihypertensives, such as ketanserin [\[250\]](#page-29-18) and furosemide [\[9\]](#page-21-8), were also associated with myoclonus. Interestingly, González et al. reported that enalapril-induced myoclonus was a dose-dependent side effect [\[251\]](#page-29-19).

The fixed-dose combination of sacubitril/valsartan, usually prescribed for congestive heart failure, was associated with myoclonus [\[252\]](#page-30-0). In repeated dose studies in mice and rats but not in primates, increased locomotor activity, twitches, and sensitivity to touch were observed with sacubitril/valsartan [\[253\]](#page-30-1). Also, enkephalinase inhibitors, with neutral endopeptidase inhibitors being part of this family, were previously shown to interfere with the dopaminergic system in experimental models [\[254\]](#page-30-2).

Vasopressor-induced myoclonus was already observed with dobutamine and midodrine. Sympathomimetics can facilitate neuromuscular transmission followed by the prolonged rapid stimulation of motor nerves, which is most likely mediated by alpha-1 receptors [\[255\]](#page-30-3). The cause of dobutamine-induced myoclonus is not yet fully understood, but it may be related to kidney failure and neurotoxicity [\[256\]](#page-30-4). It is suggested that inhibiting P-glycoprotein reduces the breakdown of dobutamine and makes it easier for the substance to enter the central nervous system. Also, chronic kidney disease may affect the half-life of dobutamine [\[257\]](#page-30-5).

17. Opioids

Myoclonus may occur as a result of the initial administration [\[258\]](#page-30-6), change [\[259\]](#page-30-7), or withdrawal [\[260\]](#page-30-8) of opiates. Janssen et al. reported that the distribution of opioid-induced myoclonus is usually generalized or multifocal, but focal cases were already described [\[5\]](#page-21-4). Also, myoclonus associated with opioids is frequently associated with the use of other medications, such as antidepressants and antipsychotics [\[261\]](#page-30-9). For opioids associated with myoclonus in the literature, consider reading Table S7 [\[9,](#page-21-8)[261](#page-30-9)[–273\]](#page-30-10).

The long-term use of high doses of opiates, like in palliative care, was already correlated with an increased frequency of myoclonus. A retrospective review of 48 terminally ill hospice patients who received continuous parenteral hydromorphone for pain control was studied. The authors reported that agitation, myoclonus, and seizures were independently associated with the maximal dose ($p < 0.05$) and with the duration ($p < 0.05$) of continuous parenteral hydromorphone. A possible explanation for these findings could be the hydromorphone-3-glucuronide, a metabolic product of hydromorphone, which has been implicated in neuroexcitatory symptoms in laboratory investigations [\[11\]](#page-21-10). However, McCann et al. found no association between the plasma levels of morphine-3-glucuronide or hydromorphone-3-glucuronide and myoclonus [\[274\]](#page-30-11).

Almedallah et al. reported a 24-year-old pregnant woman who was post-dated and had to undergo a cesarean section, for which epidural anesthesia with fentanyl, bupivacaine, and lidocaine was administered. She developed myoclonus, involving the upper and lower limbs, head, and torso while preserving consciousness. Myoclonus improved after fentanyl discontinuation, and no plasma concentration of fentanyl was noticed. She received morphine, tramadol, and lornoxicam without the occurrence of abnormal movements [\[24\]](#page-22-20). Similarly, Bruera et al. proposed the subcutaneous administration of an acute fentanyl overdose with 5000 mcg to a 62-year-old gentleman for the management of cancer pain, which resulted in generalized myoclonus along with confusion, restlessness, visual hallucinations, hyperalgesia, and tremors on the tactile stimulation of the arms and legs [\[275\]](#page-30-12). The pathophysiology proposed by some authors is general central excitability, with few cases reporting a full improvement of myoclonus after naloxone injections [\[276\]](#page-30-13).

The management of morphine-induced myoclonus has already been reported in several cases in which the frequency of myoclonus was reduced with the prescription of clonazepam [\[277\]](#page-30-14), dantrolene [\[278\]](#page-30-15), and midazolam [\[279\]](#page-30-16). In experimental studies, ketamine improved morphine-induced hindlimb myoclonic seizures [\[280\]](#page-30-17).

Scott et al. attributed this abnormal motor activity either to myoclonus produced by fentanyl causing the depression of higher central nervous system inhibitory centers or to extreme narcotic-induced rigidity [\[281\]](#page-31-0). Lane et al. found that fentanyl infusion withdrawal can cause myoclonus in the pediatric population [\[282\]](#page-31-1). A retrospective analysis described by Smith et al. showed that out of 127 surgical patients, 93 developed mild to moderate rigidity and received fentanyl, sufentanyl, and elfentanyl, as recorded by EEG and EMG [\[283\]](#page-31-2). It is noteworthy that some other anesthetics besides fentanyl can also cause myoclonus, so cases of fentanyl-induced myoclonus in surgical settings should be described with all the medications prescribed for the subject. Furthermore, a significant increase in myoclonus related to opioids, especially fentanyl, is likely to be observed in the following decades due to the "opioid crisis" [\[284\]](#page-31-3).

18. Others

18.1. Heavy Metals

Bismuth salts are among the most commonly reported medications associated with myoclonus. They can cause encephalopathy and myoclonus, characterized by generalized, asymmetric, and stimulus-sensitive jerks during rest or action [\[285\]](#page-31-4). In severe cases, status epilepticus, coma, and death have been reported [\[286\]](#page-31-5). After drug withdrawal, the encephalopathy usually resolves, but approximately 10% of the individuals will have a bad outcome, including mortality and neuropsychiatric sequels [\[287\]](#page-31-6). Electroencephalographic findings are usually unspecific, and CSF analysis can show increased 5-hydroxy-indoleacetic acid levels [\[288\]](#page-31-7).

18.2. Lithium

Rissardo et al. found 97 reports of lithium-induced myoclonus, and the distribution was focal, multifocal, and generalized. The mean age was 53.1, and the lithium dose was 942.7 mg/day [\[289\]](#page-31-8). Dyson et al. found an incidence of 25% of myoclonus in intoxicated individuals [\[290\]](#page-31-9), and Bender et al. found a prevalence of 4.5% in individuals with affective disorder [\[291\]](#page-31-10). Based on the electrodiagnostic studies, we can hypothesize that myoclonus could occur due to dysfunctional cerebellar output, leading to cortical hyperexcitability [\[292\]](#page-31-11). Moreover, giant somatosensory evoked potentials are commonly reported as a remarkable feature of myoclonus occurring with lithium monotherapy or when combined with antidepressants [\[293\]](#page-31-12).

19. Discussion

19.1. Drug-Induced Myoclonus Pathophysiological Mechanism

The myoclonus caused by most medications cannot be hypothesized because the reports did not include electrodiagnostic studies. Table [2](#page-16-0) describes some of the proposed mechanisms for DIM. Increased serotonergic transmission may be the most commonly proposed pathophysiological mechanism for developing myoclonus [\[92\]](#page-24-6). Also, apparently, there are some risk factors associated with DIM, including advanced age, neurodegenerative comorbidities, a history of epilepsy, impaired renal function, electrolyte imbalance, and polypharmacy [\[9\]](#page-21-8).

Table 2. Proposed mechanisms for drug-induced myoclonus.

hydroxytryptophan.

The neurotransmitters likely associated with DIM are serotonin, dopamine, GABA, and glutamate at various levels of the neuraxis (Figure [2\)](#page-17-0). The schema of anatomical and neurophysiological mechanisms can help in clinical decisions and facilitate mechanismbased intervention. For anatomical localization, clinically, spinal segmental myoclonus can involve multiple groups of muscles innervated by 1–3 adjacent spinal levels, contrasting with peripherally generated myoclonus involving specific nerve roots, plexus, trasting with peripherally generated myoclonus involving specific nerve roots, plexus, and peripheral nerves. Another interesting fact distinguishing propriospinal myoclonus and peripheral nerves. Another interesting fact distinguishing propriospinal myoclonus from corticospinal myoclonus is the relatively slow conduction of propriospinal pathways from corticospinal myoclonus is the relatively slow conduction of propriospinal pathways related to high latencies involving the axial musculature compared to corticospinal origin. related to high latencies involving the axial musculature compared to corticospinal origin. Nonetheless, adding to this, myoclonus with underlying encephalopathy can present with Nonetheless, adding to this, myoclonus with underlying encephalopathy can present with appendicular and axial myoclonus and cranial muscle involvement [\[308\]](#page-31-27). appendicular and axial myoclonus and cranial muscle involvement [308].

Figure 2. Classification of drug-induced myoclonus based on possible pathophysiological mechanisms.

nisms. *19.2. Drug-Induced Asterixis*

Asterixis was uncommonly associated with medications (Table S8). Asterixis can be attributed to the direct or indirect effect of the medications on the CNS. The direct effect can be explained by abnormalities in the neurotransmitter levels and toxic levels of the medications. On the other hand, the indirect effect is usually observed with medications that lead to hepatic dysfunction and increased levels of ammonia, predisposing the individual to the development of asterixis [\[181\]](#page-27-19). Drug-induced asterixis is restricted to a lower number of medications compared to myoclonus in general, so this can help in the differential diagnosis of patients presenting with asterixis. For medications associated with asterixis in the literature, consider reading Table S8 [\[58](#page-23-12)[,142](#page-26-7)[,145,](#page-26-10)[147](#page-26-12)[,148,](#page-26-13)[173,](#page-27-11)[309](#page-31-28)[–330\]](#page-32-0).

19.3. Proposed Classifications

We would like to propose a classification for DIM based on its symptomatology and main mechanism (Table [3\)](#page-18-0). This classification is not strict; a significant overlap can be observed with a single medication. For example, valproate can be associated with types 2A and 2B, but it is not usually related to type 3. We noticed these features after observing some common characteristics in the literature and after reports of some rare and anecdotal drugs causing myoclonus.

Table 3. Proposed classification for drug-induced myoclonus.

First, the most commonly reported etiology for DIM is probably serotonin syndrome. Some patients developed the full syndrome, and others with the same combination of medications developed only myoclonus. The isolated myoclonus is part of a spectrum, which we would like to call the serotonin syndrome spectrum. The most common criteria used for diagnosing serotonin syndrome are Sternbach's, Radomski's, and Hunter's criteria [\[331\]](#page-32-1).

Second, some patients developed clinical encephalopathy before the development of myoclonus, without the fulfillment of serotonin syndrome criteria. In this context, we found two groups of individuals: one that developed associated liver injury and the other that did not have liver injury. Both groups of individuals had an overall worse prognosis compared to serotonin syndrome alone. It is noteworthy that the non-hepatic encephalopathy associated with myoclonus can also be categorized as drug-induced Creutzfeldt–Jakob-like syndrome. Drug-induced Creutzfeldt–Jakob-like syndrome is characterized by confusion, myoclonus, and EEG abnormalities associated with the use of a drug [\[332\]](#page-32-2). The most frequently reported medications associated with this specific constellation of symptoms are lithium [\[289\]](#page-31-8), TCA [\[111\]](#page-25-6), and carbamazepine [\[218\]](#page-28-21).

A third group of individuals were observed to have only myoclonus and did not develop any other sign of serotonin syndrome or encephalopathy. Interestingly, this group of patients had an overall better prognosis than the first and second groups. However, further investigations regarding the management and follow-up of these individuals are needed.

19.4. Management

There has been an increased number of medications associated with myoclonus within recent decades. The recommended approach is to follow systematic reviews that have already been performed with the specific medication class, but the number of these studies is still limited. Therefore, we would like to provide some basic rules for managing DIM.

Management usually includes stopping the offending drug or modifying its dosage. However, this is a complex recommendation in clinical practice since sometimes it is not feasible due to underlying neuropsychiatric conditions, along with the risks and benefits of the responsible drug. Other options include adding specific treatments like drugs that act on the idiopathic myoclonus, such as benzodiazepines and antiseizure medications [\[92\]](#page-24-6). Another possible approach is noninvasive brain stimulation procedures, such as transcranial magnetic and electrical stimulation techniques. Occupational therapy, speech therapy, and physiotherapy can also be implemented in selected and resistant cases [\[333\]](#page-32-3).

Classifying myoclonus on neurophysiological subtypes is important for management. Pena et al. explained that levetiracetam, valproic acid, and clonazepam are often used to treat cortical myoclonus. For cortical–subcortical myoclonus, the treatment of myoclonus is prioritized; hence, valproic acid is the mainstay of therapy. Subcortical–nonsegmental myoclonus can be treated with clonazepam. However, many other drugs can be used, according to the etiology. Segmental and peripheral myoclonus are difficult to treat and often remain resistant to treatment, but anticonvulsants and botulinum toxin injections can be attempted. The choice of drugs depends on the adverse effects, efficacy, and evidencebased knowledge, which is limited and unreliable due to non[sta](#page-21-6)ndard data [7]. Noteworthy, this approach is based on idiopathic myoclonus, so similar results may not be obtained wit[h D](#page-19-0)IM (Figure 3).

Figure 3. Algorithm of management of drug-induced myoclonus. **Figure 3.** Algorithm of management of drug-induced myoclonus.

20. Future Studies 20. Future Studies

Significant progress has been made in understanding the pathophysiology and man-Significant progress has been made in understanding the pathophysiology and management of idiopathic myoclonus. However, the literature on drug-induced myoclonus is agement of idiopathic myoclonus. However, the literature on drug-induced myoclonus is scarce. Future studies should assess the epidemiological data regarding myoclonus sec-scarce. Future studies should assess the epidemiological data regarding myoclonus secondary to medications. Online databases such as the FDA Adverse Event Reporting System (FAERS) should register subjects with more clinical details, including basic demographic features and electrodiagnostic studies. Most data published about myoclonus still lack lack basic descriptions of EEG and EMG, and therefore no source can be identified, which basic descriptions of EEG and EMG, and therefore no source can be identified, which can result in misleading information about management.

The effect of medications on different neurotransmitters should be further investigated. There are studies in the literature with completely opposite results regarding neurotransmitters when using the same medication. Also, it is not uncommon to find published studies in the literature that were never replicated. In this way, there could be misleading information about attempts to provide pathophysiological explanations based on neurotransmitters.

While an increase in cases of DIM has been observed, more epidemiological data are required to understand the drugs' nature, side effects, management, economic burden, and prognosis. More studies need to be performed on stable populations, with more pharmacodynamic investigations of the pharmacological agents. Similarly, more studies should be conducted by combining the medications used to control myoclonic symptoms. Ideally, to maximize reliability, multicenter clinical trials must be performed on cohorts of patients with myoclonus but without underlying neurological conditions and renal dysfunction. The lack of homogenous clinical features means that the number of drugs that could possibly cause myoclonus is underestimated. Providing reliable data about the prevalence and outcomes of this motor side effect to patients and caregivers when using individual medications in various healthcare settings remains a challenge.

21. Conclusions

The subclasses of medications with level A evidence were intravenous anesthetics (etomidate), cephalosporins (ceftazidime, cefepime), fluoroquinolones (ciprofloxacin), selective serotonin reuptake inhibitors (citalopram, escitalopram, paroxetine, sertraline), tricyclic antidepressant (amitriptyline), glutamate antagonist (amantadine), atypical antipsychotics (clozapine, quetiapine), antiseizure medications (carbamazepine, oxcarbazepine, phenytoin, gabapentin, pregabalin, valproate), pure opioid agonists (fentanyl, morphine), bismuth salts, and mood stabilizers (lithium).

