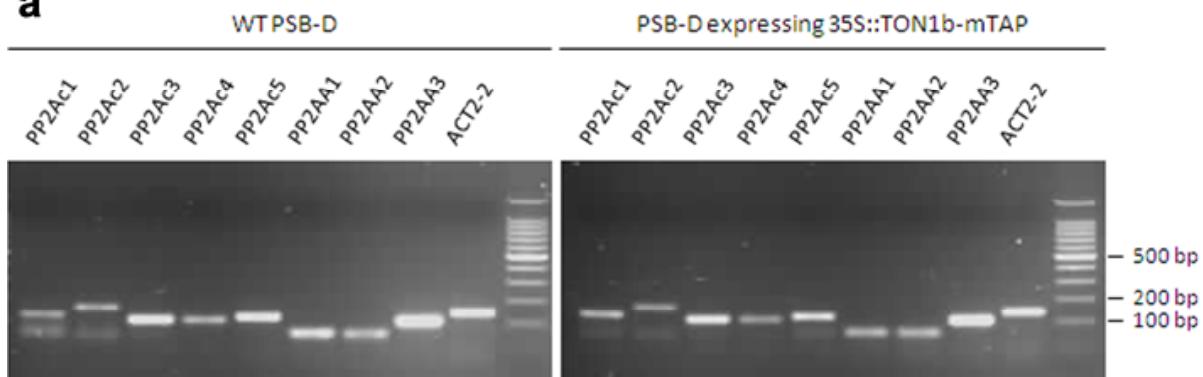
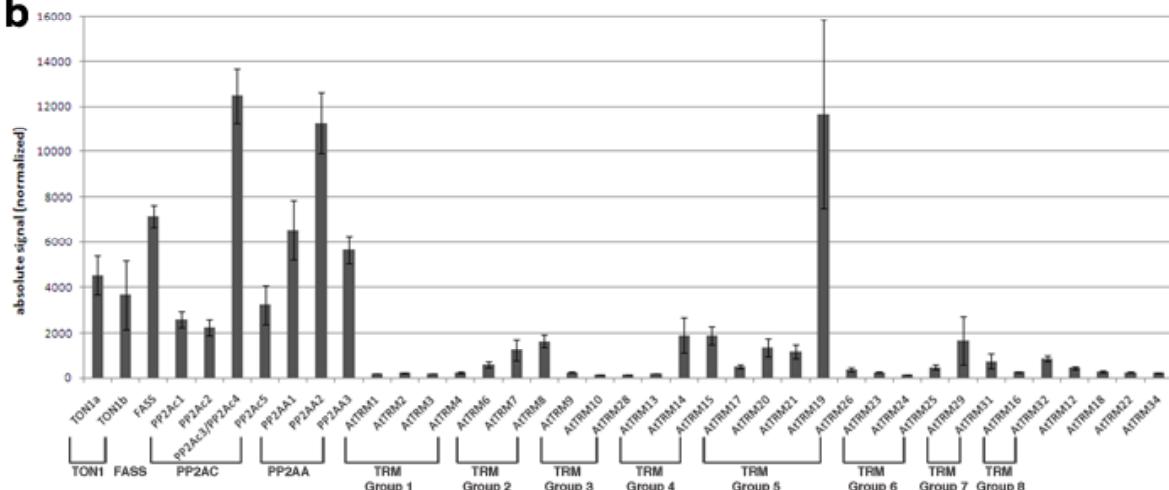


## Supplementary Figure S1

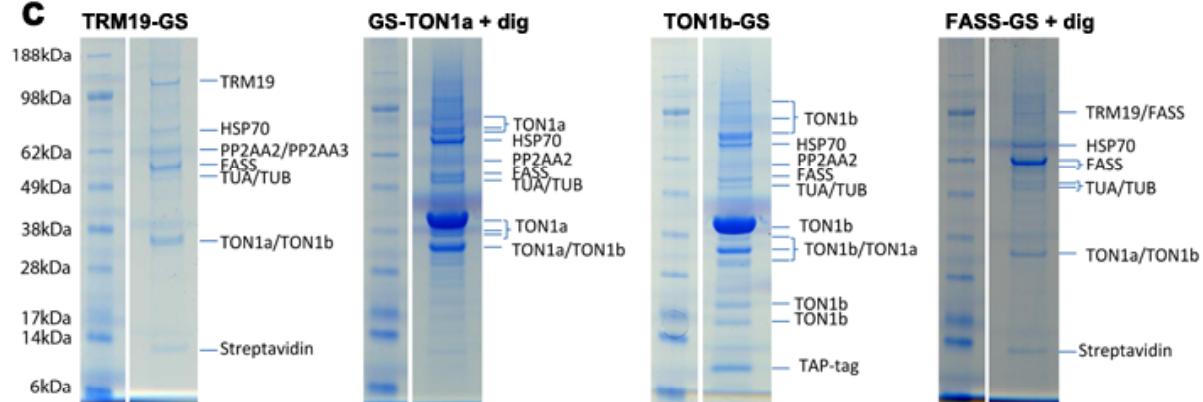
**a**



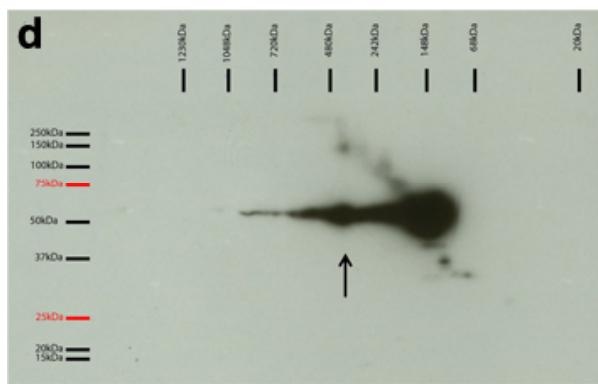
**b**



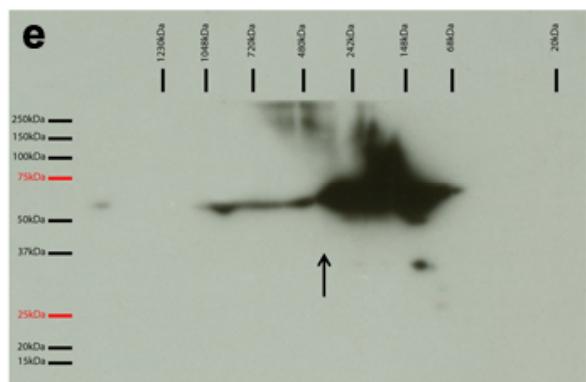
**c**



**d**



**e**

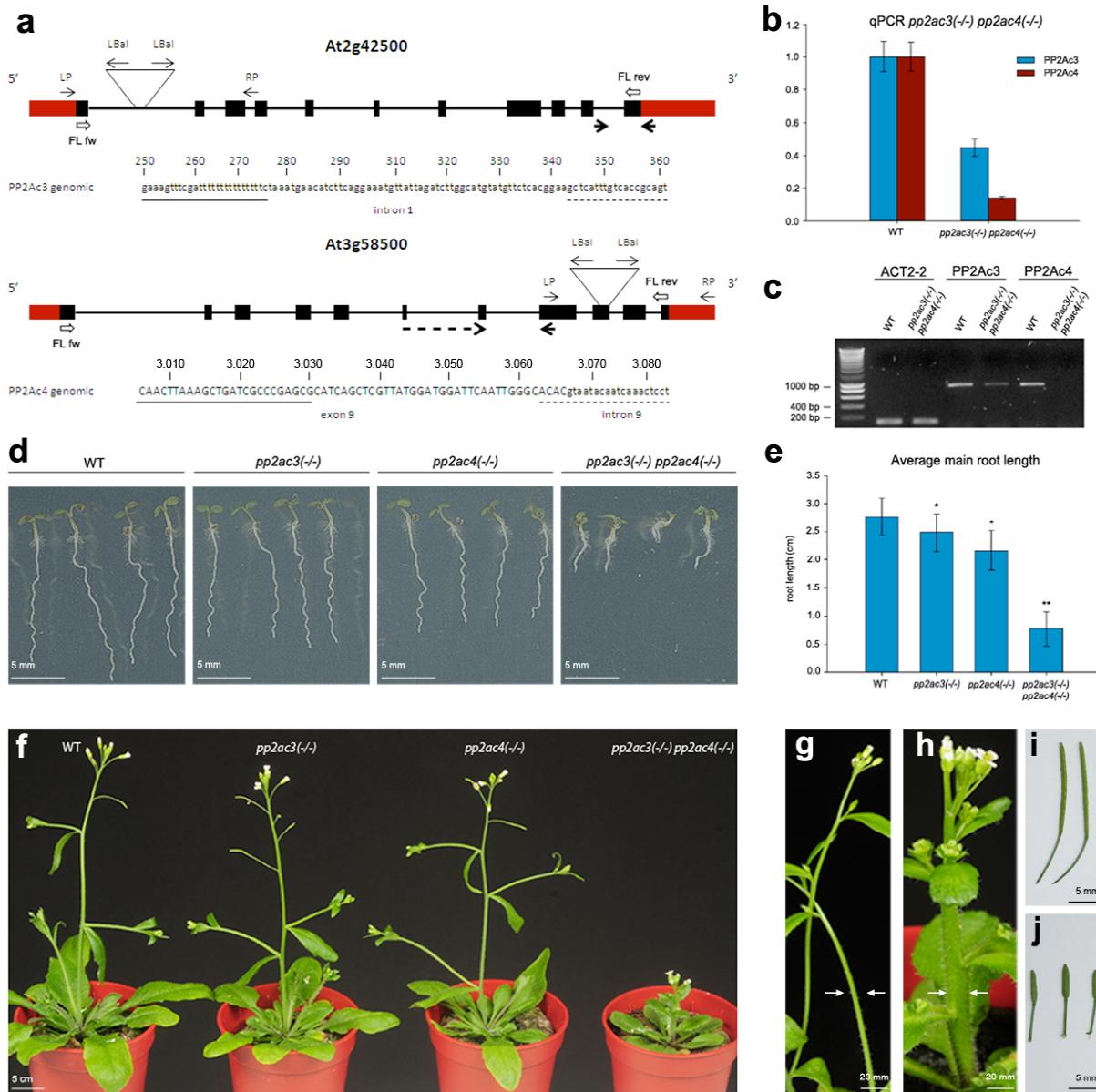


**Supplementary Figure S1 : Expression and TAP analysis of TTP members in *Arabidopsis* cell culture**

- (a) RT-PCR analysis of *Arabidopsis* PP2AA and PP2AC subunits in control cells and in Pro35S:TON1b-mTAP expressing cells.
- (b) Gene expression analysis of the TTP complex genes in *Arabidopsis* cell culture using the Genevestigator tool<sup>61</sup>. No Affymetrix probes are available for the *TRM5*, *TRM11*, *TRM27*, *TRM30* and *TRM33* genes.
- (c) Representative Coomassie stained gels for all bait proteins used in the TAP experiments (TON1a, TON1b, FASS and TRM19) followed by MALDI TOF/TOF analysis. Proteins in the TAP eluates were separated on a 4-12% gradient NuPAGE gel (Invitrogen) and visualized with colloidal Coomassie Brilliant Blue staining. The most abundant proteins detected in each visible band on gel are indicated. In brief, the identified proteins were traced back to their gel slice number via MALDI spot numbers, making it possible to indicate the most abundant identified protein(s) in each visible band on SDS-PAGE. Some of the visible bands on SDS-PAGE correspond to non-specific co-purified proteins like HSP70, TUA/TUB, streptavidin etc... Such background proteins are not presented in the final table of interactors, as known background proteins (obtained from cultures expressing TAP-tagged mock proteins GUS, RFP and GFP) are systematically subtracted from the interactor lists as indicated in the Methods section. Identified proteins which do not give a clear visible band on SDS-PAGE or which are not the most abundant protein in a visible band on the SDS-PAGE gels are not indicated. Left lanes of each panel correspond to molecular weight standards.
- (d-e) 2D Blue Native/SDS-PAGE on total protein extracts of PSB-D *Arabidopsis* cell suspension cells expressing Pro35S::TON1b-GS (d) or Pro35S::GS-TON1a (e) followed by Western blot detection using the Peroxidase Anti-Peroxidase (PAP) antibody that binds the GS-tag. The observed smear indicates incorporation of the GS-fused proteins in high molecular weight range complexes with an accumulation at an estimated size of 300-350 kDa (arrows). The strong signal at around 100kDa likely reflects the migration of uncomplexed TON1 proteins as dimers.