The distribution of myoclonus ranges from focal to generalized, even amongst patients using the same drug, which suggests various neuro-anatomical generators. In many cases, DIM subsides with the cessation of the responsible drug, but specialized treatments and therapies are common. Due to the heterogeneous nature of myoclonus secondary to drugs, DIM should always be assessed as a differential diagnosis of myoclonus. The dosedependent progression of myoclonus from multifocal to generalized and the involvement of CNS symptoms like confusion and visual hallucinations are observed with an increase in the dosage of opioids like fentanyl, indicating the need for the modification of the offending drug's dose. Pretreatment with certain drugs like alfentanil, dexmedetomidine, propofol, butorphanol, dezocine, and midazolam in etomidate-induced myoclonus emerged as a therapeutic option for suppressing myoclonus. Patients with isolated myoclonus who do not develop any serotonin syndrome or encephalopathy are likely to have a better prognosis. Besides cephalosporins and fluoroquinolones, antibiotics such as lipopeptides, glycopeptides, and tetracyclines were rarely associated with DIM. Certain drugs like TCAs and lithium, when used in combination, are more likely to cause DIM compared to their use in isolation.

In conclusion, our overview confirms the level of evidence supporting various drugs causing myoclonus. Our refinement of DIM may offer clinicians a practical approach to discussing therapeutic benefits and the side effects of drugs, including myoclonus, while maintaining patients' autonomy and well-being.

Supplementary Materials: The following supporting information can be downloaded at: [https:](https://www.mdpi.com/article/10.3390/medicina61010131/s1) [//www.mdpi.com/article/10.3390/medicina61010131/s1,](https://www.mdpi.com/article/10.3390/medicina61010131/s1) Table S1: Articles of Myoclonus Associated with Drugs in PubMed; Table S2: Anesthetic-Induced Myoclonus; Table S3: Antibiotic-Induced Myoclonus; Table S4: Antidepressant-Induced Myoclonus; Table S5: Antipsychotic-Induced My-

oclonus; Table S6: Antiseizure Medication-Induced Myoclonus; Table S7: Opioid-Induced Myoclonus; Table S8: Drug-Induced Asterixis.

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References

- 1. Dijk, J.M.; Tijssen, M.A.J. Management of Patients with Myoclonus: Available Therapies and the Need for an Evidence-Based Approach. *Lancet Neurol.* **2010**, *9*, 1028–1036. [\[CrossRef\]](https://doi.org/10.1016/S1474-4422(10)70193-9) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20864054)
- 2. Chandarana, M.; Saraf, U.; Divya, K.P.; Krishnan, S.; Kishore, A. Myoclonus—A Review. *Ann. Indian Acad. Neurol.* **2021**, *24*, 327–338. [\[CrossRef\]](https://doi.org/10.4103/aian.AIAN_1180_20) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34446993)
- 3. Friedreich, N. Paramyoclonus Multiplex. *Virchows Arch.* **1881**, *86*, 421–430. [\[CrossRef\]](https://doi.org/10.1007/BF01915725)
- 4. Kojovic, M.; Cordivari, C.; Bhatia, K. Myoclonic Disorders: A Practical Approach for Diagnosis and Treatment. *Ther. Adv. Neurol. Disord.* **2011**, *4*, 47–62. [\[CrossRef\]](https://doi.org/10.1177/1756285610395653) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21339907)
- 5. Janssen, S.; Bloem, B.R.; van de Warrenburg, B.P. The Clinical Heterogeneity of Drug-Induced Myoclonus: An Illustrated Review. *J. Neurol.* **2017**, *264*, 1559–1566. [\[CrossRef\]](https://doi.org/10.1007/s00415-016-8357-z) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27981352)
- 6. Caviness, J.N.; Brown, P. Myoclonus: Current Concepts and Recent Advances. *Lancet Neurol.* **2004**, *3*, 598–607. [\[CrossRef\]](https://doi.org/10.1016/S1474-4422(04)00880-4)
- 7. Pena, A.B.; Caviness, J.N. Physiology-Based Treatment of Myoclonus. *Neurotherapeutics* **2020**, *17*, 1665–1680. [\[CrossRef\]](https://doi.org/10.1007/s13311-020-00922-6)
- 8. Shibasaki, H.; Hallett, M. Electrophysiological Studies of Myoclonus. *Muscle Nerve* **2005**, *31*, 157–174. [\[CrossRef\]](https://doi.org/10.1002/mus.20234)
- 9. Brefel-Courbon, C.; Gardette, V.; Ory, F.; Montastruc, J.L. Drug-Induced Myoclonus: A French Pharmacovigilance Database Study. *Neurophysiol. Clin.* **2006**, *36*, 333–336. [\[CrossRef\]](https://doi.org/10.1016/j.neucli.2006.12.003)
- 10. Caviness, J.N.; Alving, L.I.; Maraganore, D.M.; Black, R.A.; McDonnell, S.K.; Rocca, W.A. The Incidence and Prevalence of Myoclonus in Olmsted County, Minnesota. *Mayo Clin. Proc.* **1999**, *74*, 565–569. [\[CrossRef\]](https://doi.org/10.4065/74.6.565)
- 11. Thwaites, D.; McCann, S.; Broderick, P. Hydromorphone Neuroexcitation. *J. Palliat. Med.* **2004**, *7*, 545–550. [\[CrossRef\]](https://doi.org/10.1089/jpm.2004.7.545) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15353098)
- 12. Page, M.J.; McKenzie, J.E.; Bossuyt, P.M.; Boutron, I.; Hoffmann, T.C.; Mulrow, C.D.; Shamseer, L.; Tetzlaff, J.M.; Akl, E.A.; Brennan, S.E.; et al. The PRISMA 2020 Statement: An Updated Guideline for Reporting Systematic Reviews. *BMJ* **2021**, *372*, n71. [\[CrossRef\]](https://doi.org/10.1136/bmj.n71)
- 13. De Vries, E.; Schoonvelde, M.; Schumacher, G. No Longer Lost in Translation: Evidence That Google Translate Works for Comparative Bag-of-Words Text Applications. *Polit. Anal.* **2018**, *26*, 417–430. [\[CrossRef\]](https://doi.org/10.1017/pan.2018.26)
- 14. Naranjo, C.A.; Busto, U.; Sellers, E.M.; Sandor, P.; Ruiz, I.; Roberts, E.A.; Janecek, E.; Domecq, C.; Greenblatt, D.J. A Method for Estimating the Probability of Adverse Drug Reactions. *Clin. Pharmacol. Ther.* **1981**, *30*, 239–245. [\[CrossRef\]](https://doi.org/10.1038/clpt.1981.154) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7249508)
- 15. Ng, A.T. Prolonged Myoclonic Contractions after Enflurane Anaesthesia—A Case Report. *Can. Anaesth. Soc. J.* **1980**, *27*, 502–503. [\[CrossRef\]](https://doi.org/10.1007/BF03007053) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7448613)
- 16. Laughlin, T.P.; Newberg, L.A. Prolonged Myoclonus after Etomidate Anesthesia. *Anesth. Analg.* **1985**, *64*, 80–82. [\[CrossRef\]](https://doi.org/10.1213/00000539-198501000-00017)
- 17. Wu, M.-S.; Hsu, Y.-D.; Lin, J.-C.; Chen, S.-C.; Lee, J.-T. Spinal Myoclonus in Subacute Combined Degeneration Caused by Nitrous Oxide Intoxication. *Acta Neurol. Taiwanica* **2007**, *16*, 102–105.
- 18. Tam, M.K.P.; Irwin, M.G.; Tse, M.L.; Lui, Y.W.A.; Law, K.I.; Ng, P.W. Prolonged Myoclonus after a Single Bolus Dose of Propofol. *Anaesthesia* **2009**, *64*, 1254–1257. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2044.2009.06035.x)
- 19. Sutter, R.; Ristic, A.; Rüegg, S.; Fuhr, P. Myoclonus in the Critically Ill: Diagnosis, Management, and Clinical Impact. *Clin. Neurophysiol.* **2016**, *127*, 67–80. [\[CrossRef\]](https://doi.org/10.1016/j.clinph.2015.08.009)
- 20. Fox, E.J.; Villanueva, R.; Schutta, H.S. Myoclonus Following Spinal Anesthesia. *Neurology* **1979**, *29*, 379–380. [\[CrossRef\]](https://doi.org/10.1212/WNL.29.3.379)
- 21. Harrison, J.L. Postoperative Seizures after Isoflurane Anesthesia. *Anesth. Analg.* **1986**, *65*, 1235–1236. [\[CrossRef\]](https://doi.org/10.1213/00000539-198611000-00025) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/3767024)
- 22. Conreux, F.; Best, O.; Preckel, M.P.; Lhopitault, C.; Beydon, L.; Pouplard, F.; Granry, J.C. Electroencephalographic effects of sevoflurane in pediatric anesthesia: A prospective study of 20 cases. *Ann. Fr. Anesth. Reanim.* **2001**, *20*, 438–445. [\[CrossRef\]](https://doi.org/10.1016/S0750-7658(01)00393-8) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11419238)
- 23. Magny, J.F.; d'Allest, A.M.; Nedelcoux, H.; Zupan, V.; Dehan, M. Midazolam and Myoclonus in Neonate. *Eur. J. Pediatr.* **1994**, *153*, 389–390. [\[CrossRef\]](https://doi.org/10.1007/BF01956430) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8033934)
- 24. Almedallah, D.K.; Alshamlan, D.Y.; Shariff, E.M. Acute Opioid-Induced Myoclonic Reaction after Use of Fentanyl as an Anesthetic Drug for an Emergency Cesarean Section. *Case Rep. Neurol.* **2018**, *10*, 130–134. [\[CrossRef\]](https://doi.org/10.1159/000486891)
- 25. Boscan, P.; Pypendop, B.H.; Solano, A.M.; Ilkiw, J.E. Cardiovascular and Respiratory Effects of Ketamine Infusions in Isoflurane-Anesthetized Dogs before and during Noxious Stimulation. *Am. J. Vet. Res.* **2005**, *66*, 2122–2129. [\[CrossRef\]](https://doi.org/10.2460/ajvr.2005.66.2122)
- 26. Lee, J.J.; Hwang, S.M.; Lee, J.S.; Jang, J.S.; Lim, S.-Y.; Hong, S.J. Recurrent Spinal Myoclonus after Two Episodes of Spinal Anesthesia at a 1-Year Interval—A Case Report. *Korean J. Anesthesiol.* **2010**, *59*, S62–S64. [\[CrossRef\]](https://doi.org/10.4097/kjae.2010.59.S.S62)
- 27. Watanabe, S.; Sakai, K.; Ono, Y.; Seino, H.; Naito, H. Alternating Periodic Leg Movement Induced by Spinal Anesthesia in an Elderly Male. *Anesth. Analg.* **1987**, *66*, 1031–1032. [\[CrossRef\]](https://doi.org/10.1213/00000539-198710000-00024)
- 28. Nadkarni, A.V.; Tondare, A.S. Localized Clonic Convulsions after Spinal Anesthesia with Lidocaine and Epinephrine. *Anesth. Analg.* **1982**, *61*, 945–947. [\[CrossRef\]](https://doi.org/10.1213/00000539-198211000-00011)
- 29. Fores Novales, B.; Aguilera Celorrio, L. Spinal Myoclonus Following Intrathecal Anaesthesia with Prilocaine. *Anaesth. Intensive Care* **2009**, *37*, 498–499.
- 30. Kang, H.Y.; Lee, S.W.; Hong, E.P.; Sim, Y.H.; Lee, S.-M.; Park, S.W.; Kang, J.-M. Myoclonus-like Involuntary Movements Following Cesarean Delivery Epidural Anesthesia. *J. Clin. Anesth.* **2016**, *34*, 392–394. [\[CrossRef\]](https://doi.org/10.1016/j.jclinane.2016.05.014)
- 31. Drummond, J.C.; Todd, M.M.; Shapiro, H.M. Minimal Alveolar Concentrations for Halothane, Enflurane, and Isoflurane in the Cat. *J. Am. Vet. Med. Assoc.* **1983**, *182*, 1099–1101. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/6863122)
- 32. Sepahvand, M.; Rashidi, S.; Emamikhah, M.; Rohani, M.; Yazdi, N. Laughing Ceased, Nitrous Oxide-Induced Myelopathy Evolved. *Can. J. Neurol. Sci.* **2024**, *51*, 469–471. [\[CrossRef\]](https://doi.org/10.1017/cjn.2023.44) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37066709)
- 33. Hasbaoui, B.E.; Mebrouk, N.; Saghir, S.; Yajouri, A.E.; Abilkassem, R.; Agadr, A. Vitamin B12 Deficiency: Case Report and Review of Literature. *Pan. Afr. Med. J.* **2021**, *38*, 237. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34046142)
- 34. Feng, Y.; Chen, X.-B.; Zhang, Y.-L.; Chang, P.; Zhang, W.-S. Propofol Decreased the Etomidate-Induced Myoclonus in Adult Patients: A Meta-Analysis and Systematic Review. *Eur. Rev. Med. Pharmacol. Sci.* **2023**, *27*, 1322–1335. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36876671)
- 35. Walder, B.; Tramèr, M.R.; Seeck, M. Seizure-like Phenomena and Propofol: A Systematic Review. *Neurology* **2002**, *58*, 1327–1332. [\[CrossRef\]](https://doi.org/10.1212/WNL.58.9.1327)
- 36. Chao, S.; Khan, R.; Lieberman, J.; Buren, M. Propofol-Induced Myoclonus during Maintenance of Anaesthesia. *Anaesth. Rep.* **2023**, *11*, e12253. [\[CrossRef\]](https://doi.org/10.1002/anr3.12253)
- 37. Hellyer, P.W.; Freeman, L.C.; Hubbell, J.A. Induction of Anesthesia with Diazepam-Ketamine and Midazolam-Ketamine in Greyhounds. *Vet. Surg.* **1991**, *20*, 143–147. [\[CrossRef\]](https://doi.org/10.1111/j.1532-950X.1991.tb00324.x)
- 38. Doenicke, A.W.; Roizen, M.F.; Kugler, J.; Kroll, H.; Foss, J.; Ostwald, P. Reducing Myoclonus after Etomidate. *Anesthesiology* **1999**, *90*, 113–119. [\[CrossRef\]](https://doi.org/10.1097/00000542-199901000-00017)
- 39. Feng, Y.; Chang, P.; Kang, Y.; Liao, P.; Li, C.-Y.; Liu, J.; Zhang, W.-S. Etomidate-Induced Myoclonus in Sprague-Dawley Rats Involves Neocortical Glutamate Accumulation and N-Methyl-d-Aspartate Receptor Activity. *Anesth. Analg.* **2023**, *137*, 221–233. [\[CrossRef\]](https://doi.org/10.1213/ANE.0000000000006292)
- 40. Feng, Y.; Liu, J.; Zhang, W.-S. Etomidate-Induced Myoclonus Correlates with the Dysfunction of Astrocytes and Glutamate Transporters in the Neocortex of Sprague-Dawley Rats. *Eur. Rev. Med. Pharmacol. Sci.* **2022**, *26*, 6221–6235.
- 41. Zhou, C.; Zhu, Y.; Liu, Z.; Ruan, L. Effect of Pretreatment with Midazolam on Etomidate-Induced Myoclonus: A Meta-Analysis. *J. Int. Med. Res.* **2017**, *45*, 399–406. [\[CrossRef\]](https://doi.org/10.1177/0300060516682882) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28415947)
- 42. Nooraei, N.; Solhpour, A.; Mohajerani, S.A. Priming with Atracurium Efficiently Suppresses Etomidate-Induced Myoclonus. *Acta Anaesthesiol. Taiwanica* **2013**, *51*, 145–148. [\[CrossRef\]](https://doi.org/10.1016/j.aat.2013.12.005) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/24529669)
- 43. Collin, R.I.; Drummond, G.B.; Spence, A.A. Alfentanil Supplemented Anaesthesia for Short Procedures. A Double-Blind Study of Alfentanil Used with Etomidate and Enflurane for Day Cases. *Anaesthesia* **1986**, *41*, 477–481. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2044.1986.tb13270.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/3089048)
- 44. Du, X.; Zhou, C.; Pan, L.; Li, C. Effect of Dexmedetomidine in Preventing Etomidate-Induced Myoclonus: A Meta-Analysis. *Drug Des. Devel. Ther.* **2017**, *11*, 365–370. [\[CrossRef\]](https://doi.org/10.2147/DDDT.S121979)
- 45. Hua, J.; Miao, S.; Shi, M.; Tu, Q.; Wang, X.; Liu, S.; Wang, G.; Gan, J. Effect of Butorphanol on Etomidate-Induced Myoclonus: A Systematic Review and Meta-Analysis. *Drug Des. Devel. Ther.* **2019**, *13*, 1213–1220. [\[CrossRef\]](https://doi.org/10.2147/DDDT.S191982)
- 46. Zhu, Y.; Yang, Y.; Zhou, C.; Bao, Z. Using Dezocine to Prevent Etomidate-Induced Myoclonus: A Meta-Analysis of Randomized Trials. *Drug Des. Devel. Ther.* **2017**, *11*, 2163–2170. [\[CrossRef\]](https://doi.org/10.2147/DDDT.S137464)
- 47. Greenwood, J.; Crull, A.; Graves, M.; Ledvina, M. Pharmacological Interventions for Reducing the Incidence of Myoclonus in Patients Receiving Etomidate for Induction of General Anesthesia: An Umbrella Review. *JBI Evid. Synth.* **2024**, *22*, 66–89. [\[CrossRef\]](https://doi.org/10.11124/JBIES-22-00390)
- 48. Shiratori, T.; Hotta, K.; Satoh, M. Spinal Myoclonus Following Neuraxial Anesthesia: A Literature Review. *J. Anesth.* **2019**, *33*, 140–147. [\[CrossRef\]](https://doi.org/10.1007/s00540-018-02607-z)
- 49. Slaker, R.A.; Danielson, B. Neurotoxicity Associated with Ceftazidime Therapy in Geriatric Patients with Renal Dysfunction. *Pharmacotherapy* **1991**, *11*, 351–352. [\[CrossRef\]](https://doi.org/10.1002/j.1875-9114.1991.tb04302.x)
- 50. Kurtzman, N.A.; Rogers, P.W.; Harter, H.R. Neurotoxic Reaction to Penicillin and Carbenicillin. *JAMA* **1970**, *214*, 1320–1321. [\[CrossRef\]](https://doi.org/10.1001/jama.1970.03180070084016)
- 51. Calandra, G.B.; Brown, K.R.; Grad, L.C.; Ahonkhai, V.I.; Wang, C.; Aziz, M.A. Review of Adverse Experiences and Tolerability in the First 2,516 Patients Treated with Imipenem/Cilastatin. *Am. J. Med.* **1985**, *78*, 73–78. [\[CrossRef\]](https://doi.org/10.1016/0002-9343(85)90104-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/3859218)
- 52. Durand, J.M.; Telle, H.; Quilès, N.; Taramasco, V.; Kaplanski, G.; Soubeyrand, J. Confusion syndrome, myoclonus and treatment with pefloxacin. *Ann. Med. Interne* **1993**, *144*, 495–496.
- 53. New, P.S.; Wells, C.E. Cerebral Toxicity Associated with Massive Intravenous Penicillin Therapy. *Neurology* **1965**, *15*, 1053–1058. [\[CrossRef\]](https://doi.org/10.1212/WNL.15.11.1053)