<sup>61</sup>Zimmermann, P., Hirsch-Hoffmann, M., Hennig, L., and Gruissem, W. GENEVESTIGATOR. *Arabidopsis* microarray database and analysis toolbox. *Plant Physiol* **136**, 2621-2632 (2004)

## Supplementary Figure S2

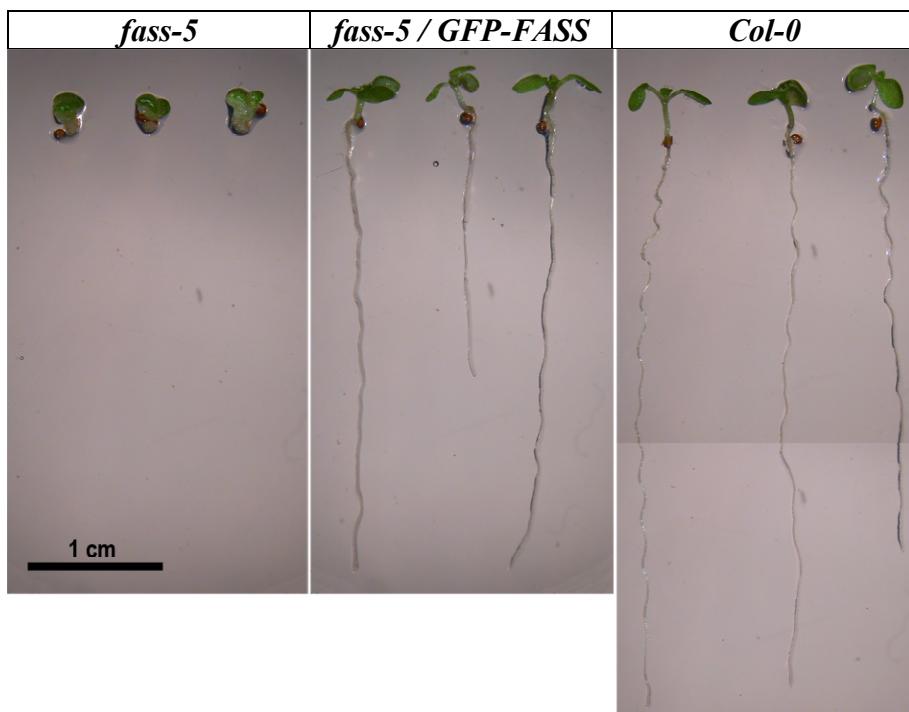


Supplementary Figure S2 : Analysis of *pp2ac3-c4* double mutant

(a) Gene models of *PP2AC3* and *PP2AC4* with indication of the respective T-DNA insertions and primers used for genotyping (LP, RP, LBal), qPCR (bold arrows) and RT-PCR (FL fw and FL rev). Sequencing both the LBal-LP and LBal-RP PCR products allowed us to determine the exact insertion positions (underlined and dashed sequences) and to confirm the presence of tandem repeat T-DNA insertions in both lines. (b) Quantitative PCR (qPCR) analysis between wild type (Col-0) and the *pp2ac3-c4* double mutant showing significant reduction in *PP2AC3* and *PP2AC4* messenger levels. Expression of *PP2AC3* appears less reduced than *PP2AC4*. Both actin and eif4A were used as normalization genes. (c) RT-PCR analysis showing residual expression of full-length *PP2AC3* mRNA, while no full-length *PP2AC4* mRNA could be detected in the *pp2ac3-c4* double mutant. Actin (Act2-2) primers were used for normalization. (d) Representative images of wild type (Col-0), *pp2ac3*, *pp2ac4* and *pp2ac3-c4* double mutant seedlings showing severely affected main root growth of the *pp2ac3-c4* double mutant. (e) Average main root length of wild type (n=96), *pp2ac3* (n=38),

*pp2a-c4* ( $n=35$ ) and the *pp2ac3-ac4* double mutant ( $n=52$ ). Single *pp2ac* mutants already show a statistical reduction in average main root length compared to wild type (T-test,  $P<10^{-4}$  and  $P<10^{-10}$  for *pp2ac3* and *pp2ac4* respectively) and the defects in main root growth are strongly enhanced (T-test,  $P<10^{-60}$ ) in the *pp2ac3-c4* double mutant. (f-j) The *pp2ac3* and *pp2ac4* single mutants develop to wild type looking mature plants while the *pp2ac3-c4* double mutant are dwarfed plant with small, thick and round leaves, thick flowering stems (g, h) and shorter siliques (i, j).

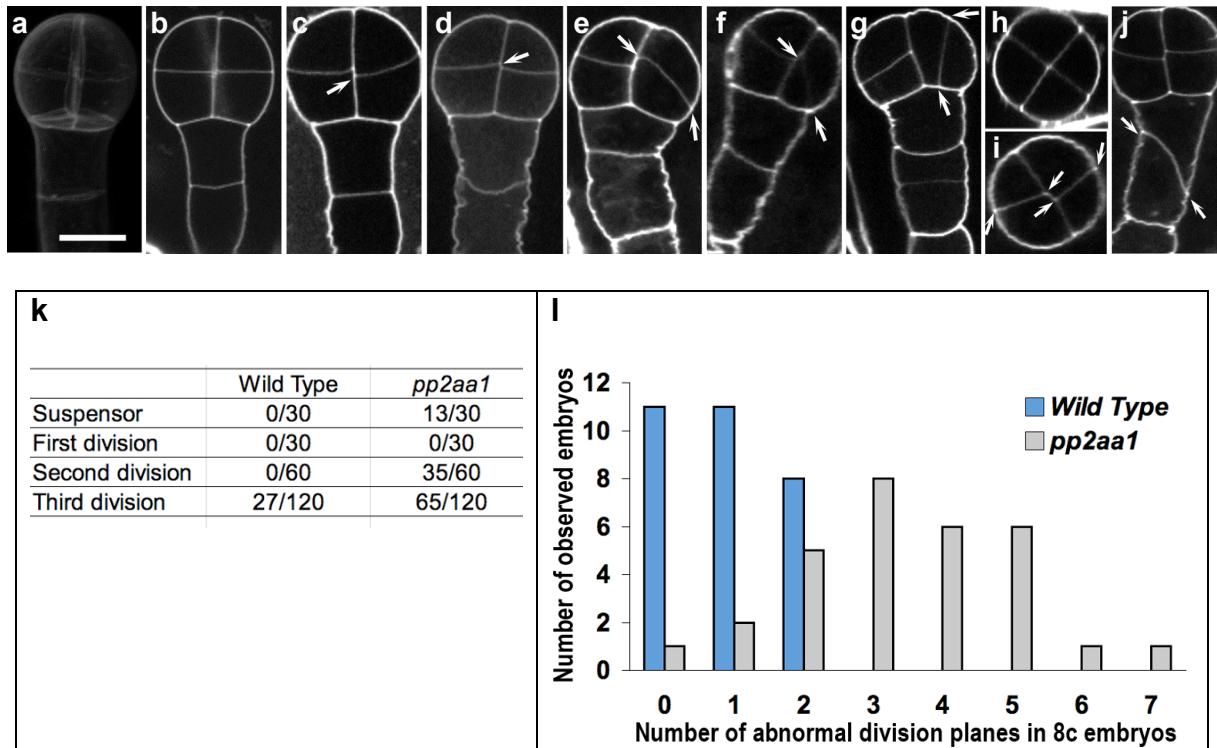
### Supplementary Figure S3



**Supplementary Figure S3: The GFP-FASS fusion complements the *fass* mutant phenotype**

Wild-type *Arabidopsis* and heterozygous *fass-5* plants were infiltrated with *Agrobacterium* carrying the Pro35S:GFP-FASS construct. At least 20 primary transformants were obtained for each genotype. To determine whether the GFP-FASS construct complements the *fass* phenotype, the progenies were analyzed for segregation of the *fass* phenotype and of the hygromycin resistance carried by the T-DNA (GFP-FASS). No plant presenting both hygromycin resistance and the *fass* mutant phenotype was recovered indicating that the GFP-FASS construct complements the *fass* phenotype. Root growth of *fass-5* complemented plants was almost fully restored (compare one-week old seedlings in the middle panel to *fass-5* (left panel) or *Col-0* (right panel)).

## Supplementary Figure S4

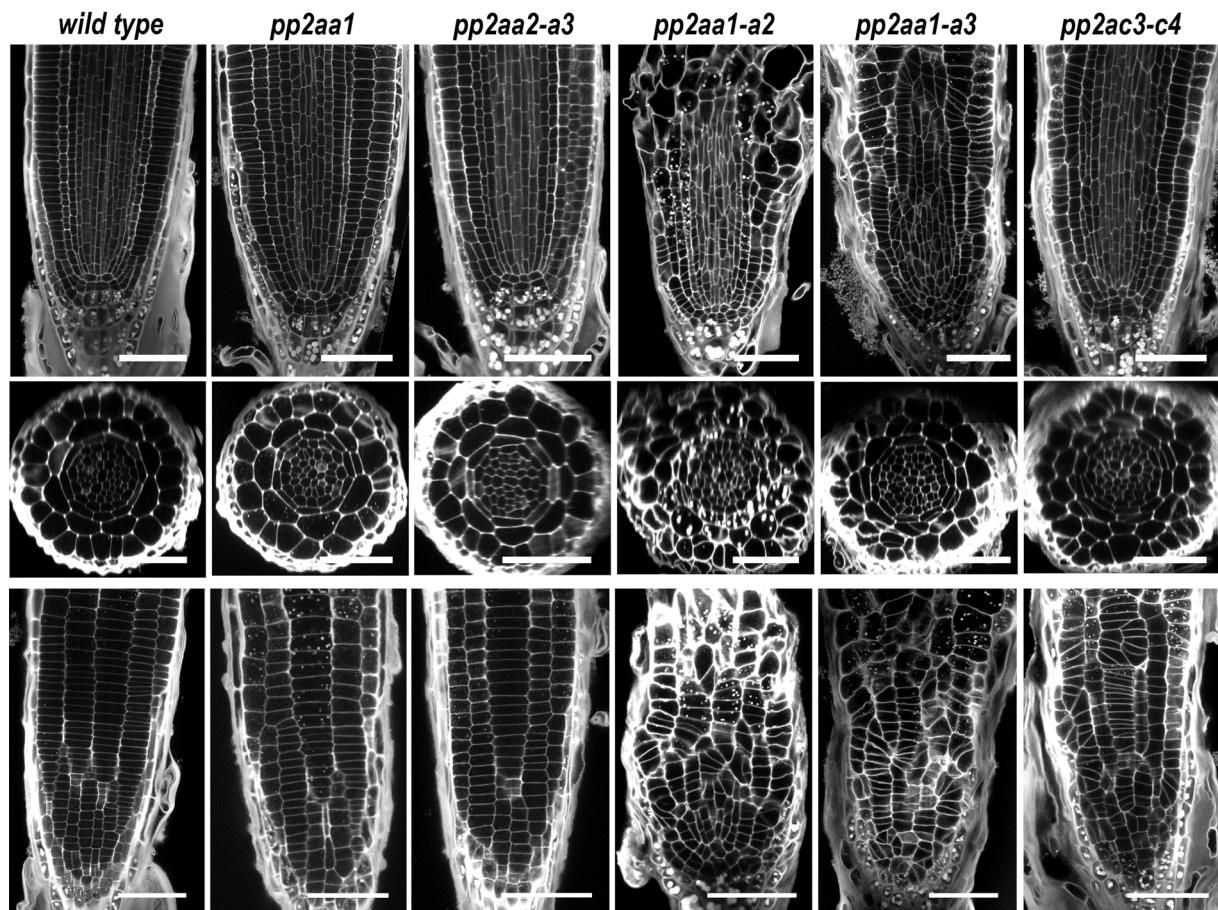


**Supplementary Figure S4 : Cell division plane defects in 8-celled *pp2aa1* mutant embryos**

(a-j) Propidium Iodide (PI) staining of 8-celled wild type (a-c and h) and *pp2aa1* (d-g, i, j) embryos. In a is shown a 3D reconstruction, in b-g and j, longitudinal views, and in h-i transverse sections of 8-celled embryos. 8-celled embryos result from three rounds of division (seven divisions in total, ie 1+2+4) in the embryo proper. The first division of the apical cell is longitudinal. The two-celled embryo then divides by a second longitudinal division perpendicular to the plane of the previous division. The four resulting cells then divide by transverse divisions, giving rise to the eight-celled embryo. PI imaging and 3D reconstruction of 8-celled embryos allowed to estimate division plane defects during these three rounds of division : Transverse sections enabled to follow the first and second round of division, whereas longitudinal sections revealed abnormal division plane during the third one. The table in k reports cell division plane defects observed in 30 embryos of *pp2aa1* compared to wild type. Abnormal division planes are visible in the second round of divisions in *pp2aa1* embryos where 35 planes were tilted (as shown in i) compared to wild type embryos (h). Defects are even more pronounced during the third round of division. At this stage, slightly tilted planes are present occasionally in wild-type embryos (27/120; as shown in c). Among 65 abnormal division planes observed in *pp2aa1* embryos, 35 appeared slightly tilted as in the wild type (c-d), while, in contrast to wild type, 15 were strongly misplaced as in e and 16, as in f or g. Aberrant division planes are also frequent in the suspensor of *pp2aa1* embryos (in 13/30 embryos as shown in j) but never observed in wild type. (l) Number of embryos displaying 0 to 7 abnormal division planes in wild type compared to *pp2aa1* embryos. Most *pp2aa1* embryos have more than 2 abnormal division planes. Wild type embryos reported here as having abnormal planes only have slightly tilted division planes.