- 54. Park-Matsumoto, Y.C.; Tazawa, T. Piperacillin-Induced Encephalopathy. *J. Neurol. Sci.* **1996**, *140*, 141–142. [\[CrossRef\]](https://doi.org/10.1016/0022-510X(96)00119-0)
- 55. Kallay, M.C.; Tabechian, H.; Riley, G.R.; Chessin, L.N. Neurotoxicity Due to Ticarcillin in Patient with Renal Failure. *Lancet* **1979**, *1*, 608–609. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(79)91032-8)
- 56. Sackellares, J.C.; Smith, D.B. Myoclonus with Electrocerebral Silence in a Patient Receiving Penicillin. *Arch. Neurol.* **1979**, *36*, 857–858. [\[CrossRef\]](https://doi.org/10.1001/archneur.1979.00500490071013)
- 57. Viloria-Alebesque, A.; Povar-Echeverría, M.; Bruscas-Alijarde, M.J.; Gracia-Gutiérrez, A.; Royo-Trallero, L.; Al-Cheikh-Felices, P. Myoclonus Induced by Amoxicillin-Clavulanic Acid. *Epilepsy Behav. Rep.* **2020**, *14*, 100367. [\[CrossRef\]](https://doi.org/10.1016/j.ebr.2020.100367)
- 58. Herishanu, Y.O.; Zlotnik, M.; Mostoslavsky, M.; Podgaietski, M.; Frisher, S.; Wirguin, I. Cefuroxime-Induced Encephalopathy. *Neurology* **1998**, *50*, 1873–1875. [\[CrossRef\]](https://doi.org/10.1212/WNL.50.6.1873)
- 59. Uchihara, T.; Tsukagoshi, H. Myoclonic Activity Associated with Cefmetazole, with a Review of Neurotoxicity of Cephalosporins. *Clin. Neurol. Neurosurg.* **1988**, *90*, 369–371. [\[CrossRef\]](https://doi.org/10.1016/0303-8467(88)90013-3)
- 60. Hagiya, H.; Miyawaki, K.; Yamamoto, N.; Yoshida, H.; Kitagawa, A.; Asaoka, T.; Eguchi, H.; Akeda, Y.; Tomono, K. Ceftriaxone-Induced Neurotoxicity in a Patient after Pancreas-Kidney Transplantation. *Intern. Med.* **2017**, *56*, 3103–3107. [\[CrossRef\]](https://doi.org/10.2169/internalmedicine.8774-16)
- 61. Chan, S.; Turner, M.R.; Young, L.; Gregory, R. Cephalosporin-Induced Myoclonus. *Neurology* **2006**, *66*, E20. [\[CrossRef\]](https://doi.org/10.1212/01.wnl.0000190256.30385.13)
- 62. Cho, I.; Bertoni, J.M.; Hopkins, L. Moxalactam Myoclonus, Seizures, and Encephalopathy. *Drug Intell. Clin. Pharm.* **1986**, *20*, 223–224. [\[CrossRef\]](https://doi.org/10.1177/106002808602000310) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/3956381)
- 63. Khasani, S. Cefepime-Induced Jaw Myoclonus. *Neurology* **2015**, *84*, 1183. [\[CrossRef\]](https://doi.org/10.1212/WNL.0000000000001365) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/25780020)
- 64. Sonck, J.; Laureys, G.; Verbeelen, D. The Neurotoxicity and Safety of Treatment with Cefepime in Patients with Renal Failure. *Nephrol. Dial. Transplant.* **2008**, *23*, 966–970. [\[CrossRef\]](https://doi.org/10.1093/ndt/gfm713) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18175786)
- 65. Zimmermann, P.; Camenzind, D.; Beer, J.H.; Tarnutzer, A.A. Negative Myoclonus as the Leading Symptom in Acute Cefepime Neurotoxicity. *BMJ Case Rep.* **2021**, *14*, e239744. [\[CrossRef\]](https://doi.org/10.1136/bcr-2020-239744)
- 66. Lau, K.K.; Kink, R.J.; Jones, D.P. Myoclonus Associated with Intraperitoneal Imipenem. *Pediatr. Nephrol.* **2004**, *19*, 700–701. [\[CrossRef\]](https://doi.org/10.1007/s00467-004-1467-3)
- 67. Rivera, M.; Crespo, M.; Teruel, J.L.; Marcén, R.; Ortuño, J. Neurotoxicity Due to Imipenem/Cilastatin in Patients on Continuous Ambulatory Peritoneal Dialysis. *Nephrol. Dial. Transplant.* **1999**, *14*, 258–259. [\[CrossRef\]](https://doi.org/10.1093/ndt/14.1.258)
- 68. Frucht, S.; Eidelberg, D. Imipenem-Induced Myoclonus. *Mov. Disord.* **1997**, *12*, 621–622. [\[CrossRef\]](https://doi.org/10.1002/mds.870120430)
- 69. Baraboutis, I.G.; Marangos, M.N.; Skoutelis, A.; Bassaris, H. Meropenem-Aggravated Seizure Activity in Progressive Myoclonus Epilepsy. *Int. J. Antimicrob. Agents* **2008**, *31*, 177–179. [\[CrossRef\]](https://doi.org/10.1016/j.ijantimicag.2007.09.011)
- 70. Spina Silva, T.; Dal-Prá Ducci, R.; Zorzetto, F.P.; Braatz, V.L.; de Paola, L.; Kowacs, P.A. Meropenem-Induced Myoclonus: A Case Report. *Seizure* **2014**, *23*, 912–914. [\[CrossRef\]](https://doi.org/10.1016/j.seizure.2014.06.017)
- 71. Apodaca, K.; Baker, J.; Bin-Bilal, H.; Raskin, Y.; Quinn, D.K. Ertapenem-Induced Delirium: A Case Report and Literature Review. *Psychosomatics* **2015**, *56*, 561–566. [\[CrossRef\]](https://doi.org/10.1016/j.psym.2015.02.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26002226)
- 72. Jayathissa, S.; Woolley, M.; Ganasegaram, M.; Holden, J.; Cu, E. Myoclonus and Delirium Associated with Ciprofloxacin. *Age Ageing* **2010**, *39*, 762. [\[CrossRef\]](https://doi.org/10.1093/ageing/afq107) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/20817933)
- 73. Post, B.; Koelman, J.H.T.M.; Tijssen, M.A.J. Propriospinal Myoclonus after Treatment with Ciprofloxacin. *Mov. Disord.* **2004**, *19*, 595–597. [\[CrossRef\]](https://doi.org/10.1002/mds.10717)
- 74. Javed, H.; Ali, H.T.; Soliman, Z.A.; Caprara, A.L.F.; Rissardo, J.P. Three Cases of Myoclonus Secondary to Ciprofloxacin: "Ciproclonus". *Clin. Neuropharmacol.* **2023**, *46*, 200–203. [\[CrossRef\]](https://doi.org/10.1097/WNF.0000000000000565) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37748004)
- 75. Kayipmaz, S.; Altınöz, A.E.; Ok, N.E.G. Lithium Intoxication: A Possible Interaction with Moxifloxacin. *Clin. Psychopharmacol. Neurosci.* **2017**, *15*, 407–409. [\[CrossRef\]](https://doi.org/10.9758/cpn.2017.15.4.407)
- 76. Reddy, V.A.S.K.; Mittal, G.K.; Sekhar, S.; Singhdev, J.; Mishra, R. Levofloxacin-Induced Myoclonus and Encephalopathy. *Ann. Indian Acad. Neurol.* **2020**, *23*, 405–407.
- 77. Marinella, M.A. Myoclonus and Generalized Seizures Associated with Gatifloxacin Treatment. *Arch. Intern. Med.* **2001**, *161*, 2261–2262. [\[CrossRef\]](https://doi.org/10.1001/archinte.161.18.2261)
- 78. Bagon, J.A. Neuropsychiatric Complications Following Quinolone Overdose in Renal Failure. *Nephrol. Dial. Transplant.* **1999**, *14*, 1337. [\[CrossRef\]](https://doi.org/10.1093/ndt/14.5.1337a)
- 79. Michtell, G.A.J. Extrapyramidal Reaction in a Patient on Combined Drug Therapy: Report of a Case. *Anesth. Prog.* **1971**, *18*, 78–79.
- 80. Vadalá, S.F.; Pellegrini, D.; Silva, E.D.; Miñarro, D.; Finn, B.C.; Bruetman, J.E.; Nápoli, G.; Young, P. Lethargic encephalitis. Report of one case. *Rev. Med. Chil.* **2013**, *141*, 531–534. [\[CrossRef\]](https://doi.org/10.4067/S0034-98872013000400016)
- 81. Sarva, H.; Panichpisal, K. Gentamicin-Induced Myoclonus: A Case Report and Literature Review of Antibiotics-Induced Myoclonus. *Neurologist* **2012**, *18*, 385–388. [\[CrossRef\]](https://doi.org/10.1097/NRL.0b013e3182704d78) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23114671)
- 82. Dib, E.G.; Bernstein, S.; Benesch, C. Multifocal Myoclonus Induced by Trimethoprim-Sulfamethoxazole Therapy in a Patient with Nocardia Infection. *N. Engl. J. Med.* **2004**, *350*, 88–89. [\[CrossRef\]](https://doi.org/10.1056/NEJM200401013500121) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14702438)
- 83. Scolari, M.J.; Pellegrini, D. Daptomycin Associated Myoclonus: A Case Report. *Farm. Hosp.* **2021**, *46*, 40–42. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35379091)
- 84. Patel, U.C.; Fowler, M.A. Ertapenem-Associated Neurotoxicity in the Spinal Cord Injury (SCI) Population: A Case Series. *J. Spinal Cord. Med.* **2018**, *41*, 735–740. [\[CrossRef\]](https://doi.org/10.1080/10790268.2017.1368960)
- 85. Jacob, J.S.; Cohen, P.R. Doxycycline-Induced Hand Tremors: Case Report and Review of Antibiotic-Associated Tremors. *Cureus* **2020**, *12*, e10782. [\[CrossRef\]](https://doi.org/10.7759/cureus.10782)
- 86. Ferreira, Â.; Sobrosa, P.; Costa, M.; Miranda, I.; Guerra, D. Linezolid Toxicity: A Clinical Case Report. *Cureus* **2024**, *16*, e55672. [\[CrossRef\]](https://doi.org/10.7759/cureus.55672)
- 87. Chow, K.M.; Hui, A.C.; Szeto, C.C. Neurotoxicity Induced by Beta-Lactam Antibiotics: From Bench to Bedside. *Eur. J. Clin. Microbiol. Infect. Dis.* **2005**, *24*, 649–653. [\[CrossRef\]](https://doi.org/10.1007/s10096-005-0021-y)
- 88. Bhattacharyya, S.; Darby, R.R.; Raibagkar, P.; Gonzalez Castro, L.N.; Berkowitz, A.L. Antibiotic-Associated Encephalopathy. *Neurology* **2016**, *86*, 963–971. [\[CrossRef\]](https://doi.org/10.1212/WNL.0000000000002455)
- 89. Conway, N.; Beck, E.; Somerville, J. Penicillin Encephalopathy. *Postgrad. Med. J.* **1968**, *44*, 891–897. [\[CrossRef\]](https://doi.org/10.1136/pgmj.44.518.891)
- 90. Lerner, P.I.; Smith, H.; Weinstein, L. Penicillin Neurotoxicity. *Ann. N. Y. Acad. Sci.* **1967**, *145*, 310–318. [\[CrossRef\]](https://doi.org/10.1111/j.1749-6632.1967.tb50228.x)
- 91. Grill, M.F.; Maganti, R.K. Neurotoxic Effects Associated with Antibiotic Use: Management Considerations. *Br. J. Clin. Pharmacol.* **2011**, *72*, 381–393. [\[CrossRef\]](https://doi.org/10.1111/j.1365-2125.2011.03991.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/21501212)
- 92. Jiménez-Jiménez, F.J.; Puertas, I.; de Toledo-Heras, M. Drug-Induced Myoclonus: Frequency, Mechanisms and Management. *CNS Drugs* **2004**, *18*, 93–104. [\[CrossRef\]](https://doi.org/10.2165/00023210-200418020-00003) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14728056)
- 93. Badhwar, A.; Berkovic, S.F.; Dowling, J.P.; Gonzales, M.; Narayanan, S.; Brodtmann, A.; Berzen, L.; Caviness, J.; Trenkwalder, C.; Winkelmann, J.; et al. Action Myoclonus-Renal Failure Syndrome: Characterization of a Unique Cerebro-Renal Disorder. *Brain* **2004**, *127*, 2173–2182. [\[CrossRef\]](https://doi.org/10.1093/brain/awh263) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15364701)
- 94. Sugimoto, M.; Uchida, I.; Mashimo, T.; Yamazaki, S.; Hatano, K.; Ikeda, F.; Mochizuki, Y.; Terai, T.; Matsuoka, N. Evidence for the Involvement of GABA(A) Receptor Blockade in Convulsions Induced by Cephalosporins. *Neuropharmacology* **2003**, *45*, 304–314. [\[CrossRef\]](https://doi.org/10.1016/S0028-3908(03)00188-6)
- 95. Lam, S.; Gomolin, I.H. Cefepime Neurotoxicity: Case Report, Pharmacokinetic Considerations, and Literature Review. *Pharmacotherapy* **2006**, *26*, 1169–1174. [\[CrossRef\]](https://doi.org/10.1592/phco.26.8.1169)
- 96. Chow, K.M.; Szeto, C.C.; Hui, A.C.-F.; Wong, T.Y.-H.; Li, P.K.-T. Retrospective Review of Neurotoxicity Induced by Cefepime and Ceftazidime. *Pharmacotherapy* **2003**, *23*, 369–373. [\[CrossRef\]](https://doi.org/10.1592/phco.23.3.369.32100)
- 97. Rissardo, J.P.; Caprara, A.L.F. Fluoroquinolone-Associated Movement Disorder: A Literature Review. *Medicines* **2023**, *10*, 33. [\[CrossRef\]](https://doi.org/10.3390/medicines10060033)
- 98. Cannon, J.P.; Lee, T.A.; Clark, N.M.; Setlak, P.; Grim, S.A. The Risk of Seizures among the Carbapenems: A Meta-Analysis. *J. Antimicrob. Chemother.* **2014**, *69*, 2043–2055. [\[CrossRef\]](https://doi.org/10.1093/jac/dku111)
- 99. Kufel, W.D.; Parsels, K.A.; Blaine, B.E.; Steele, J.M.; Seabury, R.W.; Asiago-Reddy, E.A. Real-World Evaluation of Linezolid-Associated Serotonin Toxicity with and without Concurrent Serotonergic Agents. *Int. J. Antimicrob. Agents* **2023**, *62*, 106843. [\[CrossRef\]](https://doi.org/10.1016/j.ijantimicag.2023.106843)
- 100. Neau, J.P.; Rogez, R.; Boissonnot, L.; Simmat, G.; Gil, R.; Lefevre, J.P. Neurologic adverse effects of piperazine. *Acta Neurol. Belg.* **1984**, *84*, 26–34.
- 101. Yagi, S.; Moriya, O.; Nakajima, M.; Umeki, S.; Hino, J.; Soejima, R. A case of tuberculous pleurisy associated with myoclonus and Quincke's edema due to isoniazid and isoniazid sodium methanesulfonate. *Kekkaku* **1989**, *64*, 407–412. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/2796112)
- 102. Moellentin, D.; Picone, C.; Leadbetter, E. Memantine-Induced Myoclonus and Delirium Exacerbated by Trimethoprim. *Ann. Pharmacother.* **2008**, *42*, 443–447. [\[CrossRef\]](https://doi.org/10.1345/aph.1K619) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18303133)
- 103. Pei, L.J.; Tianzhi, I.L.; Lim, W.S. Memantine-Induced Myoclonus Precipitated by Renal Impairment and Drug Interactions. *J. Am. Geriatr. Soc.* **2015**, *63*, 2643–2644. [\[CrossRef\]](https://doi.org/10.1111/jgs.13847) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26691710)
- 104. Kornhuber, J.; Weller, M.; Schoppmeyer, K.; Riederer, P. Amantadine and Memantine Are NMDA Receptor Antagonists with Neuroprotective Properties. *J. Neural Transm. Suppl.* **1994**, *43*, 91–104.
- 105. Rissardo, J.P.; Fornari Caprara, A.L. Action Myoclonus Secondary to Donepezil: Case Report and Literature Review. *Rambam Maimonides Med. J.* **2023**, *14*, e0023. [\[CrossRef\]](https://doi.org/10.5041/RMMJ.10510)
- 106. Whateley, J.M.; Huffman, A.J.; Henderson, E.J. Acute Inability to Mobilise Resulting from Probable Donepezil-Induced Myoclonus. *Age Ageing* **2018**, *47*, 907–908. [\[CrossRef\]](https://doi.org/10.1093/ageing/afy093)
- 107. Tombini, M.; Boscarino, M.; Di Lazzaro, V. Tackling Seizures in Patients with Alzheimer's Disease. *Expert Rev. Neurother.* **2023**, *23*, 1131–1145. [\[CrossRef\]](https://doi.org/10.1080/14737175.2023.2278487)
- 108. Brody, D.J.; Gu, Q. Antidepressant Use Among Adults: United States, 2015–2018. *NCHS Data Brief* **2020**, *377*, 1–8.
- 109. Czeisler, M.É.; Lane, R.I.; Wiley, J.F.; Czeisler, C.A.; Howard, M.E.; Rajaratnam, S.M.W. Follow-up Survey of US Adult Reports of Mental Health, Substance Use, and Suicidal Ideation During the COVID-19 Pandemic, September 2020. *JAMA Netw. Open* **2021**, *4*, e2037665. [\[CrossRef\]](https://doi.org/10.1001/jamanetworkopen.2020.37665)
- 110. Lauterbach, E.C. Reversible Intermittent Rhythmic Myoclonus with Fluoxetine in Presumed Pick's Disease. *Mov. Disord.* **1994**, *9*, 343–346. [\[CrossRef\]](https://doi.org/10.1002/mds.870090314)
- 111. Rissardo, J.P.; Caprara, A.L.F. The Link Between Amitriptyline and Movement Disorders: Clinical Profile and Outcome. *Ann. Acad. Med. Singap.* **2020**, *49*, 236–251. [\[CrossRef\]](https://doi.org/10.47102/annals-acadmed.sg.202023) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32419008)
- 112. Forsberg-Gillving, M.; Bode, M.; Sindrup, S.H. Myoclonus as a side effect to citalopram treatment in a patient with liver cirrhosis. *Ugeskr. Laeger* **2015**, *177*, V04150325.
- 113. Sato, Y.; Nakamura, K.; Yasui-Furukori, N. Serotonin Syndrome Induced by the Readministration of Escitalopram after a Short-Term Interruption in an Elderly Woman with Depression: A Case Report. *Neuropsychiatr. Dis. Treat.* **2015**, *11*, 2505–2507.
- 114. Arora, B.; Kannikeswaran, N. The Serotonin Syndrome-the Need for Physician's Awareness. *Int. J. Emerg. Med.* **2010**, *3*, 373–377. [\[CrossRef\]](https://doi.org/10.1007/s12245-010-0195-7)
- 115. Takahashi, C.; Goto, E.; Taira, S.; Kataoka, N.; Nishihara, M.; Katsumata, T.; Goto, I.; Takiuchi, H. Serotonin syndrome in a patient with small cell lung cancer. *Gan Kagaku Ryoho* **2013**, *40*, 1059–1061.
- 116. Correia, P.; Ribeiro, J.A.; Bento, C.; Sales, F. Negative Myoclonus Secondary to Paroxetine Intake. *BMJ Case Rep.* **2018**, *2018*, bcr-2018-224586. [\[CrossRef\]](https://doi.org/10.1136/bcr-2018-224586)
- 117. Bušková, J.; Vorlová, T.; Piško, J.; Sonka, K. Severe Sleep-Related Movement Disorder Induced by Sertraline. *Sleep Med.* **2012**, *13*, 769–770. [\[CrossRef\]](https://doi.org/10.1016/j.sleep.2012.01.006)
- 118. Trigo López, J.; Martínez Pías, E.; Carrancho García, A.; Pedraza Hueso, M.I. Opsoclonus-myoclonus syndrome secondary to duloxetine poisoning. *Neurologia* **2021**, *36*, 250–252. [\[CrossRef\]](https://doi.org/10.1016/j.nrl.2020.05.009)
- 119. Koshiishi, T.; Okuyama, K. Probable Serotonin Syndrome and Withdrawal Symptoms Caused by Milnacipran. *Yakugaku Zasshi* **2016**, *136*, 1675–1679. [\[CrossRef\]](https://doi.org/10.1248/yakushi.16-00032)
- 120. Necpál, J.; Skorvanek, M. Opsoclonus-Myoclonus Ataxia Syndrome Secondary to Venlafaxine Intoxication. *J. Neurol. Sci.* **2017**, *372*, 19–20. [\[CrossRef\]](https://doi.org/10.1016/j.jns.2016.11.023)
- 121. Sato, F.; Suzuki, A.; Noto, K.; Shirata, T.; Kanno, M.; Kobayashi, R.; Otani, K. Serotonin Syndrome Induced by Overdose of Atomoxetine Alone in a Patient with Attention-Deficit Hyperactivity Disorder: A Case Report. *Psychiatry Clin. Neurosci. Rep.* **2022**, *1*, e41. [\[CrossRef\]](https://doi.org/10.1002/pcn5.41) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38868692)
- 122. Riaz, A.; Ali, H.T.; Allahham, A.; Fornari Caprara, A.L.; Rissardo, J.P. Bupropion-Induced Myoclonus: Case Report and Review of the Literature. *Neurohospitalist* **2023**, *13*, 297–302. [\[CrossRef\]](https://doi.org/10.1177/19418744231173283) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37441201)
- 123. Majewska, M.; Szponar, J.; Pyra, E.; Kostek, H.; Kujawa, A. Serotonin syndrome in the course of drug-poisoning—Case presentation. *Przegl. Lek.* **2011**, *68*, 523–526.
- 124. Hernández, J.L.; Ramos, F.J.; Infante, J.; Rebollo, M.; González-Macías, J. Severe Serotonin Syndrome Induced by Mirtazapine Monotherapy. *Ann. Pharmacother.* **2002**, *36*, 641–643. [\[CrossRef\]](https://doi.org/10.1345/aph.1A302)
- 125. Mason, P.J.; Morris, V.A.; Balcezak, T.J. Serotonin Syndrome. Presentation of 2 Cases and Review of the Literature. *Medicine* **2000**, *79*, 201–209. [\[CrossRef\]](https://doi.org/10.1097/00005792-200007000-00001)
- 126. Patel, H.C.; Bruza, D.; Yeragani, V. Myoclonus with Trazodone. *J. Clin. Psychopharmacol.* **1988**, *8*, 152. [\[CrossRef\]](https://doi.org/10.1097/00004714-198804000-00026)
- 127. Thumtecho, S.; Wainipitapong, S.; Suteparuk, S. Escitalopram, Bupropion, Lurasidone, Lamotrigine and Possible Vortioxetine Overdose Presented with Serotonin Syndrome and Diffuse Encephalopathy: A Case Report. *Toxicol. Rep.* **2021**, *8*, 1846–1848. [\[CrossRef\]](https://doi.org/10.1016/j.toxrep.2021.11.003)
- 128. Bloem, B.R.; Lammers, G.J.; Roofthooft, D.W.; De Beaufort, A.J.; Brouwer, O.F. Clomipramine Withdrawal in Newborns. *Arch. Dis. Child. Fetal Neonatal Ed.* **1999**, *81*, F77. [\[CrossRef\]](https://doi.org/10.1136/fn.81.1.F77a)
- 129. Masand, P. Desipramine-Induced Oral-Pharyngeal Disturbances: Stuttering and Jaw Myoclonus. *J. Clin. Psychopharmacol.* **1992**, *12*, 444–445. [\[CrossRef\]](https://doi.org/10.1097/00004714-199212000-00014)
- 130. Black, K.J.; Kilzieh, N. Severe Imipramine-Induced Myoclonus in a Patient with Psychotic Bipolar Depression, Catatonia, and Schizencephaly. *Ann. Clin. Psychiatry* **1994**, *6*, 45–49. [\[CrossRef\]](https://doi.org/10.3109/10401239409148839)
- 131. Kettl, P.; DePaulo, J.R.J. Maprotiline-Induced Myoclonus. *J. Clin. Psychopharmacol.* **1983**, *3*, 264–265. [\[CrossRef\]](https://doi.org/10.1097/00004714-198308000-00034) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/6886043)
- 132. de Larquier, A.; Vial, T.; Bréjoux, G.; Descotes, J. Serotoninergic syndrome after combining tramadol and iproniazid. *Therapie* **1999**, *54*, 767–768. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10709455)
- 133. Gillman, P.K. Possible Serotonin Syndrome with Moclobemide and Pethidine. *Med. J. Aust.* **1995**, *162*, 554. [\[CrossRef\]](https://doi.org/10.5694/j.1326-5377.1995.tb138527.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7776924)
- 134. White, P.D. Myoclonus and Episodic Delirium Associated with Phenelzine: A Case Report. *J. Clin. Psychiatry* **1987**, *48*, 340–341.
- 135. Rissardo, J.P.; Caprara, A.L.F. Buspirone-Associated Movement Disorder: A Literature Review. *Prague Med. Rep.* **2020**, *121*, 5–24. [\[CrossRef\]](https://doi.org/10.14712/23362936.2020.1)
- 136. Ritchie, E.C.; Bridenbaugh, R.H.; Jabbari, B. Acute Generalized Myoclonus Following Buspirone Administration. *J. Clin. Psychiatry* **1988**, *49*, 242–243.
- 137. Cohen, R.M.; Pickar, D.; Murphy, D.L. Myoclonus-Associated Hypomania during MAO-Inhibitor Treatment. *Am. J. Psychiatry* **1980**, *137*, 105–106.
- 138. Garvey, M.J.; Tollefson, G.D. Occurrence of Myoclonus in Patients Treated with Cyclic Antidepressants. *Arch. Gen. Psychiatry* **1987**, *44*, 269–272. [\[CrossRef\]](https://doi.org/10.1001/archpsyc.1987.01800150081010)
- 139. Revet, A.; Montastruc, F.; Roussin, A.; Raynaud, J.-P.; Lapeyre-Mestre, M.; Nguyen, T.T.H. Antidepressants and Movement Disorders: A Postmarketing Study in the World Pharmacovigilance Database. *BMC Psychiatry* **2020**, *20*, 308. [\[CrossRef\]](https://doi.org/10.1186/s12888-020-02711-z)
- 140. Evidente, V.G.; Caviness, J.N. Focal Cortical Transient Preceding Myoclonus during Lithium and Tricyclic Antidepressant Therapy. *Neurology* **1999**, *52*, 211–213. [\[CrossRef\]](https://doi.org/10.1212/WNL.52.1.211)
- 141. Skidmore, F.; Reich, S.G. Tardive Dystonia. *Curr. Treat. Options Neurol.* **2005**, *7*, 231–236. [\[CrossRef\]](https://doi.org/10.1007/s11940-005-0016-0) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15814076)
- 142. Lu, C.S.; Chu, N.S. Acute Dystonic Reaction with Asterixis and Myoclonus Following Metoclopramide Therapy. *J. Neurol. Neurosurg. Psychiatry* **1988**, *51*, 1002–1003. [\[CrossRef\]](https://doi.org/10.1136/jnnp.51.7.1002-a) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/3204387)
- 143. Harada, T.; Hirosawa, T.; Morinaga, K.; Shimizu, T. Metoclopramide-Induced Serotonin Syndrome. *Intern. Med.* **2017**, *56*, 737–739. [\[CrossRef\]](https://doi.org/10.2169/internalmedicine.56.7727) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28321081)
- 144. Hyser, C.L.; Drake, M.E.J. Myoclonus Induced by Metoclopramide Therapy. *Arch. Intern. Med.* **1983**, *143*, 2201–2202. [\[CrossRef\]](https://doi.org/10.1001/archinte.1983.00350110191040)
- 145. Yung, C.Y. Case Vignettes of Movement Disorders. *Brain Res. Bull.* **1983**, *11*, 191–194. [\[CrossRef\]](https://doi.org/10.1016/0361-9230(83)90190-9)
- 146. Dy, P.; Arcega, V.; Ghali, W.; Wolfe, W. Serotonin Syndrome Caused by Drug to Drug Interaction between Escitalopram and Dextromethorphan. *BMJ Case Rep.* **2017**, *2017*, bcr-2017-221486. [\[CrossRef\]](https://doi.org/10.1136/bcr-2017-221486)
- 147. Naguy, A. Ondansetron-Induced Myoclonus With Escitalopram and Highly Active Antiretroviral Therapy: A Closer Look at 5-HT3 Receptors. *Prim. Care Companion CNS Disord.* **2020**, *22*, 19l02524. [\[CrossRef\]](https://doi.org/10.4088/PCC.19l02524)
- 148. Chaw, S.H.; Chan, L.; Lee, P.K.; Bakar, J.A.; Rasiah, R.; Foo, L.L. Prolonged Drug-Induced Myoclonus: Is It Related to Palonosetron? *J. Anesth.* **2016**, *30*, 1063–1066. [\[CrossRef\]](https://doi.org/10.1007/s00540-016-2228-8)
- 149. Rojas, C.; Stathis, M.; Thomas, A.G.; Massuda, E.B.; Alt, J.; Zhang, J.; Rubenstein, E.; Sebastiani, S.; Cantoreggi, S.; Snyder, S.H.; et al. Palonosetron Exhibits Unique Molecular Interactions with the 5-HT3 Receptor. *Anesth. Analg.* **2008**, *107*, 469–478. [\[CrossRef\]](https://doi.org/10.1213/ane.0b013e318172fa74)
- 150. Van Sweden, B.; Kamphuisen, H.A. Cimetidine Neurotoxicity. EEG and Behaviour Aspects. *Eur. Neurol.* **1984**, *23*, 300–305. [\[CrossRef\]](https://doi.org/10.1159/000115746)
- 151. Irioka, T.; Machida, A.; Yokota, T.; Mizusawa, H. Antihistamine-Associated Myoclonus: A Case Report. *Mov. Disord.* **2008**, *23*, 1615–1616. [\[CrossRef\]](https://doi.org/10.1002/mds.22076) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/18581470)
- 152. Jacquesson, M.; Saudeau, D.; Pantin, B.; Girard, J.J.; Groussin, P. Myoclonia caused by a combination of triprolidine, pseudoephedrine and paracetamol. *Nouv. Presse Med.* **1982**, *11*, 2298–2299. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7110997)
- 153. Schipior, P.G. An Unusual Case of Antihistamine Intoxication. *J. Pediatr.* **1967**, *71*, 589–591. [\[CrossRef\]](https://doi.org/10.1016/S0022-3476(67)80115-X) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/4227701)
- 154. Ammenti, A.; Reitter, B.; Müller-Wiefel, D.E. Chlorambucil Neurotoxicity: Report of Two Cases. *Helv. Paediatr. Acta* **1980**, *35*, 281–287. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7410114)
- 155. Wyllie, A.R.; Bayliff, C.D.; Kovacs, M.J. Myoclonus Due to Chlorambucil in Two Adults with Lymphoma. *Ann. Pharmacother.* **1997**, *31*, 171–174. [\[CrossRef\]](https://doi.org/10.1177/106002809703100207)
- 156. Byrne, T.N.J.; Moseley, T.A., 3rd; Finer, M.A. Myoclonic Seizures Following Chlorambucil Overdose. *Ann. Neurol.* **1981**, *9*, 191–194. [\[CrossRef\]](https://doi.org/10.1002/ana.410090215)
- 157. Meloni, G.; Raucci, U.; Pinto, R.M.; Spalice, A.; Vignetti, M.; Iannetti, P. Pretransplant Conditioning with Busulfan and Cyclophosphamide in Acute Leukemia Patients: Neurological and Electroencephalographic Prospective Study. *Ann. Oncol.* **1992**, *3*, 145–148. [\[CrossRef\]](https://doi.org/10.1093/oxfordjournals.annonc.a058131)
- 158. Savica, R.; Rabinstein, A.A.; Josephs, K.A. Ifosfamide Associated Myoclonus-Encephalopathy Syndrome. *J. Neurol.* **2011**, *258*, 1729–1731. [\[CrossRef\]](https://doi.org/10.1007/s00415-011-5990-4)
- 159. Denison, D.J.; Alghzaly, A.A. Busulfan Induced Myoclonus. *Saudi Med. J.* **2006**, *27*, 557–558.
- 160. Maller, B.; Peguero, E.; Tanvetyanon, T. Ipilimumab/Nivolumab-Related Opsoclonus-Myoclonus-Ataxia Syndrome Variant in a Patient with Malignant Pleural Mesothelioma. *J. Immunother.* **2018**, *41*, 411–412. [\[CrossRef\]](https://doi.org/10.1097/CJI.0000000000000228)
- 161. Raskin, J.; Masrori, P.; Cant, A.; Snoeckx, A.; Hiddinga, B.; Kohl, S.; Janssens, A.; Cras, P.; Van Meerbeeck, J.P. Recurrent Dysphasia Due to Nivolumab-Induced Encephalopathy with Presence of Hu Autoantibody. *Lung Cancer* **2017**, *109*, 74–77. [\[CrossRef\]](https://doi.org/10.1016/j.lungcan.2017.05.002) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28577954)
- 163. Blank, C.U.; Enk, A. Therapeutic Use of Anti-CTLA-4 Antibodies. *Int. Immunol.* **2015**, *27*, 3–10. [\[CrossRef\]](https://doi.org/10.1093/intimm/dxu076)
- 164. Monnerat, C.; Gander, M.; Leyvraz, S. A Rare Case of Prednimustine-Induced Myoclonus. *J. Natl. Cancer Inst.* **1997**, *89*, 173–174. [\[CrossRef\]](https://doi.org/10.1093/jnci/89.2.173) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8998190)
- 165. Lazar, A.; Mau-Holzmann, U.A.; Kolb, H.; Reichenmiller, H.E.; Riess, O.; Schömig, E. Multiple Organ Failure Due to 5-Fluorouracil Chemotherapy in a Patient with a Rare Dihydropyrimidine Dehydrogenase Gene Variant. *Onkologie* **2004**, *27*, 559–562. [\[CrossRef\]](https://doi.org/10.1159/000081338)
- 166. Taylor, H.G.; Wolf, C.R.; Maitland, C.G. Neurologic Toxicity Associated with Hepatic Artery Infusion HAI of FUdR. *Cancer Chemother. Pharmacol.* **1986**, *17*, 292–293.
- 167. Escoda, L.; López-Guillermo, A.; Formigón, M.; Estrach, T.; Cervantes, F.; Montserrat, E.; Rozman, C. Treatment of various lymphoproliferative syndromes with deoxycoformycin: Results in 6 patients. *Sangre* **1990**, *35*, 421–424.