Images in a to j are at the same scale (bar = 10 µm in a).

## Supplementary Figure S5

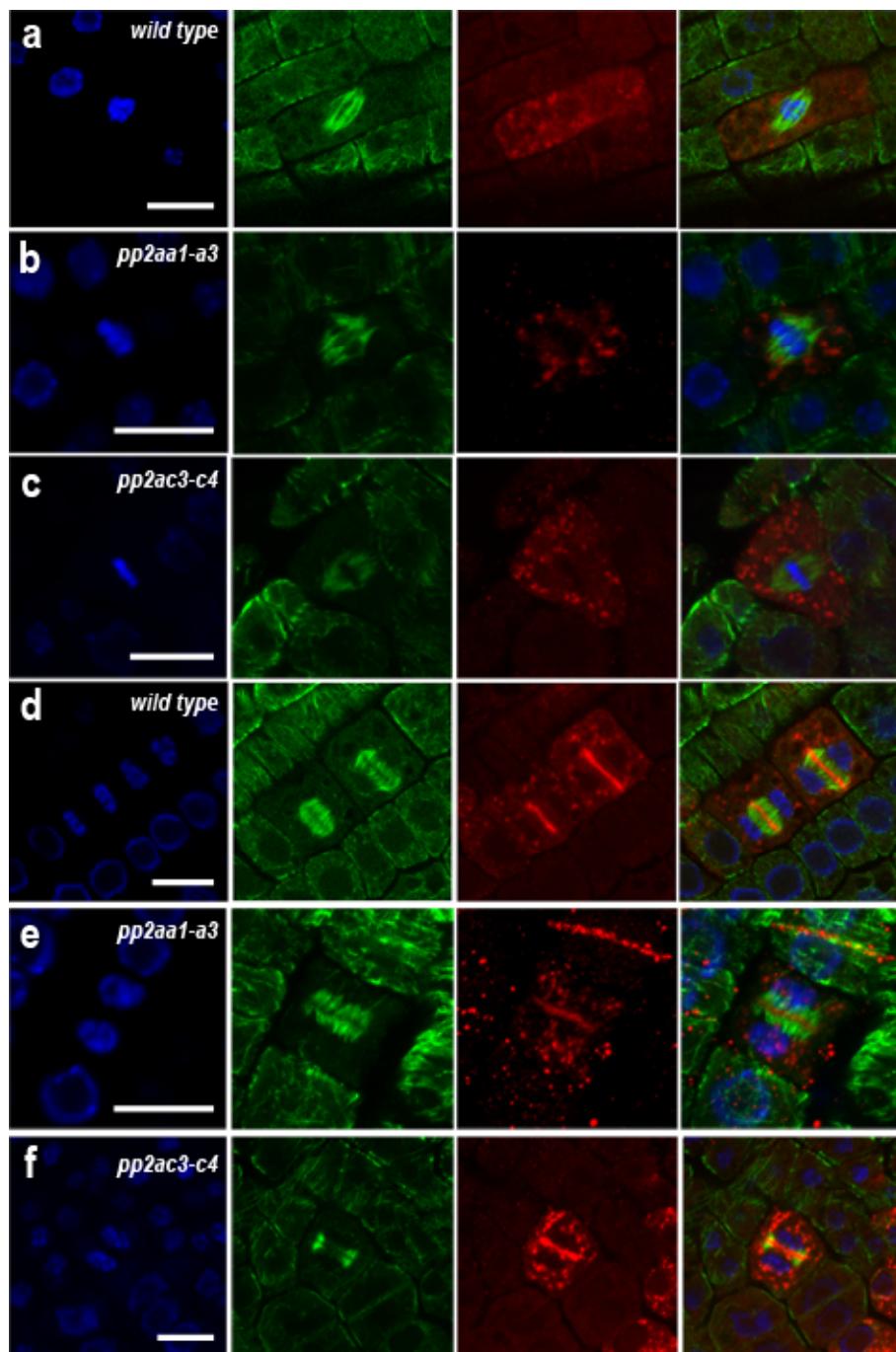


**Supplementary Figure S5 : Cell division plane defects in roots of *pp2aaa* and *pp2ac* mutants**

Longitudinal (upper panels), transverse (mid panels), and tangential views of root tips of wild-type (Col-0), *pp2aa1*, *pp2aa2-a3*, *pp2aa1-a2*, *pp2aa1-a3*, *pp2ac3-c4* three-day old seedlings stained with propidium iodide.

Bars = 50  $\mu$ m in all images.

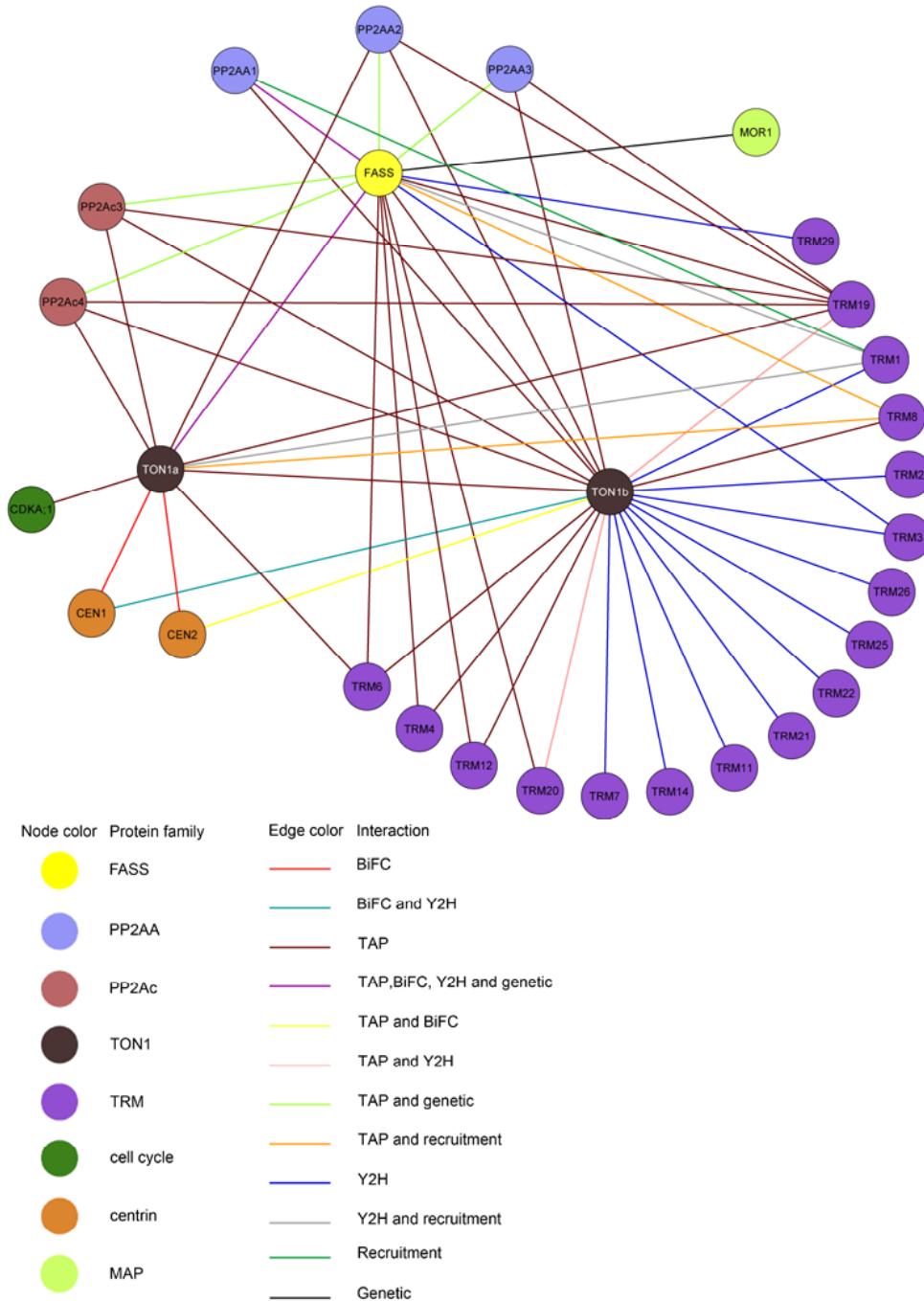
## Supplementary Figure S6



**Supplementary Figure S6 : *pp2aa* and *pp2ac* mutations don't affect spindle and phragmoplast**

(a-c) Spindles and (d-f) phragmoplasts. Whole mount immuno-localization of  $\alpha$ -tubulin, and KNOLLE in root tip cells of wild type (a and d), *pp2aa1-a3* (b and e), *pp2ac3-c4* (c and f) double mutants. Here are shown DAPI staining (in blue), anti- $\alpha$ -tubulin (in green), anti-KNOLLE (red), and overlay images. Bars = 10 $\mu$ m in a-f.

## Supplementary Figure S7



**Supplementary Figure S7: Interactome map overview of all TTP protein interactions**

This protein-protein interaction network includes all known interactions concerning the TTP proteins. The presented data is a combination of novel data described in this paper and previously published interactions<sup>10,11,12,15,25,43</sup>. The nodes represent individual proteins and the node colors are indicative of the different protein families. The lines connecting the nodes indicate known interactions and are color-coded dependent on the techniques used to identify the interactions.

**Supplementary Table S1 : MALDI MS/MS analysis of TAP eluates**

Bait	TAP Protocol	Prev			Database version	PMF data				MSMS data										Variable Modification			
		Prey Locus	Prey Name	# Found/ # exp		Protein Score	RMS error (ppm)	Sequence coverage %	Unique Peptides	Total Ion Score	Peptide Number	Start	End	Observed	Mr(Exp)	Mr(Calc)	Delta	Miss	Ions Score	Expect	Peptide		
TON1a	-dig	AT3G55005	TON1b	4/4	TAIR8	156	8.20E-12	4	42	8	106	1	237	243	863.44	862.43	0.00	0	32	2.00E-02	R.NLTSSWR.N		
TON1a	-dig	AT5G18580	FASS	4/4	TAIR8	207	6.50E-17	6	25	11	146	1	244	252	1272.56	1271.55	1271.55	0.00	0	52	1.00E-04	R.ASVEAIEEED.R.V	
TON1a	-dig	AT3G25800	PP2AA2	1/4	TAIR8	58	5.10E-02	12	32	12	/	3	126	142	1958.05	1957.04	1957.05	-0.01	0	22	7.40E-02	R.NEDSRPLLDVLEGFLK.F	
TON1a	-dig	AT2G42500/AT3 PP2AA3/PP2A4		1/4	TAIR8	68	4.60E-03	7	28	6	34	1	260	272	1589.78	1588.78	1588.77	0.01	0	15	2.80E-01	K.FFFFCDPHR.R	
TON1a	-dig	AT2G45240	MAP1A	1/4	TAIR8	47	6.50E-01	4	14	4	31	1	92	102	1310.69	1309.68	1309.68	0.00	0	50	2.50E-04	K.DTPEGLYLFR.C	
TON1a	+dig	AT3G55005	TON1b	4/4	TAIR8	441	2.60E-40	5	64	14	321	1	237	243	863.44	862.43	0.00	0	44	7.30E-04	K.EYADGTLTEIFIER.V		
TON1a	+dig	AT5G18580	FASS	4/4	TAIR8	390	3.30E-35	4	35	16	274	1	335	341	901.45	900.44	900.45	-0.01	0	41	2.40E-03	R.NLTSSWR.N	
TON1a	+dig	AT3G25800	PP2AA2	2/4	TAIR8	111	2.60E-07	9	33	13	31	1	279	287	1047.58	1046.57	1046.58	-0.01	0	40	2.50E-03	R.RASTALENQLDR.K	
TON1a	+dig	AT2G42500/AT3 PP2AA3/PP2A4		2/4	TAIR8	120	3.30E-08	4	33	8	67	2	133	142	1121.57	1120.57	1120.57	0.00	0	83	7.40E-08	R.DFSININGYELRN.N	
TON1a	+dig	AT3G06810	IBR3	1/4	TAIR8	84	1.40E-04	6	28	17	/	1	287	298	1456.72	1455.72	1455.73	-0.01	0	66	1.70E-02	K.LPFLTDIHSFLR.D	
TON1b	-dig	AT3G55000	TON1a	5/5	TAIR8	622	2.10E-58	10	51	15	487	1	241	247	863.43	862.43	0.00	0	26	8.30E-02	R.NLTSSWR.N		
TON1b	-dig	AT5G18580	FASS	5/5	TAIR8	568	5.20E-53	4	45	23	353	1	335	341	901.45	900.44	900.45	-0.01	0	35	6.00E-03	R.KLHASPSGR.L	
TON1b	-dig	AT3G25800	PP2AA2	4/5	TAIR8	225	1.00E-18	6	36	15	129	1	279	287	1047.58	1046.57	1046.58	-0.01	0	38	3.20E-03	R.KDEFNW.R.Q	Oxidation(M)
TON1b	-dig	AT2G42500/AT3 PP2AA3/PP2A4		3/5	TAIR8	218	5.20E-18	5	45	11	131	1	287	298	1456.72	1455.71	1455.73	-0.02	0	21	7.30E-02	K.FFFFCDPHR.R	Oxidation(M)
TON1b	-dig	AT1G13320	PP2AA3	1/5	TAIR8	74	1.40E-03	10	13	6	50	1	133	142	1137.58	1136.57	1136.56	0.01	0	34	2.00E-03	R.NHTFTIQFEPAPR.R	
TON1b	-dig	AT3G06810	IBR3	1/4	TAIR8	73	1.60E-03	7	16	9	39	1	18	25	1020.5	1019.49	1019.48	0.01	0	35	3.80E-03	R.GAGTYFGQDISEQFNHNTNNL.K	Oxidation(M)