- 168. Mazumder, M.A.; Gulati, S. Tacrolimus-Induced Focal Myoclonus of Unilateral Hand in a Kidney Transplant Recipient. *Indian J. Nephrol.* **2024**, *34*, 93. [\[CrossRef\]](https://doi.org/10.4103/ijn.ijn_146_23)
- 169. Gijtenbeek, J.M.; van den Bent, M.J.; Vecht, C.J. Cyclosporine Neurotoxicity: A Review. *J. Neurol.* **1999**, *246*, 339–346. [\[CrossRef\]](https://doi.org/10.1007/s004150050360)
- 170. Kang, H.G.; Park, S.K.; Wang, S.J.; Oh, S.-Y.; Ryu, H.U. Opsoclonus-Myoclonus Syndrome Following Long-Term Use of Cyclosporine. *Clin. Toxicol.* **2018**, *56*, 373–376. [\[CrossRef\]](https://doi.org/10.1080/15563650.2017.1375511)
- 171. LoRusso, P.; Foster, B.J.; Poplin, E.; McCormick, J.; Kraut, M.; Flaherty, L.; Heilbrun, L.K.; Valdivieso, M.; Baker, L. Phase I Clinical Trial of Pyrazoloacridine NSC366140 (PD115934). *Clin. Cancer Res.* **1995**, *1*, 1487–1493. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/9815948)
- 172. Shun, Y.-T.; Lai, H.-Y.; Chuang, Y.-T.; Lin, H.-F. Successful Treatment of Irinotecan-Induced Muscle Twitching: A Case Report. *Clin. Med. Insights Case Rep.* **2023**, *16*, 11795476221150354. [\[CrossRef\]](https://doi.org/10.1177/11795476221150354) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36760340)
- 173. Rissardo, J.P.; Fornari Caprara, A.L. Myoclonus Secondary to Amantadine: Case Report and Literature Review. *Clin. Pract.* **2023**, *13*, 830–837. [\[CrossRef\]](https://doi.org/10.3390/clinpract13040075) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37489424)
- 174. Pfeiffer, R.F. Amantadine-Induced "Vocal" Myoclonus. *Mov. Disord.* **1996**, *11*, 104–106. [\[CrossRef\]](https://doi.org/10.1002/mds.870110123) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8771080)
- 175. Lin, I.; Armengou-Garcia, L.; Sasikumar, S.; Kuhlman, G.; Fox, S.H.; Lang, A.E.; Espay, A.J. Amantadine-Induced Craniofacial Myoclonus: Distinctive Iatrogenic Dysarthria in Parkinson's Disease. *Mov. Disord. Clin. Pract.* **2023**, *10*, 1408–1413. [\[CrossRef\]](https://doi.org/10.1002/mdc3.13828)
- 176. Cotzias, G.C.; Papavasiliou, P.S.; Gellene, R. Modification of Parkinsonism—Chronic Treatment with L-Dopa. *N. Engl. J. Med.* **1969**, *280*, 337–345. [\[CrossRef\]](https://doi.org/10.1056/NEJM196902132800701)
- 177. Klawans, H.L.; Goetz, C.; Bergen, D. Levodopa-Induced Myoclonus. *Arch. Neurol.* **1975**, *32*, 330–334. [\[CrossRef\]](https://doi.org/10.1001/archneur.1975.00490470075011)
- 178. Nausieda, P.A.; Weiner, W.J.; Kaplan, L.R.; Weber, S.; Klawans, H.L. Sleep Disruption in the Course of Chronic Levodopa Therapy: An Early Feature of the Levodopa Psychosis. *Clin. Neuropharmacol.* **1982**, *5*, 183–194. [\[CrossRef\]](https://doi.org/10.1097/00002826-198205020-00003)
- 179. Luquin, M.R.; Scipioni, O.; Vaamonde, J.; Gershanik, O.; Obeso, J.A. Levodopa-Induced Dyskinesias in Parkinson's Disease: Clinical and Pharmacological Classification. *Mov. Disord.* **1992**, *7*, 117–124. [\[CrossRef\]](https://doi.org/10.1002/mds.870070204)
- 180. Marconi, R.; Lefebvre-Caparros, D.; Bonnet, A.M.; Vidailhet, M.; Dubois, B.; Agid, Y. Levodopa-Induced Dyskinesias in Parkinson's Disease Phenomenology and Pathophysiology. *Mov. Disord.* **1994**, *9*, 2–12. [\[CrossRef\]](https://doi.org/10.1002/mds.870090103)
- 181. Rissardo, J.P.; Muhammad, S.; Yatakarla, V.; Vora, N.M.; Paras, P.; Caprara, A.L.F. Flapping Tremor: Unraveling Asterixis—A Narrative Review. *Medicina* 2024, 60, 362. [\[CrossRef\]](https://doi.org/10.3390/medicina60030362) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38541088)
- 182. Pandey, S.; Srivanitchapoom, P. Levodopa-Induced Dyskinesia: Clinical Features, Pathophysiology, and Medical Management. *Ann. Indian Acad. Neurol.* **2017**, *20*, 190–198. [\[CrossRef\]](https://doi.org/10.4103/aian.AIAN_239_17) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28904447)
- 183. Vardi, J.; Glaubman, H.; Rabey, J.M.; Streifler, M. Myoclonic Attacks Induced by L-Dopa and Bromocryptin in Parkinson Patients: A Sleep EEG Study. *J. Neurol.* **1978**, *218*, 35–42. [\[CrossRef\]](https://doi.org/10.1007/BF00314716)
- 184. Tandberg, E.; Larsen, J.P.; Karlsen, K. A Community-Based Study of Sleep Disorders in Patients with Parkinson's Disease. *Mov. Disord.* **1998**, *13*, 895–899. [\[CrossRef\]](https://doi.org/10.1002/mds.870130606)
- 185. Hsu, Y.-C.; Yeh, Y.-W. Multidrug Overdose-Induced Myoclonus Complicated by Rhabdomyolysis: Possible Role and Mechanism of Muscle Toxicity of Risperidone. *J. Clin. Pharm. Ther.* **2014**, *39*, 698–700. [\[CrossRef\]](https://doi.org/10.1111/jcpt.12205)
- 186. Onofrj, M.; Thomas, A.; Iacono, D.; Di Iorio, A.; Bonanni, L. Switch-over from Tolcapone to Entacapone in Severe Parkinson's Disease Patients. *Eur. Neurol.* **2001**, *46*, 11–16. [\[CrossRef\]](https://doi.org/10.1159/000050749)
- 187. Cardon-Dunbar, A.; Robertson, T.; Roberts, M.S.; Isbister, G.K. Pramipexole Overdose Associated with Visual Hallucinations, Agitation and Myoclonus. *J. Med. Toxicol.* **2017**, *13*, 343–346. [\[CrossRef\]](https://doi.org/10.1007/s13181-017-0615-7)
- 188. Haddad, P.M.; Dursun, S.M. Neurological Complications of Psychiatric Drugs: Clinical Features and Management. *Hum. Psychopharmacol.* **2008**, *23* (Suppl. S1), 15–26. [\[CrossRef\]](https://doi.org/10.1002/hup.918)
- 189. Druschky, K.; Bleich, S.; Grohmann, R.; Engel, R.R.; Neyazi, A.; Stübner, S.; Toto, S. Seizure Rates under Treatment with Antipsychotic Drugs: Data from the AMSP Project. *World J. Biol. Psychiatry* **2019**, *20*, 732–741. [\[CrossRef\]](https://doi.org/10.1080/15622975.2018.1500030)
- 190. Pai, N.; Acar, M.; Juneja, P.; Kouhkamari, M.H.; Siva, S.; Mullan, J. Antipsychotic Prescribing Patterns in Australia: A Retrospective Analysis. *BMC Psychiatry* **2022**, *22*, 110. [\[CrossRef\]](https://doi.org/10.1186/s12888-022-03755-z)
- 191. Hayase, T.; Saiga, H.; Yamaguchi, T. Haloperidol-Induced Myoclonus in a Patient with Delirium. *Geriatr. Gerontol. Int.* **2023**, *23*, 243–244. [\[CrossRef\]](https://doi.org/10.1111/ggi.14550) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36709514)
- 192. Altıparmak, T.; Yurtseven, C.H.; Geniş, B.; Cosar, B. Myoclonic Seizures Induced by Antipsychotic Drugs: A Case Series and Literature Review. *Turk. J. Clin. Psychiatry* **2022**, *25*, 219–222. [\[CrossRef\]](https://doi.org/10.5505/kpd.2022.48254)
- 193. Berman, I.; Zalma, A.; DuRand, C.J.; Green, A.I. Clozapine-Induced Myoclonic Jerks and Drop Attacks. *J. Clin. Psychiatry* **1992**, *53*, 329–330.
- 194. Leroy, C.; Sigwald, J. Myoclonus and epilepsy appearing in various patients during the administration of chlorpromazine. *Encephale* **1956**, *45*, 904–909. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/13384320)
- 195. Tikka, S.K.; Pratap, A.; Sinha, V.K. Dose-Dependent Olanzapine-Induced Myoclonus. *Toxicol. Int.* **2014**, *21*, 335–336. [\[CrossRef\]](https://doi.org/10.4103/0971-6580.155393)
- 196. Ishida, T.; Uchida, H.; Kaneko, S.; Sugiyama, K.; Hamabe, Y.; Mimura, M.; Suzuki, T. Life-Threatening Serotonin Syndrome Precipitated by Discontinuation of Serotonin-Dopamine Antagonist in the Presence of Serotonergic Agents: A Case Report. *Clin. Neuropharmacol.* **2020**, *43*, 81–83. [\[CrossRef\]](https://doi.org/10.1097/WNF.0000000000000385)
- 197. Aggarwal, A.; Jiloha, R.C. Quetiapine Induced Myoclonus. *Indian J. Med. Sci.* **2008**, *62*, 422–423. [\[CrossRef\]](https://doi.org/10.4103/0019-5359.44024)
- 198. Zand, L.; Hoffman, S.J.; Nyman, M.A. 74-Year-Old Woman with New-Onset Myoclonus. *Mayo Clin. Proc.* **2010**, *85*, 955–958. [\[CrossRef\]](https://doi.org/10.4065/mcp.2009.0572)
- 199. Asahi, S.; Nishikawa, T.; Kurata, K.; Morinobu, S.; Yamawaki, S. A Case of Myclonus, Resembling Epileptic Seizure, Induced by Short-Term Sulpiride Treatment. *Int. J. Psychiatry Clin. Pract.* **2002**, *6*, 215–216. [\[CrossRef\]](https://doi.org/10.1080/136515002761581027)
- 200. Montaz, L.; Varache, N.; Harry, P.; Aymes, C.; Turcant, A.; Delille, F.; Simonin, D.; Hass, C. Torsades de pointes during sultopride poisoning. *J. Toxicol. Clin. Exp.* **1992**, *12*, 481–486.
- 201. Pisani, F.; Oteri, G.; Costa, C.; Di Raimondo, G.; Di Perri, R. Effects of Psychotropic Drugs on Seizure Threshold. *Drug Saf.* **2002**, *25*, 91–110. [\[CrossRef\]](https://doi.org/10.2165/00002018-200225020-00004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11888352)
- 202. Gopaul, M.; Altalib, H. Do Psychotropic Drugs Cause Seizures? *Epilepsy Behav. Rep.* **2024**, *27*, 100679. [\[CrossRef\]](https://doi.org/10.1016/j.ebr.2024.100679) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38881884)
- 203. Wong, J.; Delva, N. Clozapine-Induced Seizures: Recognition and Treatment. *Can. J. Psychiatry* **2007**, *52*, 457–463. [\[CrossRef\]](https://doi.org/10.1177/070674370705200708) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/17688010)
- 204. Tominaga, H.; Fukuzako, H.; Izumi, K.; Koja, T.; Fukuda, T.; Fujii, H.; Matsumoto, K.; Sonoda, H.; Imamura, K. Tardive Myoclonus. *Lancet* **1987**, *1*, 322. [\[CrossRef\]](https://doi.org/10.1016/S0140-6736(87)92042-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/2880125)
- 205. Fukuzako, H.; Tominaga, H.; Izumi, K.; Koja, T.; Nomoto, M.; Hokazono, Y.; Kamei, K.; Fujii, H.; Fukuda, T.; Matsumoto, K. Postural Myoclonus Associated with Long-Term Administration of Neuroleptics in Schizophrenic Patients. *Biol. Psychiatry* **1990**, *27*, 1116–1126. [\[CrossRef\]](https://doi.org/10.1016/0006-3223(90)90048-7)
- 206. Ortí-Pareja, M.; Jiménez-Jiménez, F.J.; Vázquez, A.; Catalán, M.J.; Zurdo, M.; Burguera, J.A.; Martínez-Martín, P.; Molina, J.A. Drug-Induced Tardive Syndromes. *Park. Relat. Disord.* **1999**, *5*, 59–65. [\[CrossRef\]](https://doi.org/10.1016/S1353-8020(99)00015-2)
- 207. Little, J.T.; Jankovic, J. Tardive Myoclonus. *Mov. Disord.* **1987**, *2*, 307–311. [\[CrossRef\]](https://doi.org/10.1002/mds.870020408)
- 208. Staedt, J.; Dewes, D.; Danos, P.; Stoppe, G. Can Chronic Neuroleptic Treatment Promote Sleep Disturbances in Elderly Schizophrenic Patients? *Int. J. Geriatr. Psychiatry* **2000**, *15*, 170–176. [\[CrossRef\]](https://doi.org/10.1002/(SICI)1099-1166(200002)15:2%3C170::AID-GPS88%3E3.0.CO;2-H)
- 209. Uvais, N.A.; Ashfaq, A.M. Very Low Single-Dose Quetiapine-Induced Myoclonus. *Prim. Care Companion CNS Disord.* **2022**, *24*, 21cr02907. [\[CrossRef\]](https://doi.org/10.4088/PCC.21cr02907)
- 210. Baysal Kirac, L.; Aydogdu, I.; Acarer, A.; Alpaydin, S.; Bayam, F.E.; Onbasi, H.; Bademkiran, F. Myoclonic Status Epilepticus in Six Patients without Epilepsy. *Epilepsy Behav. Case Rep.* **2013**, *1*, 10–13. [\[CrossRef\]](https://doi.org/10.1016/j.ebcr.2012.10.003)
- 211. Velayudhan, L.; Kirchner, V. Quetiapine-Induced Myoclonus. *Int. Clin. Psychopharmacol.* **2005**, *20*, 119–120. [\[CrossRef\]](https://doi.org/10.1097/00004850-200503000-00011) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15729090)
- 212. Magaudda, A.; Di Rosa, G. Carbamazepine-Induced Non-Epileptic Myoclonus and Tic-like Movements. *Epileptic Disord.* **2012**, *14*, 172–173. [\[CrossRef\]](https://doi.org/10.1684/epd.2012.0504) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/22584239)
- 213. Aguglia, U.; Zappia, M.; Quattrone, A. Carbamazepine-Induced Nonepileptic Myoclonus in a Child with Benign Epilepsy. *Epilepsia* **1987**, *28*, 515–518. [\[CrossRef\]](https://doi.org/10.1111/j.1528-1157.1987.tb03680.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/3653053)
- 214. Dhuna, A.; Pascual-Leone, A.; Talwar, D. Exacerbation of Partial Seizures and Onset of Nonepileptic Myoclonus with Carbamazepine. *Epilepsia* **1991**, *32*, 275–278. [\[CrossRef\]](https://doi.org/10.1111/j.1528-1157.1991.tb05255.x)
- 215. Holtmann, M.; Korn-Merker, E.; Boenigk, H.E. Carbamazepine-Induced Combined Phonic and Motor Tic in a Boy with Down's Syndrome. *Epileptic Disord.* **2000**, *2*, 39–40. [\[CrossRef\]](https://doi.org/10.1684/j.1950-6945.2000.tb00348.x)
- 216. Parmeggiani, L.; Seri, S.; Bonanni, P.; Guerrini, R. Electrophysiological Characterization of Spontaneous and Carbamazepine-Induced Epileptic Negative Myoclonus in Benign Childhood Epilepsy with Centro-Temporal Spikes. *Clin. Neurophysiol.* **2004**, *115*, 50–58. [\[CrossRef\]](https://doi.org/10.1016/S1388-2457(03)00327-4)
- 217. Sáenz-Farret, M.; Tijssen, M.A.J.; Eliashiv, D.; Fisher, R.S.; Sethi, K.; Fasano, A. Antiseizure Drugs and Movement Disorders. *CNS Drugs* **2022**, *36*, 859–876. [\[CrossRef\]](https://doi.org/10.1007/s40263-022-00937-x)
- 218. Rissardo, J.P.; Caprara, A.L.F. Carbamazepine-, Oxcarbazepine-, Eslicarbazepine-Associated Movement Disorder: A Literature Review. *Clin. Neuropharmacol.* **2020**, *43*, 66–80. [\[CrossRef\]](https://doi.org/10.1097/WNF.0000000000000387)
- 219. Rissardo, J.P.; Fornari Caprara, A.L. Cenobamate (YKP3089) and Drug-Resistant Epilepsy: A Review of the Literature. *Medicina* **2023**, *59*, 1389. [\[CrossRef\]](https://doi.org/10.3390/medicina59081389)
- 220. Wallace, S.J. Myoclonus and Epilepsy in Childhood: A Review of Treatment with Valproate, Ethosuximide, Lamotrigine and Zonisamide. *Epilepsy Res.* **1998**, *29*, 147–154. [\[CrossRef\]](https://doi.org/10.1016/S0920-1211(97)00080-6)