TON2	+dig	AT3G05750	TRM6	2/4	TAIR8	99	4,50E-06	8	12	9	73	6	286	300	1772,85	1771,84	1771,84	0,00	0	39	2,30E-03 R.HASYDTIDLPNEELR.K
TON2	+dig	AT5G26910	TRM8	1/4	TAIR8	75	1,00E-03	4	13	9	49	7	889	904	1928,99	1927,98	1927,99	-0,01	0	58	3,00E-05 R.ELOWLSLEDDIEIG.R.E
TON2	+dig	AT2G42500/AT3 PP2A3/PP2A4		1/4	TAIR8	663	1,60E-62	3	56	13	545	1	175	183	1088,57	1087,57	1087,57	0,00	0	11	2,30E+00 R.FQTETLPPR.S
TON2	+dig	AT1G13320	PP2AA3	1/4	TAIR8	169	4,10E-13	6	20	8	130	2	119	129	1439,7	1438,7	1438,7	0,00	1	41	1,90E-03 R.RNPDFDPYFLR.S
TON2	+dig	AT1G25490	PP2AA1	1/4	TAIR8	105	1,00E-06	8	11	4	91	3	134	149	1871,89	1870,89	1870,88	-0,01	0	22	1,30E-01 K.ASTWDAYENLGYVNLR.S
TON2	+dig	AT2G45240	MAP1A	4/4	TAIR8	135	1,00E-09	10	24	7	93	1	751	758	953,48	952,47	952,48	-0,01	0	18	3,60E-01 K.GGFLFEQR.D
TON2	+dig	AT1G13320	PP2AA3	1/4	TAIR8	169	4,10E-13	6	20	8	130	2	129	141	1612,77	1611,77	1611,77	0,00	0	31	1,30E-02 R.WDAYENLGYVNLR.S
TON2	+dig	AT1G25490	PP2AA1	1/4	TAIR8	105	1,00E-06	8	11	4	91	3	141	148	951,47	950,46	950,46	0,00	0	28	5,20E-02 K.YGNANWK.I
TON2	+dig	AT2G45240	MAP1A	4/4	TAIR8	135	1,00E-09	10	24	7	93	4	287	298	1456,73	1455,72	1455,73	-0,01	0	82	1,30E-07 R.NHTFIQFEPAPR.R
TON2	+dig	AT1G13320	PP2AA3	1/4	TAIR8	169	4,10E-13	6	20	8	130	5	94	108	1641,9	1640,89	1640,9	-0,01	0	58	2,30E-05 R.GYYSVETVTLVGLK.V
TON2	+dig	AT1G25490	PP2AA1	1/4	TAIR8	105	1,00E-06	8	11	4	91	6	79	93	1865,78	1864,78	1864,78	-0,01	0	39	4,80E-04 K.CPDTNYLFMGDYVDR.G
TON2	+dig	AT2G45240	MAP1A	4/4	TAIR8	135	1,00E-09	10	24	7	93	7	244	259	1910,89	1909,88	1909,89	-0,01	0	99	1,40E-09 RAHOLVMMDGFNWAHEOK.V
TON2	+dig	AT1G13320	PP2AA3	1/4	TAIR8	169	4,10E-13	6	20	8	130	8	244	259	1926,88	1925,88	1925,88	0,00	0	(79)	1,60E-07 RAHOLVMMDGFNWAHEOK.V
TON2	+dig	AT1G25490	PP2AA1	1/4	TAIR8	105	1,00E-06	8	11	4	91	9	219	239	2314,08	2340,08	2340,08	-0,01	0	89	1,40E-08 R.GAGYTFQDISEQFNHTNNLK.L
TON2	+dig	AT2G45240	MAP1A	4/4	TAIR8	135	1,00E-09	10	24	7	93	10	54	74	2412,18	2411,17	2411,17	0,00	0	72	1,10E-06 K.SPVTICDIHGQFHDLAEFL.R.I
TIM19	-dig	AT3G55000	TON1a	2/2	TAIR9	696	8,40E-66	4	55	14	574	1	241	247	863,4374	862,4301	862,4297	0,00	0	36	8,50E-03 R.NLTSSWR.N
TIM19	-dig	AT3G55005	TON1b	2/2	TAIR9	563	1,70E-52	8	53	13	452	2	67	75	952,5194	951,5121	951,5125	-0,01	1	53	1,10E-04 R.KLHASPSGR.L
TIM19	-dig	AT5G18580	FASS	2/2	TAIR9	1060	3,30E-102	2	55	25	832	3	207	213	994,4745	993,4672	993,4668	0,00	1	57	4,90E-05 R.KDEFNW.R.Q
TIM19	-dig	AT3G25800	PP2AA2	2/2	TAIR9	558	5,30E-52	4	41	17	431	4	175	185	1144,5635	1143,5562	1143,5562	0,00	0	52	1,20E-04 R.SSASDSLPPQR.R
TIM19	-dig	AT2G42500/AT3 PP2A3/PP2A4	2/2	TAIR9	269	4,20E-23	4	30	7	176	5	226	237	1300,6829	1299,6756	1299,6782	0,00	0	87	4,20E-08 R.ASAALENLQLDR.K	
TIM19	-dig	AT1G13320	PP2AA3	1/2	TAIR9	194	1,30E-15	7	27	11	130	6	34	45	1394,6445	1393,6372	1393,6361	0,00	0	92	1,10E-08 R.ASVFAEAEED.R.V
TIM19	-dig	AT3G25800	PP2AA2	2/2	TAIR9	558	5,30E-52	4	41	17	431	7	114	125	1426,6322	1425,6249	1425,6273	0,00	0	106	3,00E-10 R.DFNSNNGFELNR.N
TIM19	-dig	AT2G42500/AT3 PP2A3/PP2A4	2/2	TAIR9	269	4,20E-23	4	30	7	176	8	46	64	2051,9885	2050,9785	2050,9742	0,00	0	93	8,90E-09 R.VIENNEGPPALLGSCNDR.A	
TIM19	-dig	AT1G13320	PP2AA3	1/2	TAIR9	194	1,30E-15	7	27	11	130	1	237	243	863,4374	862,4301	862,4297	0,00	0	36	8,50E-03 R.NLTSSWR.N
TIM19	-dig	AT3G25800	PP2AA2	2/2	TAIR9	558	5,30E-52	4	41	17	431	2	175	185	952,5194	951,5121	951,4886	0,02	0	53	1,10E-04 R.QLHASPSGR.L
TIM19	-dig	AT2G42500/AT3 PP2A3/PP2A4	2/2	TAIR9	269	4,20E-23	4	30	7	176	3	175	185	1184,5616	1183,5587	1183,5582	0,00	0	55	5,80E-05 R.SSASDSLPH.R.Q	
TIM19	-dig	AT1G13320	PP2AA3	1/2	TAIR9	194	1,30E-15	7	27	11	130	4	222	233	1330,697	1329,6897	1329,6888	0,00	0	55	7,20E-05 R.RASTALENLQLDR.K
TIM19	-dig	AT3G25800	PP2AA2	2/2	TAIR9	558	5,30E-52	4	41	17	431	5	34	45	1394,6445	1393,6372	1393,6361	0,00	0	92	1,10E-08 R.ASVFAEAEED.R.V
TIM19	-dig	AT2G42500/AT3 PP2A3/PP2A4	2/2	TAIR9	269	4,20E-23	4	30	7	176	6	114	125	1441,6615	1440,6542	1440,6633	-0,01	0	102	9,50E-10 R.DFSINNYELR.N	
TIM19	-dig	AT1G13320	PP2AA3	1/2	TAIR9	194	1,30E-15	7	27	11	130	7	187	202	1499,7043	1498,697	1498,7012	0,00	0	60	1,70E-05 R.SVSVASQASGAATSGYR.K
TIM19	-dig	AT3G25800	PP2AA2	2/2	TAIR9	558	5,30E-52	4	41	17	431	8	54	60	875,5563	874,5459	874,5528	0,00	0	53	3,00E-05 R.ILLEIFK.E
TIM19	-dig	AT2G42500/AT3 PP2A3/PP2A4	2/2	TAIR9	269	4,20E-23	4	30	7	176	9	457	464	1002,4553	1001,4468	1001,4468	0,00	0	46	2,60E-04 R.GFWAHDR.N.E	
TIM19	-dig	AT1G13320	PP2AA3	1/2	TAIR9	194	1,30E-15	7	27	11	130	10	344	443	1089,5973	1088,59	1088,59	0,00	0	104	1,20E-09 K.ITALDGLLLGCK.Q
TIM19	-dig	AT3G25800	PP2AA2	2/2	TAIR9	558	5,30E-52	4	41	17	431	11	156	164	1120,5145	1119,5072	1119,5059	0,00	0	42	8,10E-04 R.FFFSPSNFMK.F
TIM19	-dig	AT2G42500/AT3 PP2A3/PP2A4	2/2	TAIR9	269	4,20E-23	4	30	7	176	12	244	252	1272,5647	1271,5574	1271,5546	0,00	0	49	1,20E-04 K.FFFFCDPH.R.C	
TIM19	-dig	AT1G13320	PP2AA3	1/2	TAIR9	194	1,30E-15	7	27	11	130	13	369	379	1311,6573	1310,65	1310,6507	0,00	0	67	4,70E-06 K.DTPEGLYLFR.C
TIM19	-dig	AT3G25800	PP2AA2	2/2	TAIR9	558	5,30E-52	4	41	17	431	14	66	77	1327,656	1326,6487	1326,6456	0,00	0	49	2,50E-04 K.SQEAGTIPSFKY.K
TIM19	-dig	AT2G42500/AT3 PP2A3/PP2A4	2/2	TAIR9	269	4,20E-23	4	30	7	176	15	444	456	1362,7041	1361,6968	1361,6973	0,00	0	50	3,50E-04 K.QGQGTAVASMLIDVR.G	
TIM19	-dig	AT1G13320	PP2AA3	1/2	TAIR9	194	1,30E-15	7	27	11	130	16	10	29	1400,7711	1399,7638	1399,7646	0,00	1	20	1,90E-01 R.KIPPAASSMLWVR.N
TIM19	-dig	AT3G25800	PP2AA2	2/2	TAIR9	558	5,30E-52	4	41	17	431	17	387	399	1477,7791	1476,7718	1476,7725	0,00	0	115	6,60E-11 R.GFLTTADHSFLSR.D
TIM19	-dig	AT2G42500/AT3 PP2A3/PP2A4	2/2	TAIR9	269	4,20E-23	4	30	7	176	18	173	184	1514,8414	1513,8341	1513,8367	0,00	0	39	2,10E-03 R.JAILPFYLYVMR.T	
TIM19	-dig	AT1G13320	PP2AA3	1/2	TAIR9	194	1,30E-15	7	27	11	130	19	119	131	1501,7883	1500,7811	1500,7824	0,00	0	13	1,10E-01 R.DMPMPAFNQMYCR.I
TIM19	-dig	AT3G25800	PP2AA2	2/2	TAIR9	558	5,30E-52	4	41	17	431	20	321	334	1656,8107	1655,8034	1655,8042	0,00	0	94	8,70E-09 R.KEYADGLTEIFIER.V
TIM19	-dig	AT2G42500/AT3 PP2A3/PP2A4	2/2	TAIR9	269	4,20E-23	4	30	7	176	21	101	119	2220,0813	2199,074	2219,0715	0,00	0	33	7,80E-03 K.QSDOLLNADLAAMWVCL.R.E	
TIM19	-dig	AT1G13320	PP2AA3	1/2	TAIR9	194	1,30E-15	7	27	11	130	22	279	287	1047,5817	1046,5744	1046,576	0,00	0	52	1,80E-04 R.TELTPAYV.R.L
TIM19	-dig	AT3G25800	PP2AA2	2/2	TAIR9	558	5,30E-52	4	41	17	431	23	133	142	1121,5769	1120,5696	1120,5665	0,00	0	71	2,20E-06 R.LAAGEWFATAR.V
TIM19	-dig	AT2G42500/AT3 PP2A3/PP2A4	2/2	TAIR9	269	4,20E-23	4	30	7	176	24	190	200	1173,6239	1172,6166	1172,6169	0,00	0	55	9,90E-05 K.FAAFTESAHLK.T	
TIM19	-dig	AT1G13320	PP2AA3	1/2	TAIR9	194	1,30E-15	7	27	11	130	25	119	131	1501,7883	1500,7811	1500,7824	0,00	0	76	6,80E-07 R.ESDLVDHFISLVR.K
TIM19	-dig	AT3G25800	PP2AA2	2/2	TAIR9	558	5,30E-52	4	41	17	431	26	337	352	1576,8361	1575,8288	1575,8364	-0,01	0	46	2,50E-03 R.SALASVIMGMAPVLGK.D
TIM19	-dig	AT2G42500/AT3 PP2A3/PP2A4	2/2	TAIR9	269	4,20E-23	4	30	7	176	27	537	552	1767,0187	1766,0114	1766,0119	-0,01	0	42	2,90E-04 K.VLQSLIPVQDSVEK.T	
TIM19	-dig	AT1G13320	PP2AA3	1/2	TAIR9	194	1,30E-15	7	27	11	130	28	310	325	1858,0563	1857,049	1857,0546	-0,01	0	61	5,90E-06 R.ILNPEIAQHILPCVK.E
TIM19	-dig	AT3G25800	PP2AA2	2/2																	

Protein identification details were obtained with the 4800 MALDI TOF/TOF<sup>TM</sup> Proteomics analyzer (AB SCIEX) and the GPS explorer v3.6 (AB SCIEX) software package combined with search engine Mascot version 2.1 or 2.2 (Matrix Science)

Column headers for Protein and Peptide data are as follows :

**Protein score:** The score calculated by the Mascot search engine for each protein. This score is based on the probability that peptide mass matches are non-random events. If the Protein Score is equal to or greater than the Mascot® Significance Level calculated for the database search, the protein match is considered to be statistically non-random at the 95% confidence interval. Protein score = -10\*Log(P), where P is the probability that the observed match is a random event. **Expect:** Protein score expectation value. **RMS error (ppm):** RMS error of the set of matched mass values, in ppm. **Sequence coverage %:** Percentage of protein sequence covered by assigned peptide matches. **Unique peptides:** The number of peptides with unique sequences matching the selected protein. **Total Ion Score:** A score calculated by weighting Ion Scores for all individual peptides matched to a given protein. **Peptide Number:** Peptide index number within the list of peptides associated with a given protein. **Start:** The starting position of the peptide in the protein. **End:** The ending position of the peptide in the protein. **Observed:** The observed monoisotopic mass of the peptide in the spectrum (m/z). **Mr (Exp):** The experimental mass of the peptide calculated from the observed m/z value. **Mr (Calc):** The theoretical mass of the peptide based on its sequence. **Delta (Da):** The difference between the theoretical (Mr (Calc)) and experimental (Mr (Exp)) masses, in daltons. **Miss:** Number of missed Trypsin cleavage sites. **Ions score:** The Ions Score is calculated by the Mascot search engine for each peptide matched from MS/MS peak lists. This score is based on the probability that ion fragmentation matches are non-random events. If the Ion Score is equal to or greater than the Mascot® Significance Level calculated for the database search, the peptide match is considered to be statistically non-random at the 95% confidence interval. Ions score = -10\*Log(P), where P is the probability that the observed match is a random event. **Best Ions score:** The highest individual Ion Score for a given protein identification. **Expect:** Ions score expectation value. **Peptide:** The amino acid sequence of the selected peptide. **Variable Modification:** Variable modification type on the peptide.

**Supplementary Table S2: VELOS orbitrap analysis of TAP TON1b eluates**

Accession	Description	Score	Coverage	# Unique Peptides	# Peptides	# AAs					
<b>AT3G55005</b>	<b>TON1b</b>	<b>237,09</b>	<b>64,20</b>	<b>17</b>	<b>18</b>	<b>257</b>					
	Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	ΔM [ppm]	m/z [Da]	RT [min]	# Missed Cleavages
	High	NEDSRPLLDVLEGLFK		1	4,69	3	1958,06035	0,47	653,35830	28,73	0
	High	NEDSRPLLDVLEGLFK	E13(Methyl)	1	4,48	3	1972,07370	-0,69	658,02942	29,64	0
	High	NEDSRPLLDVLEGLFKENNmTQVMGGSSR	M21(Oxidation)	1	4,48	4	3398,66884	-0,95	850,42267	29,10	1
	High	ASVFEAIEEDRVIVIENNEGPPALLGScNDR	C28(Carbamidomethyl)	1	4,43	4	3443,63776	-0,23	861,66490	26,02	1
	High	SEIRDFSINNGYelNLR	E13(Methyl)	1	4,13	3	1940,94687	0,37	647,65381	21,77	1
	High	RSVSASQASGAATSGYR		1	4,08	2	1655,80777	-1,14	828,40752	17,14	1
	High	FENnTQVmMGSSR	M4(Oxidation); M8(Oxidation)	1	3,79	2	1475,62496	-0,32	738,31612	17,30	0
	High	VlennNeGLPPALLGScNDR	E3(Methyl); E6(Methyl); C16(Carbamidomethyl)	1	3,74	2	2096,04546	0,61	1048,52637	23,56	0
	High	LLSALIcEYLDWAQLNHTLK	C7(Carbamidomethyl)	1	3,68	2	2401,25728	-0,51	1201,13228	27,13	0
	High	YDTEEdmPEEVmR	M6(Oxidation); M11(Oxidation)	1	3,65	2	1546,60247	-0,78	773,80487	18,49	0
	High	ASVFEAIEEDRVIVIENNEGPPALLGScNDR	E18(Methyl); C28(Carbamidomethyl)	1	3,60	3	3457,65342	-0,22	1153,22266	27,17	1
	High	ASTALENLQLDRK		1	3,58	2	1458,78997	-0,81	729,89862	20,22	1
	High	FeNMTQVmMGSSR	E2(Methyl)	1	3,55	2	1457,65068	-0,38	729,32898	21,93	0
	High	SEIRDFSINNGYelNLR		1	3,55	3	1926,93133	0,43	642,98196	21,38	1
	High	VIENNEGPPALLGScNDR	C16(Carbamidomethyl)	1	3,54	3	2068,01265	-0,11	690,00907	22,47	0
	High	ReSETESSLSDTR	E2(Methyl)	1	3,53	2	1623,78048	-1,00	812,39388	19,08	1
	High	NedSRPLLDVLEGLFK	E2(Methyl); D3(Methyl)	1	3,48	2	1986,08820	-1,27	993,54774	29,20	0
	High	ESETESSLSDTR		1	3,46	2	1453,66570	0,25	727,33649	20,03	0
	High	ASTALENLQLDR		1	3,39	2	1330,69585	-0,25	665,85156	21,68	0
	High	SVSASQASGAATSGYR		1	3,39	2	1499,70744	-0,73	750,35736	18,20	0
	High	eSETESSLSDTR	E1(Methyl)	1	3,38	2	1467,68340	1,65	734,34534	20,09	0
	High	FENnTQVmMGSSR	M4(Oxidation)	1	3,37	2	1459,62924	-0,87	730,31826	18,99	0
	High	YDTEEdmPEEVmR	M6(Oxidation); E9(Methyl)	1	3,23	2	1544,62241	-1,30	772,81485	20,28	0
	High	ASTALENLQLDR	E6(Methyl)	1	3,20	2	1344,71044	-1,03	672,85886	21,40	0
	High	DFSINNGYelNLR		1	3,20	2	1441,67035	-0,23	721,33881	22,28	0
	High	YDTEEdmPEEVmR		1	3,14	2	1514,61221	-1,09	757,80974	21,49	0
	High	SSASDSLPHQR		1	3,14	2	1184,56520	-0,24	592,78624	16,35	0
	High	eSeTESSLSDTR	E1(Methyl); E3(Methyl)	1	3,13	2	1481,69475	-1,27	741,35101	21,14	0
	High	YDTEEdmPeeVMR	M6(Oxidation); E8(Methyl); E9(Methyl)	1	3,11	2	1558,63833	-1,12	779,82280	20,89	0
	High	YDTEEdmPEEVmR	M6(Oxidation)	1	2,96	2	1530,60719	-1,03	765,80723	19,74	0
	High	ASVFEAIEEEdR	D11(Methyl)	1	2,96	2	1408,65850	-0,45	704,83289	24,03	0
	High	ASVFEAIEEEdR		1	2,92	3	1394,64457	0,77	465,55304	23,22	0
	High	ASVFEAIeDRVIVIENNEGPPALLGScNDR	E9(Methyl); E10(Methyl); C28(Carbamidomethyl)	1	2,91	4	3471,66665	-0,92	868,67212	26,83	1
	High	RSSASDSLPHQR		1	2,88	2	1340,66502	-1,18	670,83615	15,96	1
	High	YDTEEdmPeeVmR	M6(Oxidation); E8(Methyl); E9(Methyl); M11(Oxidation)	1	2,87	2	1574,63445	-0,34	787,82086	19,71	0
	High	YDTEEdmPEEVmR	E4(Methyl); M6(Oxidation); M11(Oxidation)	1	2,85	2	1560,61769	-1,05	780,81248	18,77	0
	High	RESeTeSSLSDTR	E4(Methyl); E6(Methyl)	1	2,83	3	1637,79674	-0,62	546,60376	19,58	1
	High	FeNnTQVmMGSSR	E2(Methyl); M4(Oxidation)	1	2,82	2	1473,64531	-0,57	737,32629	19,37	0
	High	VIENNEGPPALLGScNDR	E6(Methyl); C16(Carbamidomethyl)	1	2,81	3	2082,02652	-0,97	694,68036	22,75	0
	High	DFSINNGYelNLR	E9(Methyl)	1	2,80	2	1455,68541	-0,63	728,34634	22,51	0
	High	RESETESSLSDTR		1	2,80	2	1609,76513	-0,82	805,38620	19,23	1
	High	YDTEEdmPEEVmR	E4(Methyl)	1	2,52	2	1528,62847	-0,68	764,81787	22,07	0
	High	ASVFEAIeEEEdR	E8(Methyl); E9(Methyl); E10(Methyl)	1	2,39	2	1436,68880	-1,14	718,84804	25,31	0
<b>AT3G55000</b>	<b>TON1a</b>	<b>129,58</b>	<b>60,00</b>	<b>13</b>	<b>14</b>	<b>260</b>					
	Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	ΔM [ppm]	m/z [Da]	RT [min]	# Missed Cleavages
	High	ASVFEAIEEDRVIVIENNEGPPALLGScNDR	C28(Carbamidomethyl)	1	5,27	3	3427,60361	-1,06	1143,20605	25,47	1
	High	LLSALIcEYLDWAQLNHTLIVYQPESNLPK	C7(Carbamidomethyl)	1	4,89	3	3541,84348	0,96	1181,28601	28,02	0
	High	NGDSGPLLLDVLEGLFK	E13(Methyl)	1	4,82	2	1800,97258	-0,95	900,98993	31,08	0
	High	NGDSGPLLLDVLEGLFK		1	4,53	3	1786,95831	-0,18	596,32429	30,35	0
	High	SELRFNSNNGFELNR		1	4,47	3	1911,89481	0,18	637,96979	21,01	1
	High	VIENNEGPPALLGScNDR	E3(Methyl); C16(Carbamidomethyl)	1	4,09	2	2065,99785	0,30	1033,50256	21,50	0
	High	DSETESSSSLESR		1	3,99	2	1413,59636	-0,91	707,30182	17,68	0