- 221. Rissardo, J.P.; Medeiros Araujo de Matos, U.; Fornari Caprara, A.L. Gabapentin-Associated Movement Disorders: A Literature Review. *Medicines* **2023**, *10*, 52. [\[CrossRef\]](https://doi.org/10.3390/medicines10090052) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37755242)
- 222. Birnbaum, D.; Koubeissi, M. Unmasking of Myoclonus by Lacosamide in Generalized Epilepsy. *Epilepsy Behav. Case Rep.* **2017**, *7*, 28–30. [\[CrossRef\]](https://doi.org/10.1016/j.ebcr.2016.09.006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28239547)
- 223. Rissardo, J.P.; Fornari Caprara, A.L. Lamotrigine-Associated Movement Disorder: A Literature Review. *Neurol. India* **2021**, *69*, 1524–1538. [\[CrossRef\]](https://doi.org/10.4103/0028-3886.333440) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34979637)
- 224. Thomas, B.; Frucht, S.J. Myoclonus: An Update. *Curr. Opin. Neurol.* **2024**, *37*, 421–425. [\[CrossRef\]](https://doi.org/10.1097/WCO.0000000000001276)
- 225. Rosen, A.D.; Berenyi, K.J.; Laurenceau, V. Intention Myoclonus—Diazepam and Phenobarbital Treatment. *JAMA* **1969**, *209*, 772–773. [\[CrossRef\]](https://doi.org/10.1001/jama.1969.03160180118020)
- 226. Rissardo, J.P.; Caprara, A.L.F. Phenytoin-Associated Movement Disorder: A Literature Review. *Tzu Chi Med. J.* **2022**, *34*, 409–417. [\[CrossRef\]](https://doi.org/10.4103/tcmj.tcmj_74_22)
- 227. Rissardo, J.P.; Caprara, A.L.F. Pregabalin-Associated Movement Disorders: A Literature Review. *Brain Circ.* **2020**, *6*, 96–106. [\[CrossRef\]](https://doi.org/10.4103/bc.bc_57_19)
- 228. Obeso, J.A.; Artieda, J.; Rothwell, J.C.; Day, B.; Thompson, P.; Marsden, C.D. The Treatment of Severe Action Myoclonus. *Brain* **1989**, *112 Pt 3*, 765–777. [\[CrossRef\]](https://doi.org/10.1093/brain/112.3.765)
- 229. Wallace, S.J. Newer Antiepileptic Drugs: Advantages and Disadvantages. *Brain Dev.* **2001**, *23*, 277–283. [\[CrossRef\]](https://doi.org/10.1016/S0387-7604(01)00230-3)
- 230. Rissardo, J.P.; Caprara, A.L.F. Topiramate-Associated Movement Disorder: Case Series and Literature Review. *Clin. Neuropharmacol.* **2020**, *43*, 116–120. [\[CrossRef\]](https://doi.org/10.1097/WNF.0000000000000395)
- 231. Rissardo, J.P.; Caprara, A.L.F.; Durante, Í. Valproate-Associated Movement Disorder: A Literature Review. *Prague Med. Rep.* **2021**, *122*, 140–180. [\[CrossRef\]](https://doi.org/10.14712/23362936.2021.14) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34606429)
- 232. Neufeld, M.Y.; Vishnevska, S. Vigabatrin and Multifocal Myoclonus in Adults with Partial Seizures. *Clin. Neuropharmacol.* **1995**, *18*, 280–283. [\[CrossRef\]](https://doi.org/10.1097/00002826-199506000-00010) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8635188)
- 233. Warstler, A.; Bean, J. Antimicrobial-Induced Cognitive Side Effects. *Ment. Health Clin.* **2016**, *6*, 207–214. [\[CrossRef\]](https://doi.org/10.9740/mhc.2016.07.207) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29955472)
- 234. Brandariz-Nuñez, D.; Correas-Sanahuja, M.; Maya-Gallego, S.; Martín Herranz, I. Neurotoxicity Associated with Acyclovir and Valacyclovir: A Systematic Review of Cases. *J. Clin. Pharm. Ther.* **2021**, *46*, 918–926. [\[CrossRef\]](https://doi.org/10.1111/jcpt.13464) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34146428)
- 235. Haefeli, W.E.; Schoenenberger, R.A.; Weiss, P.; Ritz, R.F. Acyclovir-Induced Neurotoxicity: Concentration-Side Effect Relationship in Acyclovir Overdose. *Am. J. Med.* **1993**, *94*, 212–215. [\[CrossRef\]](https://doi.org/10.1016/0002-9343(93)90186-S)
- 236. Ernst, M.E.; Franey, R.J. Acyclovir- and Ganciclovir-Induced Neurotoxicity. *Ann. Pharmacother.* **1998**, *32*, 111–113. [\[CrossRef\]](https://doi.org/10.1345/aph.17135)
- 237. Vilter, R.W. Vidarabine-Associated Encephalopathy and Myoclonus. *Antimicrob. Agents Chemother.* **1986**, *29*, 933–935. [\[CrossRef\]](https://doi.org/10.1128/AAC.29.5.933)
- 238. Goldberg, R.J.; Huk, M. Serotonin Syndrome from Trazodone and Buspirone. *Psychosomatics* **1992**, *33*, 235–236. [\[CrossRef\]](https://doi.org/10.1016/S0033-3182(92)72007-6)
- 239. Roth, B.A.; Vinson, D.R.; Kim, S. Carisoprodol-Induced Myoclonic Encephalopathy. *J. Toxicol. Clin. Toxicol.* **1998**, *36*, 609–612. [\[CrossRef\]](https://doi.org/10.3109/15563659809028058)
- 240. Lee, D.S.; Wong, H.A.; Knoppert, D.C. Myoclonus Associated with Lorazepam Therapy in Very-Low-Birth-Weight Infants. *Biol. Neonate* **1994**, *66*, 311–315. [\[CrossRef\]](https://doi.org/10.1159/000244123)
- 241. Li, Y.; Delcher, C.; Brown, J.D.; Wei, Y.-J.; Reisfield, G.M.; Winterstein, A.G. Impact of Schedule IV Controlled Substance Classification on Carisoprodol Utilization in the United States: An Interrupted Time Series Analysis. *Drug Alcohol Depend.* **2019**, *202*, 172–177. [\[CrossRef\]](https://doi.org/10.1016/j.drugalcdep.2019.05.025) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31352307)
- 242. Cepeda, C.; Valin, A.; Calderazzo, L.; Stutzmann, J.M.; Naquet, R. Myoclonus induced by some benzodiazepines in the Papio papio. Comparison with myoclonus induced by intermittent light stimulation. *Rev. Electroencephalogr. Neurophysiol. Clin.* **1982**, *12*, 32–37. [\[CrossRef\]](https://doi.org/10.1016/S0370-4475(82)80006-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/6125000)
- 243. Pitton Rissardo, J.; Fornari Caprara, A.L. Bupropion-Associated Movement Disorders: A Systematic Review. *Ann. Mov. Disord.* **2020**, *3*, 86–98. [\[CrossRef\]](https://doi.org/10.4103/AOMD.AOMD_35_19)
- 244. Pedro-Botet, M.L.; Bonal, J.; Caralps, A. Nifedipine and Myoclonic Disorders. *Nephron* **1989**, *51*, 281. [\[CrossRef\]](https://doi.org/10.1159/000185303)
- 245. Hicks, C.B.; Abraham, K. Verapamil and Myoclonic Dystonia. *Ann. Intern. Med.* **1985**, *103*, 154. [\[CrossRef\]](https://doi.org/10.7326/0003-4819-103-1-154_1)
- 246. Vadlamudi, L.; Wijdicks, E.F.M. Multifocal Myoclonus Due to Verapamil Overdose. *Neurology* **2002**, *58*, 984. [\[CrossRef\]](https://doi.org/10.1212/WNL.58.6.984)
- 247. Wallace, E.L.; Lingle, K.; Pierce, D.; Satko, S. Amlodipine-Induced Myoclonus. *Am. J. Med.* **2009**, *122*, e7. [\[CrossRef\]](https://doi.org/10.1016/j.amjmed.2008.10.036)
- 248. Swanoski, M.T.; Chen, J.S.; Monson, M.H. Myoclonus Associated with Long-Term Use of Diltiazem. *Am. J. Health Syst. Pharm.* **2011**, *68*, 1707–1710. [\[CrossRef\]](https://doi.org/10.2146/ajhp100704)
- 249. Fernandez, H.H.; Friedman, J.H. Carvedilol-Induced Myoclonus. *Mov. Disord.* **1999**, *14*, 703. [\[CrossRef\]](https://doi.org/10.1002/1531-8257(199907)14:4%3C703::AID-MDS1029%3E3.0.CO;2-G)
- 250. Bo, P.; Patrucco, M.; Savoldi, F. Neuropharmacological Profile of Ketanserin. *Farm. Sci.* **1987**, *42*, 91–99.
- 251. González, L.; Feijóo, M. Myoclonus and angiotensin converting enzyme inhibitors. *Med. Clin.* **2005**, *125*, 398. [\[CrossRef\]](https://doi.org/10.1157/13079177) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16185554)
- 253. Hammadi, S.H.; Hassan, M.A.; Allam, E.A.; Elsharkawy, A.M.; Shams, S.S. Effect of Sacubitril/Valsartan on Cognitive Impairment in Colchicine-Induced Alzheimer's Model in Rats. *Fundam. Clin. Pharmacol.* **2023**, *37*, 275–286. [\[CrossRef\]](https://doi.org/10.1111/fcp.12837)
- 254. Marin, C.; Chase, T.N. Effects of SCH 32615, an Enkephalinase Inhibitor, on D-1 and D-2 Dopamine Receptor-Mediated Behaviors. *Neuropharmacology* **1995**, *34*, 677–682. [\[CrossRef\]](https://doi.org/10.1016/0028-3908(95)00026-3)
- 255. Ye, X.; Ling, B.; Wu, J.; Wu, S.; Ren, Y.; Zhang, H.; Song, F.; Xuan, Z.; Chen, M. Case Report: Severe Myoclonus Associated with Oral Midodrine Treatment for Hypotension. *Medicine* **2020**, *99*, e21533. [\[CrossRef\]](https://doi.org/10.1097/MD.0000000000021533)
- 256. Lee, A.Y.; Barforoshi, S.; Singh, A.; Shrestha, R.; Ha, J.; Kittleson, M. Dobutamine-Induced Myoclonus in a Patient With Advanced Heart Failure and Chronic Kidney Disease. *JACC Case Rep.* **2024**, *29*, 102255. [\[CrossRef\]](https://doi.org/10.1016/j.jaccas.2024.102255)
- 257. Wierre, L.; Decaudin, B.; Barsumau, J.; Vairon, M.X.; Horrent, S.; Odou, P.; Azar, R. Dobutamine-Induced Myoclonia in Severe Renal Failure. *Nephrol. Dial. Transplant.* **2004**, *19*, 1336–1337. [\[CrossRef\]](https://doi.org/10.1093/ndt/gfh132)
- 258. Wasey, W.; Aziz, I.; Saleh, S.; Manahil, N.; Wasey, N. Tramadol Induced Jerks. *Cureus* **2021**, *13*, e17547. [\[CrossRef\]](https://doi.org/10.7759/cureus.17547)
- 259. Essandoh, S.; Sakae, M.; Miller, J.; Glare, P.A. A Cautionary Tale from Critical Care: Resolution of Myoclonus after Fentanyl Rotation to Hydromorphone. *J. Pain Symptom Manag.* **2010**, *40*, e4–e6. [\[CrossRef\]](https://doi.org/10.1016/j.jpainsymman.2010.08.005)
- 260. Han, P.K.J.; Arnold, R.; Bond, G.; Janson, D.; Abu-Elmagd, K. Myoclonus Secondary to Withdrawal from Transdermal Fentanyl: Case Report and Literature Review. *J. Pain Symptom Manag.* **2002**, *23*, 66–72. [\[CrossRef\]](https://doi.org/10.1016/S0885-3924(01)00370-0)
- 261. Lauterbach, E.C. Hiccup and Apparent Myoclonus after Hydrocodone: Review of the Opiate-Related Hiccup and Myoclonus Literature. *Clin. Neuropharmacol.* **1999**, *22*, 87–92. [\[CrossRef\]](https://doi.org/10.1097/00002826-199903000-00004) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10202603)
- 262. López Pardo, P.; Izquierdo Zamarriego, G. Myoclonus due to fentanyl. Report of a case. *Rev. Esp. Geriatr. Gerontol.* **2016**, *51*, 60–61. [\[CrossRef\]](https://doi.org/10.1016/j.regg.2015.07.006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26318583)
- 263. Sarhill, N.; Davis, M.P.; Walsh, D.; Nouneh, C. Methadone-Induced Myoclonus in Advanced Cancer. *Am. J. Hosp. Palliat. Care* **2001**, *18*, 51–53. [\[CrossRef\]](https://doi.org/10.1177/104990910101800113) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11406880)
- 264. Jacobsen, L.S.; Olsen, A.K.; Sjøgren, P.; Jensen, N.H. Morphine-induced hyperalgesia, allodynia and myoclonus—New side-effects of morphine? *Ugeskr. Laeger* **1995**, *157*, 3307–3310. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/7543228)
- 265. Reutens, D.C.; Stewart-Wynne, E.G. Norpethidine Induced Myoclonus in a Patient with Renal Failure. *J. Neurol. Neurosurg. Psychiatry* **1989**, *52*, 1450–1451. [\[CrossRef\]](https://doi.org/10.1136/jnnp.52.12.1450)
- 266. Lau, F.; Gardiner, M. Oxycodone/Naloxone: An Unusual Adverse Drug Reaction. *Aust. Fam. Physician* **2017**, *46*, 42–43.
- 267. Hochman, M.S. Meperidine-Associated Myoclonus and Seizures in Long-Term Hemodialysis Patients. *Ann. Neurol.* **1983**, *14*, 593. [\[CrossRef\]](https://doi.org/10.1002/ana.410140520)
- 268. Delvaux, B.; Ryckwaert, Y.; Van Boven, M.; De Kock, M.; Capdevila, X. Remifentanil in the Intensive Care Unit: Tolerance and Acute Withdrawal Syndrome after Prolonged Sedation. *Anesthesiology* **2005**, *102*, 1281–1282. [\[CrossRef\]](https://doi.org/10.1097/00000542-200506000-00030)
- 269. Bowdle, T.A.; Rooke, G.A. Postoperative Myoclonus and Rigidity after Anesthesia with Opioids. *Anesth. Analg.* **1994**, *78*, 783–786. [\[CrossRef\]](https://doi.org/10.1213/00000539-199404000-00030)
- 270. Bae, S.Y.; Lee, S.-J. Negative Myoclonus Associated with Tramadol Use. *Yeungnam Univ. J. Med.* **2020**, *37*, 329–331. [\[CrossRef\]](https://doi.org/10.12701/yujm.2020.00108)
- 271. Biedlingmaier, A.J.; Koola, M.M.; Shad, M.U.; John, S.; Reddy, G.G.; Varghese, S.P. A Rare Case of Serotonin Syndrome With Buprenorphine and Other Serotonergic Medications. *J. Clin. Psychopharmacol.* **2023**, *43*, 379–381. [\[CrossRef\]](https://doi.org/10.1097/JCP.0000000000001707) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37235514)
- 272. Smolen, A.; Smolen, T.N.; van de Kamp, J.L. The Effect of Naloxone Administration on Pregnancy-Associated Seizures. *Life Sci.* **1986**, *38*, 1899–1905. [\[CrossRef\]](https://doi.org/10.1016/0024-3205(86)90218-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/3713430)
- 273. Behnoush, B.; Memarian, A.; Teimoory, M. Naltrexone Induced Serotonin Syndrome. *Int. J. Med. Toxicol. Forensic Med.* **2013**, *3*, 64–66.
- 274. McCann, S.; Yaksh, T.L.; von Gunten, C.F. Correlation between Myoclonus and the 3-Glucuronide Metabolites in Patients Treated with Morphine or Hydromorphone: A Pilot Study. *J. Opioid Manag.* **2010**, *6*, 87–94. [\[CrossRef\]](https://doi.org/10.5055/jom.2010.0008)
- 275. Bruera, E.; Pereira, J. Acute Neuropsychiatric Findings in a Patient Receiving Fentanyl for Cancer Pain. *Pain* **1997**, *69*, 199–201. [\[CrossRef\]](https://doi.org/10.1016/S0304-3959(96)03238-1)
- 276. Sprung, J.; Schedewie, H.K. Apparent Focal Motor Seizure with a Jacksonian March Induced by Fentanyl: A Case Report and Review of the Literature. *J. Clin. Anesth.* **1992**, *4*, 139–143. [\[CrossRef\]](https://doi.org/10.1016/0952-8180(92)90031-U)
- 277. Eisele, J.H.J.; Grigsby, E.J.; Dea, G. Clonazepam Treatment of Myoclonic Contractions Associated with High-Dose Opioids: Case Report. *Pain* **1992**, *49*, 231–232. [\[CrossRef\]](https://doi.org/10.1016/0304-3959(92)90146-3)
- 278. Mercadante, S. Dantrolene Treatment of Opioid-Induced Myoclonus. *Anesth. Analg.* **1995**, *81*, 1307–1308.
- 279. Holdsworth, M.T.; Adams, V.R.; Chavez, C.M.; Vaughan, L.J.; Duncan, M.H. Continuous Midazolam Infusion for the Management of Morphine-Induced Myoclonus. *Ann. Pharmacother.* **1995**, *29*, 25–29. [\[CrossRef\]](https://doi.org/10.1177/106002809502900105)