High	ASVFEAIEEEDRViNNEGPPALLGScNDR	E15(Methyl); C28(Carbamidomethyl)	1	3,95	3	3441,62063	-0,66	1147,87839	25,03	1
High	RDSETESSSSLeSR		1	3,33	3	1569,69815	-0,39	523,90424	16,67	1
High	VIENNEGPPALLGScNDR	C16(Carbamidomethyl)	1	3,32	2	2051,97952	-1,01	1026,49340	21,15	0
High	SSASDSLPPQR		1	3,31	2	1144,55866	-0,60	572,78297	17,35	0
High	RDSETESSSSLeSR	E12(Methyl)	1	3,31	3	1583,71410	-0,20	528,57622	16,82	1
High	FESmTQGmGSSSR	M4(Oxidation); M8(Oxidation)	1	3,30	2	1436,57681	-0,92	718,79204	16,35	0
High	DFNSNNGFELNR		1	3,28	2	1426,63337	-0,89	713,82032	21,74	0
High	RSSASDSLPPQR		1	3,24	2	1300,65837	-1,61	650,83282	16,58	1
High	dSETESSSSLESR	D1(Methyl)	1	3,09	2	1427,61283	-0,32	714,31005	17,43	0
High	RPVSASQASDR		1	2,98	2	1173,59650	-0,53	587,30189	15,45	0
High	ASVFEAIEEEDR	D11(Methyl)	1	2,96	2	1408,65850	-0,45	704,83289	24,03	0
High	ASVFEAIEEEDR		1	2,92	3	1394,64457	0,77	465,55304	23,22	0
High	FESMTQGmGSSSR	M8(Oxidation)	1	2,87	2	1420,58199	-0,87	710,79463	17,67	0
High	QGNQDTHeeVTR	E9(Methyl)	1	2,53	2	1427,64940	-1,13	714,32834	16,28	0
High	ASVFEAIEeeDR	E8(Methyl); E9(Methyl); E10(Methyl)	1	2,39	2	1436,68880	-1,14	718,84804	25,31	0
<b>AT5G18580</b>	<b>FASS</b>	<b>127,05</b>	<b>67,50</b>	<b>22</b>	<b>22</b>	<b>480</b>				

Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	ΔM [ppm]	m/z [Da]	RT [min]	# Missed Cleavages
High	EmDFDSFLDFVLALENK	M2(Oxidation)	1	4,90	2	2048,94927	-1,45	1024,97827	30,00	0
High	QELKEYADGTLTEIFIER		1	4,89	3	2155,09128	-0,26	719,03528	24,03	1
High	MNYEDFchIASVcTEQIGPK	C7(Carbamidomethyl); C13(Carbamidomethyl)	1	4,82	3	2399,04288	-1,53	800,35248	22,80	0
High	YIGSGAGLGLSEALMELETK		1	4,44	2	1925,95305	0,25	963,48016	24,17	0
High	KKPEEGSISQR		1	4,27	3	1258,67521	0,12	420,22992	15,40	1
High	mNYEDFchIASVcTEQIGPK	(Oxidation); C7(Carbamidomethyl); C13(Carbamidomethyl)	1	3,99	3	2415,03824	-1,34	805,68427	21,90	0
High	IcDmFLALDKDmSGSLcK	IcDmFLALDKDmSGSLcK	1	3,93	2	2135,94780	-0,15	1068,47754	21,84	1
High	QSDILLNADDLAAWMWvLR	C17(Carbamidomethyl)	1	3,93	3	2204,08261	-0,61	735,36572	28,42	0
High	YIGSGAGLGLSEALmELETK	M14(Oxidation); E15(Methyl)	1	3,86	2	1955,96282	-0,16	978,48505	24,45	0
High	YIGSGAGLGLSEALmELETK	M14(Oxidation)	1	3,78	2	1941,94621	-0,65	971,47675	23,73	0
High	IDmSELDEDSDGFLHSDEMESYIGGLIPNLAQLR	M3(Oxidation)	1	3,70	3	3825,73899	-2,14	1275,91785	27,76	0
High	IcDMFLALDKDmSGSLcK	Carbamidomethyl); M12(Oxidation); C17(Carbamidomethyl)	1	3,64	2	2119,94902	-1,97	1060,47815	23,11	1
High	KIPPASSmLWVR	M8(Oxidation)	1	3,51	3	1400,77150	-0,32	467,59535	20,34	1
High	ILLSnLQELmELHQSEEETDTEQAEWNWFLTSaQR	C6(Carbamidomethyl); M11(Oxidation)	1	3,44	4	4524,07680	-1,13	1131,77466	26,34	0
High	WIEGGNYELcIEDVR	C10(Carbamidomethyl)	1	3,43	2	1852,85307	-0,23	926,93018	23,67	0
High	QGGTVASmLIDVR	M8(Oxidation)	1	3,37	2	1362,70378	-0,64	681,85553	21,77	0
High	IDmSELDEDSDGFLHSDEmESYIGGLIPNLAQLR	M3(Oxidation); M19(Oxidation)	1	3,35	3	3841,74643	1,13	1281,25366	26,33	0
High	IAILPFYLYVmR	M11(Oxidation)	1	3,34	2	1514,84295	-0,71	757,92511	26,65	0
High	DmPPAFNQMYcR	M2(Oxidation); C11(Carbamidomethyl)	1	3,21	2	1545,62773	-0,43	773,31750	21,30	0
High	IAILPFYLYVmR		1	3,16	2	1498,84734	-1,18	749,92731	27,98	0
High	DmPPAFNQMycR	M2(Oxidation); M9(Oxidation); C11(Carbamidomethyl)	1	3,13	2	1561,62419	0,57	781,31573	19,81	0
High	GFWAHHDNRENLQEEEEEPEEESQ		1	3,13	3	2912,25840	-0,78	971,42432	22,16	1
High	mNyEdFchIASVcTEQIGPK	tion); E4(Methyl); C7(Carbamidomethyl); C13(Carbamidomethyl)	1	3,03	3	2429,05442	-1,11	810,35632	22,33	0
High	SQEAGTIPSFYK		1	2,99	2	1327,65325	0,26	664,33026	21,27	0
High	IcDMFLALDK	C2(Carbamidomethyl)	1	2,92	2	1225,59514	-0,39	613,30121	24,30	0
High	YIGSGAGLGLSEALmELETK	M14(Oxidation); E15(Methyl); E17(Methyl)	1	2,89	2	1969,97820	-0,30	985,49274	25,07	0
High	EYADGTLTEIFIER		1	2,89	3	1656,81076	-0,51	552,94177	24,89	0
High	ITLGDLGGK	C9(Carbamidomethyl)	1	2,69	2	1089,59714	-0,16	545,30221	24,73	0
High	ILLEIFK		1	2,63	2	875,55997	-0,17	438,28362	24,56	0
High	QGGTVASmLIDVR		1	2,61	2	1346,70891	-0,62	673,85809	23,55	0
High	IcDmFLALDK	C2(Carbamidomethyl); M4(Oxidation)	1	2,56	2	1241,58916	-1,10	621,29822	22,61	0
High	ENcVIDDATGAcK	C3(Carbamidomethyl); E12(Methyl)	1	2,46	2	1435,63603	-0,69	718,32166	20,31	0
High	KPEEGSISQR		1	2,42	2	1130,57930	-0,69	565,79329	15,93	0

Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	ΔM [ppm]	m/z [Da]	RT [min]	# Missed Cleavages
High	LAIIIEYIPLLASQLGVGFFDK		1	5,62	3	2422,32859	0,68	808,11438	31,48	0
High	YmVANQLYELcAVGPEPTR	M2(Oxidation); C11(Carbamidomethyl)	1	4,92	2	2356,09331	-0,67	1178,55029	23,66	0
High	TDVMSmFEDLTQDDQDSVR	M6(Oxidation)	1	4,57	2	2247,93486	-1,46	1124,47107	24,43	0
High	FFANQALQS1DNVMmSS	M15(Oxidation)	1	4,40	2	1918,87187	2,30	959,93958	27,11	0
High	FFANQALQS1DNVmMSS	M14(Oxidation); M15(Oxidation)	1	4,10	2	1934,86165	-0,37	967,93446	24,43	0
High	SLYTQLcQDDmPmVR	C7(Carbamidomethyl); M11(Oxidation); M13(Oxidation)	1	4,07	2	1888,82247	-0,74	944,91487	21,55	0
High	SALASVImGmAPVLGK	M8(Oxidation); M10(Oxidation)	1	4,00	3	1576,84348	-0,22	526,28601	22,87	0

High	LAIIEYIPLLASQLGVGFFdK	D21(Methyl)	1	3,98	3	2436,33982	-1,14	812,78479	31,43	0
High	TIRPGLVELSEPPDVDR		1	3,82	2	2010,05022	-0,05	1005,52875	21,89	0
High	ILNPEIAIQHILPcVK	C14(Carbamidomethyl)	1	3,78	2	1858,06035	-0,88	929,53381	24,89	0
High	VLSQSLIPDVQSVWEK		1	3,76	3	1767,02597	-0,19	589,68018	24,53	0
High	SALASVImGMAPVLGK	M8(Oxidation)	1	3,65	2	1560,84636	-1,63	780,92682	25,63	0
High	TIRPGLVELSeDPDVDR	E11(Methyl)	1	3,44	3	2024,06595	-0,01	675,36017	22,51	0
High	LLEPQDcVQHILPVNVFSQDK	C7(Carbamidomethyl)	1	3,32	3	2592,34824	-0,35	864,78760	25,31	0
High	AVSLLAPVmGSEITcSK	M9(Oxidation); C15(Carbamidomethyl)	1	3,31	2	1778,90068	-1,18	889,95398	22,42	0
High	ESDLVDHFISLKV		1	3,29	2	1501,78813	-1,08	751,39771	25,17	0
High	LLRDNEAEVR		1	2,93	2	1214,64739	-1,20	607,82733	17,65	1
High	VSACGVFHIAYPSPAdmLK	C4(Carbamidomethyl); M17(Oxidation)	1	2,62	2	2079,00127	-1,26	1040,00427	22,52	0
High	FAATVeSAHLK	E6(Methyl)	1	2,61	2	1187,64128	-0,56	594,32428	19,47	0
High	ELSSDSSOHV		1	2,59	2	1244,58501	-1,29	622,79614	16,21	0

Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	$\Delta M$ [ppm]	m/z [Da]	RT [min]	# Missed Cleavages	AT3G06810.1	IBR3	86,92	36,65	19	19	824
High	VSQFGHGQSNTPLIEVGSGSSLKR		1	4,98	3	2632,35056	1,01	878,12170	21,69	1							
High	SGFAmTEPVASSADATNIcSIR	M5(Oxidation); C20(Carbamidomethyl)	1	4,74	2	2487,11235	-0,16	1244,05981	21,82	0							
High	ALASLHSADVDAIGLEK		1	4,56	2	1709,10544	-0,87	855,45636	21,30	0							
High	IADGPDEVHILGTIGK		1	4,36	2	1521,78942	-0,93	761,39835	20,50	0							
High	NTGVQANELVESALGYIAR		1	4,26	2	2005,03667	0,84	1003,02197	26,06	0							
High	NIPAEDESTGATSGLVLHGDFR		1	4,14	2	2043,97295	-0,08	1022,49011	20,79	0							
High	FAADNVSGFPNTNPQSFK		1	4,09	2	1826,86897	-1,04	913,93812	22,32	0							
High	YGNKEQISEWLIPPLEGR		1	3,92	3	2145,13547	0,70	715,71667	26,20	1							
High	EGLWNLFVPVDSAAR		1	3,90	2	1673,86363	-0,62	837,43545	27,07	0							
High	ELAATENKHNLGK		1	3,76	2	1511,77995	-0,90	756,39362	16,56	1							
High	SFDQLFGEGLTNLEGYLcEImGR	C19(Carbamidomethyl); M22(Oxidation)	1	3,76	3	2828,28916	-0,55	943,43457	28,54	0							
High	FIAQHGSFVSDLAK		1	3,76	2	1519,78874	-1,10	760,39801	21,01	0							
High	VSQFGHGQSNTPLIEVGSGSSLK		1	3,39	3	2476,24308	-1,50	826,08588	22,61	0							
High	SVWARQVFNgAPDTGNnEVILR	C10(Carbamidomethyl); M18(Oxidation)	1	3,19	3	2577,21970	-1,12	859,74475	24,01	0							
High	FmETHIYpMNEFSK	M2(Oxidation); M9(Oxidation)	1	2,99	2	1934,82915	-0,44	967,91821	20,30	0							
High	LLVLEAADHLDK		1	2,98	2	1336,74687	-0,22	668,87708	22,20	0							
High	WLMGNASAGER	M3(Oxidation)	1	2,90	2	1207,55156	-0,78	604,27942	19,23	0							
High	QGDSYVINGTK		1	2,78	2	1181,57939	-0,29	591,29333	18,90	0							
High	mFELVDWLR	M1(Oxidation)	1	2,67	2	1224,60784	-0,32	612,80756	26,26	0							
High	FMETHIYpMNEFSK	M9(Oxidation)	1	2,63	2	1918,83464	-0,23	959,92096	21,06	0							

Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	$\Delta M$ [ppm]	m/z [Da]	RT [min]	# Missed Cleavages	AT3G53540	TRM19	48,20	19,16	12	12	924
High	SHSSYNSSPEVTPSLSK		1	4,62	2	2006,96501	-0,79	1003,98615	19,96	0							
High	EFNDALEALDSNKLILLKK		1	4,55	2	2048,05571	0,48	1024,53149	24,33	1							
High	ELQWLISLEDDIEIIGR		1	4,09	2	1928,99358	-1,51	965,00043	27,33	0							
High	VNALNLQAEAMAFIR		1	3,84	2	1577,80950	-0,65	789,40839	23,49	0							
High	HASYDTIDLPNEELR		1	3,82	2	1772,84490	-0,08	886,92609	21,28	0							
High	TAmSSEALDLSTVTSVTDPSISR	M3(Oxidation)	1	3,59	2	2412,14263	-0,88	1206,57495	23,27	0							
High	LmGLDVLPSQSSHK	M2(Oxidation)	1	3,57	2	1614,81304	-1,62	807,91016	20,74	0							
High	LTHKFEHEIEISR		1	3,28	4	1638,85903	-0,53	410,47021	18,39	1							
High	KLLFDQISR		1	2,92	3	1119,65165	-0,42	373,88873	21,21	1							
High	TFASPSSSSDEFR		1	2,87	2	1417,62322	0,10	709,31525	20,06	0							
High	FLQHPDSLFTK		1	2,74	2	1332,69646	1,31	666,85187	21,15	0							
High	FKDVFEVLDAK		1	2,62	2	1310,69878	-0,28	655,85303	23,37	1							

Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	$\Delta M$ [ppm]	m/z [Da]	RT [min]	# Missed Cleavages	AT1G13320	PP2A3	45,71	26,92	3	10	587
High	YmVANQLYElcEAVGPEPTR	M2(Oxidation); C11(Carbamidomethyl)	1	4,92	2	2356,09331	-0,67	1178,55029	23,66	0							
High	SALASVImGmAPVLGK	M8(Oxidation); M10(Oxidation)	1	4,00	3	1576,84348	-0,22	526,28601	22,87	0							
High	LAIIYEIPLLASQLGVGFDEK		2	3,98	3	2436,33982	-1,14	812,78479	31,43	0							
High	SIYGLcQDDmPmVR	C7(Carbamidomethyl); M11(Oxidation); M13(Oxidation)	1	3,84	2	1844,79436	-1,79	922,90082	21,03	0							
High	ILNPEIAIQHILPcVK	C14(Carbamidomethyl)	1	3,78	2	1858,06035	-0,88	929,53381	24,89	0							
High	SALASVImGMAPVLGK	M8(Oxidation)	1	3,65	2	1560,84636	-1,63	780,92682	25,63	0							
High	YFANQALQSIDNVmmSS	M14(Oxidation); M15(Oxidation)	1	3,39	2	1950,85198	-2,71	975,92963	23,81	0							
High	AVSLLAPVmGSEITcSK	M9(Oxidation); C15(Carbamidomethyl)	1	3,31	2	1778,90068	-1,18	889,95398	22,42	0							

	High	ESDLVEHFTPLAK		1	2,65	2	1485,75713	-0,89	743,38220	21,97	0
	High	ELSSDSQHVR		1	2,59	2	1244,58501	-1,29	622,79614	16,21	0
	High	FAATIESAHLK		2	2,55	2	1187,64128	-0,56	594,32428	19,47	0
<b>AT3G58500</b>	<b>PP2A-4</b>	<b>41,53</b>	<b>45,37</b>	<b>10</b>	<b>10</b>	<b>313</b>					
Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	ΔM [ppm]	m/z [Da]	RT [min]	# Missed Cleavages	
High	GAGYTFGQDISEQFNHTNNLK		1	4,78	2	2341,08257	-0,80	1171,04492	22,30	0	
High	cPDTNYLFMDGYVDR	C1(Carbamidomethyl)	1	4,51	2	1865,78093	-1,30	933,39410	24,23	0	
High	GYYSVETVLLVGLK		1	4,37	2	1641,90911	-0,45	821,45819	25,96	0	
High	SPVTIcGDIHQFHDLAEFLR	C6(Carbamidomethyl)	1	3,71	3	2412,17466	-0,80	804,72974	24,54	0	
High	cPDTNYLFmGDYYVDR	C1(Carbamidomethyl); M9(Oxidation)	1	3,55	2	1881,77556	-1,45	941,39142	22,89	0	
High	AHQVLmDGFnWAHEQK	M6(Oxidation)	1	3,27	3	1926,89096	-0,34	642,96851	20,35	0	
High	cGNnASLLEVDDCR	Carbamidomethyl); M4(Oxidation); C13(Carbamidomethyl)	1	3,22	2	1655,68266	0,22	828,34497	20,96	0	
High	AKEILmDESNVQPVK	M6(Oxidation)	1	3,18	2	1716,88078	-1,73	858,94403	18,90	1	
High	NHTFIQFEPAPR		1	2,78	2	1456,73174	-1,03	728,86951	20,98	0	
High	EILmDESNVQPVK	M4(Oxidation)	1	2,56	2	1517,74986	-1,19	759,37857	19,86	0	
High	VVTIIFASPNCYR	C11(Carbamidomethyl)	1	2,47	2	1589,77725	-0,54	795,39226	23,10	0	
<b>AT5G26910</b>	<b>TRM8</b>	<b>38,62</b>	<b>21,83</b>	<b>11</b>	<b>11</b>	<b>852</b>					
Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	ΔM [ppm]	m/z [Da]	RT [min]	# Missed Cleavages	
High	VLESESVSdCTSFYDK	C11(Carbamidomethyl)	1	4,33	2	1952,84294	-0,05	976,92511	21,00	0	
High	SDScAASSVTSDDGQGTR	C5(Carbamidomethyl); C6(Carbamidomethyl)	1	4,28	2	1976,75078	-2,50	988,87903	17,47	0	
High	EYALGmATDVLPAISLFDEmEGR	M6(Oxidation); M19(Oxidation)	1	4,24	2	2447,10674	-1,60	1224,05701	25,32	0	
High	GLSSDSLSSTQGIGQDTDSAVSFNIGGDSLNLALLEQK		1	4,21	3	3711,77817	-1,39	1237,93091	25,68	0	
High	LmGLESPLPVNVQePR	M2(Oxidation); E14(Methyl)	1	3,76	2	1808,95256	-2,78	904,97992	23,58	0	
High	WDAYENLGYYNLR		1	3,69	2	1612,77402	-0,91	806,89065	23,68	0	
High	EmDVISFTFSSPIK	M2(Oxidation)	1	3,30	2	1616,78630	-0,87	808,89679	24,57	0	
High	EGWSALVEPHLLSAF		1	3,19	2	1667,89958	-0,48	834,45343	26,25	0	
High	FQSETFPKR		1	2,60	2	1108,54094	-1,16	554,77411	21,18	0	
High	NPIYVmEAASR	M6(Oxidation)	1	2,56	2	1266,61394	-0,65	633,81061	19,46	0	
High	ITTSLLTPSTS	K	1	2,44	2	1236,66716	-0,85	618,83722	19,02	0	
<b>AT4G28760</b>	<b>TRM20</b>	<b>35,29</b>	<b>13,53</b>	<b>9</b>	<b>9</b>	<b>924</b>					
Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	ΔM [ppm]	m/z [Da]	RT [min]	# Missed Cleavages	
High	LLQELVVEAVIDLTR		1	4,99	2	1740,97331	-0,56	870,99030	29,93	0	
High	DSGEDmDANSLAESLVK	M6(Oxidation)	1	4,96	2	1867,82109	-0,85	934,41418	21,79	0	
High	DASQCCSmSQLAAPSPVLTGK	C5(Carbamidomethyl); M8(Oxidation)	1	3,87	2	2252,05107	-1,05	1126,52917	21,46	0	
High	INSIVSETTTTR		1	3,85	2	1321,69499	-0,63	661,35114	19,64	0	
High	TSSTLGEmlALTETK	M8(Oxidation)	1	3,69	2	1597,79778	-0,77	799,40253	21,60	0	
High	AVSSQQSSPR		1	3,06	2	1005,49584	-0,16	503,25156	15,45	0	
High	VSNLFFFK		1	2,70	2	1001,54588	0,36	501,27658	24,50	0	
High	LASSSQETGWGNR		1	2,61	2	1392,64812	-1,56	696,82770	18,83	0	
High	GYFDEPEDVETK		1	2,52	2	1428,61565	-0,66	714,81146	21,22	0	
<b>AT2G42500</b>	<b>PP2A-3</b>	<b>28,75</b>	<b>36,74</b>	<b>1</b>	<b>8</b>	<b>313</b>					
Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	ΔM [ppm]	m/z [Da]	RT [min]	# Missed Cleavages	
High	GAGYTFGQDISEQFNHTNNLK		1	4,78	2	2341,08257	-0,80	1171,04492	22,30	0	
High	SPVTIcGDIHQFHDLAEFLR	C6(Carbamidomethyl)	1	3,71	3	2412,17	-0,80	804,73	24,54	0	
High	cGNnASLLEVDDCR	Carbamidomethyl); M4(Oxidation); C13(Carbamidomethyl)	1	3,22	2	1655,68	0,22	828,34	20,96	0	
High	AKEILmDESNVQPVK	M6(Oxidation)	1	3,18	2	1716,88	-1,73	858,94	18,90	1	
High	IGGMcPDTNYLFmGDYYVDR	M4(Oxidation); C5(Carbamidomethyl); M13(Oxidation)	1	2,92	2	2255,94	-0,86	1128,47	23,08	0	
High	NHTFIQFEPAPR		1	2,78	2	1456,73	-1,03	728,87	20,98	0	
High	EILmDESNVQPVK	M4(Oxidation)	1	2,56	2	1517,75	-1,19	759,38	19,86	0	
High	VVTIIFASPNCYR	C11(Carbamidomethyl)	1	2,47	2	1589,78	-0,54	795,39	23,10	0	
<b>AT3G05750</b>	<b>TRM6</b>	<b>25,87</b>	<b>13,23</b>	<b>7</b>	<b>7</b>	<b>801</b>					
Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	ΔM [ppm]	m/z [Da]	RT [min]	# Missed Cleavages	
High	SDSSCSTPTPTSDDGGSK	C5(Carbamidomethyl)	1	4,16	2	1903,74394	-1,00	952,37561	16,26	0	
High	LmGLESIPVPNALEPR	M2(Oxidation)	1	3,82	2	1751,93328	-1,63	876,47028	23,88	0	
High	QNQFAETSVSNQR		1	3,31	2	1508,70842	-0,29	754,85785	18,55	0	
High	dmDVISFTFSSPIK	D1(Methyl); M2(Oxidation)	2	3,30	2	1616,78630	-0,87	808,89679	24,57	0	
High	DmDVISFTFSSPIK	M2(Oxidation)	1	3,12	2	1602,77104	-0,64	801,88916	25,04	0	
High	SKKPANGVQEAGVNNSDK		1	2,98	3	1728,88565	-1,12	576,96674	15,81	1	