- 280. Kolesnikov, Y.; Jain, S.; Wilson, R.; Pasternak, G.W. Blockade of Morphine-Induced Hindlimb Myoclonic Seizures in Mice by Ketamine. *Pharmacol. Biochem. Behav.* **1997**, *56*, 423–425. [\[CrossRef\]](https://doi.org/10.1016/S0091-3057(96)00221-3)
- 281. Scott, J.C.; Sarnquist, F.H. Seizure-like Movements during a Fentanyl Infusion with Absence of Seizure Activity in a Simultaneous EEG Recording. *Anesthesiology* **1985**, *62*, 812–814. [\[CrossRef\]](https://doi.org/10.1097/00000542-198506000-00024) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/4003808)
- 282. Lane, J.C.; Tennison, M.B.; Lawless, S.T.; Greenwood, R.S.; Zaritsky, A.L. Movement Disorder after Withdrawal of Fentanyl Infusion. *J. Pediatr.* **1991**, *119*, 649–651. [\[CrossRef\]](https://doi.org/10.1016/S0022-3476(05)82421-7) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/1919900)
- 283. Smith, N.T.; Benthuysen, J.L.; Bickford, R.G.; Sanford, T.J.; Blasco, T.; Duke, P.C.; Head, N.; Dec-Silver, H. Seizures during Opioid Anesthetic Induction—Are They Opioid-Induced Rigidity? *Anesthesiology* **1989**, *71*, 852–862. [\[CrossRef\]](https://doi.org/10.1097/00000542-198912000-00008) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/2531560)
- 284. Zai, X. Beyond the Brink: Unraveling the Opioid Crisis and Its Profound Impacts. *Econ. Hum. Biol.* **2024**, *53*, 101379. [\[CrossRef\]](https://doi.org/10.1016/j.ehb.2024.101379)
- 285. Gordon, M.F.; Abrams, R.I.; Rubin, D.B.; Barr, W.B.; Correa, D.D. Bismuth Subsalicylate Toxicity as a Cause of Prolonged Encephalopathy with Myoclonus. *Mov. Disord.* **1995**, *10*, 220–222. [\[CrossRef\]](https://doi.org/10.1002/mds.870100215)
- 286. Borbinha, C.; Serrazina, F.; Salavisa, M.; Viana-Baptista, M. Bismuth Encephalopathy- a Rare Complication of Long-Standing Use of Bismuth Subsalicylate. *BMC Neurol.* **2019**, *19*, 212. [\[CrossRef\]](https://doi.org/10.1186/s12883-019-1437-9)
- 287. Buge, A.; Rancurel, G.; Dechy, H. Bismuth myoclonic encephalopathies. Their course and lasting or definitive late complications. *Rev. Neurol.* **1977**, *133*, 401–415.
- 288. Molina, J.A.; Calandre, L.; Bermejo, F. Myoclonic Encephalopathy Due to Bismuth Salts: Treatment with Dimercaprol and Analysis of CSF Transmitters. *Acta Neurol. Scand.* **1989**, *79*, 200–203. [\[CrossRef\]](https://doi.org/10.1111/j.1600-0404.1989.tb03739.x)
- 289. Rissardo, J.P.; Caprara, A.L.F.; Durante, Í.; Rauber, A. Lithium-Associated Movement Disorder: A Literature Review. *Brain Circ.* **2022**, *8*, 76–86. [\[CrossRef\]](https://doi.org/10.4103/bc.bc_77_21)
- 290. Dyson, E.H.; Simpson, D.; Prescott, L.F.; Proudfoot, A.T. Self-Poisoning and Therapeutic Intoxication with Lithium. *Hum. Toxicol.* **1987**, *6*, 325–329. [\[CrossRef\]](https://doi.org/10.1177/096032718700600410)
- 291. Bender, S.; Linka, T.; Wolstein, J.; Gehendges, S.; Paulus, H.-J.; Schall, U.; Gastpar, M. Safety and Efficacy of Combined Clozapine-Lithium Pharmacotherapy. *Int. J. Neuropsychopharmacol.* **2004**, *7*, 59–63. [\[CrossRef\]](https://doi.org/10.1017/S1461145703003870) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/14731311)
- 292. Naramoto, A.; Koizumi, N.; Itoh, N.; Shigematsu, H. An Autopsy Case of Cerebellar Degeneration Following Lithium Intoxication with Neuroleptic Malignant Syndrome. *Acta Pathol. Jpn.* **1993**, *43*, 55–58. [\[CrossRef\]](https://doi.org/10.1111/j.1440-1827.1993.tb02914.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/8257473)
- 293. Sarrigiannis, P.G.; Zis, P.; Unwin, Z.C.; Blackburn, D.J.; Hoggard, N.; Zhao, Y.; Billings, S.A.; Khan, A.A.; Yianni, J.; Hadjivassiliou, M. Tremor after Long Term Lithium Treatment; Is It Cortical Myoclonus? *Cerebellum Ataxias* **2019**, *6*, 5. [\[CrossRef\]](https://doi.org/10.1186/s40673-019-0100-y)
- 294. Jundt, F.; Lempert, T.; Dörken, B.; Pezzutto, A. Trimethoprim-Sulfamethoxazole Exacerbates Posthypoxic Action Myoclonus in a Patient with Suspicion of Pneumocystis Jiroveci Infection. *Infection* **2004**, *32*, 176–178. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15188079)
- 295. Segal, J.A.; Harris, B.D.; Kustova, Y.; Basile, A.; Skolnick, P. Aminoglycoside Neurotoxicity Involves NMDA Receptor Activation. *Brain Res.* **1999**, *815*, 270–277. [\[CrossRef\]](https://doi.org/10.1016/S0006-8993(98)01123-8)
- 296. Matsunaga, K.; Uozumi, T.; Qingrui, L.; Hashimoto, T.; Tsuji, S. Amantadine-Induced Cortical Myoclonus. *Neurology* **2001**, *56*, 279–280. [\[CrossRef\]](https://doi.org/10.1212/WNL.56.2.279)
- 297. Domínguez, C.; Benito-León, J.; Bermejo-Pareja, F. Multifocal Myoclonus Induced by Haloperidol. *Neurol. Sci.* **2009**, *30*, 385–386. [\[CrossRef\]](https://doi.org/10.1007/s10072-009-0104-0)
- 298. Praharaj, S.K.; Venkatesh, B.G.M.; Sarkhel, S.; Zia-ul-Haq, M.; Sinha, V.K. Clozapine-Induced Myoclonus: A Case Study and Brief Review. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2010**, *34*, 242–243. [\[CrossRef\]](https://doi.org/10.1016/j.pnpbp.2009.10.006)
- 299. Valin, A.; Cepeda, C.; Rey, E.; Naquet, R. Opposite Effects of Lorazepam on Two Kinds of Myoclonus in the Photosensitive Papio Papio. *Electroencephalogr. Clin. Neurophysiol.* **1981**, *52*, 647–651. [\[CrossRef\]](https://doi.org/10.1016/0013-4694(81)91439-5)
- 300. García-Ruiz, P.J.; Javier Jiménez-Jiménez, F.; García de Yébenes, J. Calcium Channel Blocker-Induced Parkinsonism: Clinical Features and Comparisons with Parkinson's Disease. *Park. Relat. Disord.* **1998**, *4*, 211–214. [\[CrossRef\]](https://doi.org/10.1016/S1353-8020(98)00032-7)
- 301. Klawans, H.L.; Carvey, P.M.; Tanner, C.M.; Goetz, C.G. Drug-Induced Myoclonus. *Adv. Neurol.* **1986**, *43*, 251–264. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/3484854)
- 302. Weiner, W.J.; Carvey, P.M.; Nausieda, P.A.; Klawans, H.L. Dopaminergic Antagonism of L-5-Hydroxytryptophan-Induced Myoclonic Jumping Behavior. *Neurology* **1979**, *29*, 1622–1625. [\[CrossRef\]](https://doi.org/10.1212/WNL.29.12.1622) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/41197)
- 303. Giménez-Roldán, S.; Mateo, D.; Muradas, V.; De Yebenes, J.G. Clinical, Biochemical, and Pharmacological Observation in a Patient with Postasphyxic Myoclonus: Association to Serotonin Hyperactivity. *Clin. Neuropharmacol.* **1988**, *11*, 151–160. [\[CrossRef\]](https://doi.org/10.1097/00002826-198804000-00006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/2454156)
- 304. Feighner, J.P.; Boyer, W.F.; Tyler, D.L.; Neborsky, R.J. Adverse Consequences of Fluoxetine-MAOI Combination Therapy. *J. Clin. Psychiatry* **1990**, *51*, 222–225.
- 305. Sternbach, H. The Serotonin Syndrome. *Am. J. Psychiatry* **1991**, *148*, 705–713.
- 306. Eison, A.S.; Wright, R.N.; Freeman, R.P.; Gylys, J.A. 5-HT-Dependent Myoclonus in Guinea Pigs: Mediation through 5-HT1A-5- HT2 Receptor Interaction. *Brain Res. Bull.* **1993**, *30*, 687–689. [\[CrossRef\]](https://doi.org/10.1016/0361-9230(93)90100-P)
- 307. Pappert, E.J.; Goetz, C.G.; Stebbins, G.T.; Belden, M.; Carvey, P.M. 5-Hydroxytryptophan-Induced Myoclonus in Guinea Pigs: Mediation through 5-HT1/2 Receptor Subtypes. *Eur. J. Pharmacol.* **1998**, *347*, 51–56. [\[CrossRef\]](https://doi.org/10.1016/S0014-2999(98)00086-7)
- 308. Termsarasab, P.; Thammongkolchai, T.; Frucht, S.J. Spinal-Generated Movement Disorders: A Clinical Review. *J. Clin. Mov. Disord.* **2015**, *2*, 18. [\[CrossRef\]](https://doi.org/10.1186/s40734-015-0028-1)
- 309. Pal, G.; Lin, M.M.; Laureno, R. Asterixis: A Study of 103 Patients. *Metab. Brain Dis.* **2014**, *29*, 813–824. [\[CrossRef\]](https://doi.org/10.1007/s11011-014-9514-7)
- 310. Hillsley, R.E.; Massey, E.W. Truncal Asterixis Associated with Ceftazidime, a Third-Generation Cephalosporin. *Neurology* **1991**, *41*, 2008. [\[CrossRef\]](https://doi.org/10.1212/WNL.41.12.2008)
- 311. Umemoto, D.; Kuroda, H.; Nishioka, H. Negative Myoclonus as a Manifestation of Cefepime Neurotoxicity. *Clin. Case Rep.* **2024**, *12*, e8380. [\[CrossRef\]](https://doi.org/10.1002/ccr3.8380) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38161642)
- 312. Levine, P.H.; Regelson, W.; Holland, J.F. Chloramphenicol-Associated Encephalopathy. *Clin. Pharmacol. Ther.* **1970**, *11*, 194–199. [\[CrossRef\]](https://doi.org/10.1002/cpt1970112194) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/5416851)
- 313. Zhang, H.; Zhang, J.; Dong, H. Fatal Hepatotoxicity Due to Viaminate. *Am. J. Med. Sci.* **2018**, *356*, 84–86. [\[CrossRef\]](https://doi.org/10.1016/j.amjms.2018.01.001)
- 314. Gray, D.A.; Foo, D. Reversible Myoclonus, Asterixis, and Tremor Associated with High Dose Trimethoprim-Sulfamethoxazole: A Case Report. *J. Spinal Cord. Med.* **2016**, *39*, 115–117. [\[CrossRef\]](https://doi.org/10.1179/2045772315Y.0000000018) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/26111222)
- 315. Kobayashi, I.; Osawa, M.; Ohta, K.; Maruyama, S. L-Dopa-Induced Asterixis. *Folia Psychiatr. Neurol. Jpn.* **1985**, *39*, 507–513. [\[CrossRef\]](https://doi.org/10.1111/j.1440-1819.1985.tb00804.x)
- 316. Rittmannsberger, H. Asterixis Induced by Psychotropic Drug Treatment. *Clin. Neuropharmacol.* **1996**, *19*, 349–355. [\[CrossRef\]](https://doi.org/10.1097/00002826-199619040-00008)
- 317. Rittmannsberger, H.; Leblhuber, F. Asterixis Induced by Carbamazepine Therapy. *Biol. Psychiatry* **1992**, *32*, 364–368. [\[CrossRef\]](https://doi.org/10.1016/0006-3223(92)90040-7)
- 318. Wahba, M.; Waln, O. Asterixis Related to Gabapentin Intake: A Case Report and Review. *Postgrad. Med.* **2013**, *125*, 139–141. [\[CrossRef\]](https://doi.org/10.3810/pgm.2013.09.2696)
- 319. Cerminara, C.; Montanaro, M.L.; Curatolo, P.; Seri, S. Lamotrigine-Induced Seizure Aggravation and Negative Myoclonus in Idiopathic Rolandic Epilepsy. *Neurology* **2004**, *63*, 373–375. [\[CrossRef\]](https://doi.org/10.1212/01.WNL.0000130195.62670.A6)
- 320. Chi, W.M.; Chua, K.S.; Kong, K.H. Phenytoin-Induced Asterixis—Uncommon or under-Diagnozed? *Brain Inj.* **2000**, *14*, 847–850.
- 321. Heckmann, J.G.; Ulrich, K.; Dütsch, M.; Neundörfer, B. Pregabalin Associated Asterixis. *Am. J. Phys. Med. Rehabil.* **2005**, *84*, 724. [\[CrossRef\]](https://doi.org/10.1097/01.phm.0000176355.97155.f5) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16141753)
- 322. Katano, H.; Fukushima, T.; Karasawa, K.; Sugiyama, N.; Ohkura, A.; Kamiya, K. Primidone-Induced Hyperammonemic Encephalopathy in a Patient with Cerebral Astrocytoma. *J. Clin. Neurosci.* **2002**, *9*, 79–81. [\[CrossRef\]](https://doi.org/10.1054/jocn.2001.1011) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11749025)
- 323. Rottach, K.G.; Weiss-Brummer, J.; Wieland, U.; Schmauss, M. Valproic acid in prophylaxis of bipolar disorder. A case of valproate-induced encephalopathy. *Nervenarzt* **2000**, *71*, 401–403. [\[CrossRef\]](https://doi.org/10.1007/s001150050575) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/10846716)
- 324. Rubin, B.; Horowitz, G.; Katz, R.I. Asterixis Following Metrizamide Myelography. *Arch. Neurol.* **1980**, *37*, 522. [\[CrossRef\]](https://doi.org/10.1001/archneur.1980.00500570070012)
- 325. Dysken, M.W.; Comaty, J.E.; Pandey, G.N.; Davis, J.M. Asterixis Associated with a High RBC Lithium Concentration. *Am. J. Psychiatry* **1979**, *136*, 1610. [\[CrossRef\]](https://doi.org/10.1176/ajp.136.12.1610-a)
- 326. Adair, J.C.; Gilmore, R.L. Meperidine Neurotoxicity after Organ Transplantation. *J. Toxicol. Clin. Toxicol.* **1994**, *32*, 325–328. [\[CrossRef\]](https://doi.org/10.3109/15563659409017968)
- 327. Conn, H.O. Asterixis. Its Occurrence in Chronic Pulmonary Disease, with a Commentary on Its General Mechanism. *N. Engl. J. Med.* **1958**, *259*, 564–569. [\[CrossRef\]](https://doi.org/10.1056/NEJM195809182591203)
- 328. Meyer, T.; Ludolph, A.C.; Münch, C. Ifosfamide Encephalopathy Presenting with Asterixis. *J. Neurol. Sci.* **2002**, *199*, 85–88. [\[CrossRef\]](https://doi.org/10.1016/S0022-510X(02)00077-1)
- 329. Drayer, B.P.; Warner, M.A.; Sudilovsky, A.; Luther, J.; Wilkins, R.; Allen, S.; Bates, M. Iopamidol vs Metrizamide: A Double Blind Study for Cervical Myelography. *Neuroradiology* **1982**, *24*, 77–84. [\[CrossRef\]](https://doi.org/10.1007/BF00339195)
- 330. Anderson, R.J. Asterixis as a Manifestation of Salicylate Toxicity. *Ann. Intern. Med.* **1981**, *95*, 188–189. [\[CrossRef\]](https://doi.org/10.7326/0003-4819-95-2-188)
- 331. Uddin, M.F.; Alweis, R.; Shah, S.R.; Lateef, N.; Shahnawaz, W.; Ochani, R.K.; Dharani, A.M.; Shah, S.A. Controversies in Serotonin Syndrome Diagnosis and Management: A Review. *J. Clin. Diagn. Res.* **2017**, *11*, OE05–OE07. [\[CrossRef\]](https://doi.org/10.7860/JCDR/2017/29473.10696) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29207768)
- 332. Finelli, P.F. Drug-Induced Creutzfeldt-Jakob like Syndrome. *J. Psychiatry Neurosci.* **1992**, *17*, 103–105. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/1390620)
- 333. Naro, A.; Pignolo, L.; Billeri, L.; Porcari, B.; Portaro, S.; Tonin, P.; Calabrò, R.S. A Case of Psychogenic Myoclonus Responding to a Novel Transcranial Magnetic Stimulation Approach: Rationale, Feasibility, and Possible Neurophysiological Basis. *Front. Hum. Neurosci.* **2020**, *14*, 292. [\[CrossRef\]](https://doi.org/10.3389/fnhum.2020.00292) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32848667)

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