	High	GQSPQISNGTcNNK	C11(Carbamidomethyl)	1	2,63	2	1504,68108	0,09	752,84418	16,73	0
	High	FSSSDSSSSLPmK	M12(Oxidation)	1	2,55	2	1375,60356	-0,79	688,30542	18,19	0
<b>AT1G74160</b>	<b>TRM4</b>	<b>23,22</b>	<b>9,37</b>	<b>7</b>	<b>7</b>	<b>1025</b>					
	Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	ΔM [ppm]	m/z [Da]	RT [min]	# Missed Cleavages
	High	SLQQVDDQLSQASNESR		1	4,73	2	1904,89250	-1,05	952,94989	20,71	0
	High	SGNWADFSGEmSGLVLVER	M11(Oxidation)	1	3,99	2	2184,98296	-1,73	1092,99512	24,41	0
	High	YISEILLASGLLLR		1	3,44	2	1560,93437	-1,06	780,97083	27,60	0
	High	LSESFSEcSSSSK	C9(Carbamidomethyl)	1	3,29	2	1521,63555	-1,22	761,32141	18,69	0
	High	ELcSAIETQK	C3(Carbamidomethyl)	1	2,85	2	1306,62993	-0,66	653,81860	19,09	0
	High	SPSVIEAK		1	2,54	2	901,49932	0,38	451,25330	18,43	0
	High	QQQSTNFAVQR		1	2,38	2	1306,64934	-0,41	653,82831	18,56	0
<b>AT1G25490</b>	<b>PP2A1</b>	<b>17,70</b>	<b>13,10</b>	<b>1</b>	<b>5</b>	<b>588</b>					
	Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	ΔM [ppm]	m/z [Da]	RT [min]	# Missed Cleavages
	High	LLNPELAIQHTLPCVK	C14(Carbamidomethyl)	1	3,78	2	1858,06035	-0,88	929,53381	24,89	0
	High	LAIIEYVPLLASQLGIGFFDdk	D21(Methyl)	4	2,94	3	2436,33982	-1,14	812,78479	31,43	0
	High	LLRDNEAEVR		1	2,93	2	1214,64739	-1,20	607,82733	17,65	1
	High	ELSSDSQHVR		1	2,59	2	1244,58501	-1,29	622,79614	16,21	0
	High	YFANQALNSIDGSTAAQS		1	2,54	2	1857,86443	1,62	929,43585	23,14	0
<b>AT5G57580</b>	<b>Calmodulin-binding protein</b>	<b>11,50</b>	<b>6,96</b>	<b>3</b>	<b>3</b>	<b>647</b>					
	Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	ΔM [ppm]	m/z [Da]	RT [min]	# Missed Cleavages
	High	EGVGTGELVFTDNSSWIR		1	4,41	2	2080,03520	0,26	1040,52124	26,17	0
	High	LTAEGINTVEDFLR		1	3,87	2	1577,81584	-0,76	789,41156	24,82	0
	High	NLDGNDDQPER		1	3,22	2	1387,57158	-0,38	694,28943	17,51	0
<b>AT1G63670</b>	<b>TRM12</b>	<b>5,77</b>	<b>3,92</b>	<b>2</b>	<b>2</b>	<b>689</b>					
	Peptide confidence	Sequence	Modifications	Rank	XCorr	Charge	MH+ [Da]	ΔM [ppm]	m/z [Da]	RT [min]	# Missed Cleavages
	High	IIGLSFDPPNIDLIEK		1	2,91	2	1688,87175	-1,47	844,93951	24,77	0
	High	TSDVSTVGR		1	2,85	2	1246,60198	-0,23	623,80463	16,47	0

Protein identification details were obtained with the LTQ Orbitrap Velos (Thermo Fisher Scientific) and the Proteome Discoverer software package version 1.3 (Thermo Fisher Scientific) combined with search engine SEQUEST.

Proteins and peptides headers generated by the software package and used in the table are listed below.

**Score:** Displays the protein score, which is the sum of the scores of the individual peptides. For SEQUEST results, the score is the sum of all peptide Xcorr values above the specified score threshold. The score threshold is calculated as follows:  $0.8 + \text{peptide\_charge} \times \text{peptide\_relevance\_factor}$  where peptide\_relevance\_factor is an advanced parameter of the SEQUEST node in the "Protein Scoring Option" category with a default value of 0.4. For each spectrum, only the highest-scoring match is used. For each spectrum and sequence, the Proteome Discoverer application uses only the highest scored peptide. When it performs a search using dynamic modifications, one spectrum might have multiple matches because of permutations of the modification site. **Coverage:** Displays by default the percentage of the protein sequence covered by identified peptides. **# Unique Peptides:** Displays the number of peptide sequences unique to a protein group. **# Peptides:** Displays the number of distinct peptide sequences in the protein group. **# AAs:** Shows by default the sequence length of the protein. **Peptide Confidence:** Indicates a confidence level associated with the peptide sequence: high confidence (>99%), medium confidence (>95%, <99%), low confidence (<95%).

**Sequence:** Displays the sequence of amino acids that compose the peptide. **Modifications:** Displays the static and dynamic modifications identified in the peptide. **XCorr (search-dependent):** Scores the number of fragment ions that are common to two different peptides with the same precursor mass and calculates the cross-correlation score for all candidate peptides queried from the database (SEQUEST searches only).

**Rank:** Displays the ordering of peptides by rank. **# Missed Cleavages:** Displays the number of cleavage sites in a peptide sequence that a cleavage reagent (enzyme) did not cleave. This number excludes cases where an amino acid such as proline inhibits the cleaving enzyme. For example, if proline resides next to lysine or arginine, trypsin does not cleave the lysine or arginine. The # of Missed Cleavages value is 0 for a complete digest of all peptides. **Charge:** Displays the charge state of the peptide. **m/z [Da]:** Displays the mass-to-charge ratio of the precursor ion, in daltons. **MH+ [Da]:** Displays the protonated monoisotopic mass of the peptides, in daltons. **ΔM [ppm]:** Displays the difference between the theoretical mass of the peptide and the experimental mass of the precursor ion. **RT [min]:** Displays the retention time when the peptide was observed, in minutes.

**Supplementary Table S3: VELOS orbitrap analysis of TAP TON1a and FASS eluates**

TON1A	Accession	Description	Score	Coverage % Unique Peptide # Peptides # AAs													
				Modifications				Rank	Peptide score	Expect	Charge	Mr(Exp)	Mr(Calc)	ΔM [Da]	m/z [Da]	# Missed Cleavages	
TON1A	AT5G57580	Calmodulin-binding protein	313					14.8	7	8	647						
Peptide confidence		Sequence						Rank	Peptide score	Expect	Charge	Mr(Exp)	Mr(Calc)	ΔM [Da]	m/z [Da]	# Missed Cleavages	
TON1A	High	LTAEGINTVEDFLR			1	105.31	2.20E-09	2	1576.809	1576.81	-3E-04	789.412			0		
TON1A	High	EGVGTLQELVFTDNSSWIR			1	91.18	4.80E-08	2	2079.028	2079.027	4E-04	1040.521			0		
TON1A	High	LYYYAEDSR			1	78.34	7.10E-07	2	1291.61	1291.608	0.002	646.8123			0		
TON1A	High	MWDALVEHAK			1	68.57	5.60E-06	2	1198.58	1198.581	0	600.2975			0		
TON1A	High	LCSLPLEPLR	C2(Carbamidomethyl)		1	59.75	6.50E-05	2	1186.638	1186.638	-3E-04	594.3261			0		
TON1A	High	AVVYGEPEASAK			1	51.35	5.10E-04	2	1090.565	1090.566	-8E-04	546.2898			0		
TON1A	High	LSPLPFTGGK			1	44.04	8.10E-04	2	1031.601	1031.602	-1E-04	516.808			0		
TON1A	High	HYPPLALNDVVVR			1	38.77	7.20E-03	2	1481.704	1481.705	-0.001	741.8592			0		
TON1A																	
TON1A	Accession	Description	Score														
TON1A	AT3G53540	TRM19	307					11.8	9	9	924						
Peptide confidence		Sequence						Rank	Peptide score	Expect	Charge	Mr(Exp)	Mr(Calc)	ΔM [Da]	m/z [Da]	# Missed Cleavages	
TON1A	High	SHSSYNSPESVPTPSLSK			1	141.16	4.30E-13	2	2005.959	2005.959	2E-04	1003.987			0		
TON1A	High	ELQWLSEDDIEIGR			1	105.19	1.50E-09	2	1927.988	1927.988	-0.001	965.0011			0		
TON1A	High	LLFDQISR			1	57.35	7.70E-05	2	990.5495	990.5495	-3E-04	496.282			0		
TON1A	High	EGDQPSPVSVLEASFDDDVSSGSECFSVSADLR	C25(Carbamidomethyl)		1	54.53	5.40E-05	3	3617.568	3616.564	1.005	1206.863			0		
TON1A	High	MQLQLLK			1	54.28	2.10E-04	2	872.5149	872.5153	-4E-04	437.2647			0		
TON1A	High	KLLFDQISR	C3(Carbamidomethyl)		1	47.98	3.90E-04	2	1118.644	1118.645	-3E-04	560.3295			1		
TON1A	High	LPCTTTHGR			1	45.24	8.50E-04	2	1041.502	1041.503	-6E-04	521.7583			0		
TON1A	High	IQETLR			1	39.35	7.20E-03	2	758.4288	758.4286	2E-04	380.2217			0		
TON1A	High	FEHEIEISR			1	36.28	9.20E-03	2	1158.567	1158.567	-4E-04	580.2905			0		
TON1A																	
TON1A	Accession	Description	Score														
TON1A	AT2G42500/AT3G5850	PP2A3/PPI2A4	203					31	8	8	313						
Peptide confidence		Sequence						Rank	Peptide score	Expect	Charge	Mr(Exp)	Mr(Calc)	ΔM [Da]	m/z [Da]	# Missed Cleavages	
TON1A	High	QITQVYGFYDECLR			C12(Carbamidomethyl)	1	86.57	1.00E-07	2	1790.828	1790.828	-0.002	896.4212			0	
TON1A	High	NHTIFQFEPAPR			1	64.52	1.80E-05	2	1455.726	1455.726	-4E-04	728.87			0		
TON1A	High	VVTIFASPNYCYR	C11(Carbamidomethyl)		1	63.31	2.60E-05	2	1588.77	1588.771	-0.001	795.3921			0		
TON1A	High	EILMDESNVOPVK	M4(Oxidation)		1	47.99	9.40E-04	2	1516.744	1516.744	-8E-04	759.379			0		
TON1A	High	YGNANVWK			1	45.9	1.40E-03	2	950.4602	950.461	-8E-04	476.2374			0		
TON1A	High	RGEPDVTR			1	41.29	2.00E-03	2	928.4738	928.4726	0.001	465.2442			1		
TON1A	High	GAGYTFGQDISEQFNHTNNLK			1	40.63	3.50E-03	3	2340.075	2340.077	-0.002	781.0324			0		
TON1A	High	PLSEQQR			1	40.4	2.60E-03	2	955.5088	955.5087	1E-04	478.7617			0		
TON1A																	
TON1A	Accession	Description	Score														
TON1A	AT3G05750	TRM6	179					6.4	4	4	801						
Peptide confidence		Sequence						Rank	Peptide score	Expect	Charge	Mr(Exp)	Mr(Calc)	ΔM [Da]	m/z [Da]	# Missed Cleavages	
TON1A	High	EFSLGMATDILPLSLDFTEGK			C12(Carbamidomethyl)	1	101.6	4.10E-09	2	2412.175	2412.177	-0.002	1207.095			0	
TON1A	High	QNQFAETSVSNQR			1	83.53	2.10E-07	2	1507.702	1507.702	-0.002	754.8571			0		
TON1A	High	QDIFLER			1	66.95	1.60E-05	2	919.4759	919.4763	-4E-04	460.7452			0		
TON1A	High	EILADQVLK			1	36.03	6.30E-03	2	1027.591	1027.591	-5E-04	514.8027			0		
TON1A																	
TON1A	Accession	Description	Score														
TON1A	AT5G26910	TRM8	86					4.6	2	2	853						
Peptide confidence		Sequence						Rank	Peptide score	Expect	Charge	Mr(Exp)	Mr(Calc)	ΔM [Da]	m/z [Da]	# Missed Cleavages	
TON1A	High	ETYEEGIDIEGEIVSTLVDDLVNLDVSGTQR			C2(Carbamidomethyl); C12(Carbamidomethyl); C17(Carbamidomethyl)	1	78.91	5.00E-07	3	3407.643	3407.647	-0.003	1136.888			0	
TON1A	High	TLDFVNK			1	37.73	5.20E-03	2	982.5128	982.5124	4E-04	492.2637			0		
TON1A																	
TON1A	Accession	Description	Score														
TON1A	AT2G45240	MAP1A	50					5.3	2	2	398						
Peptide confidence		Sequence						Rank	Peptide score	Expect	Charge	Mr(Exp)	Mr(Calc)	ΔM [Da]	m/z [Da]	# Missed Cleavages	
TON1A	High	HATMSGLSVR			M4(Oxidation)	1	41.17	4.30E-03	2	1172.595	1172.597	-0.002	587.3049			0	
TON1A	High	AIAIVKPGVR			1	35.45	4.00E-04	2	1022.66	1022.66	-2E-04	512.3372			1		
TON1A																	
FASS	Accession	Description	Score														
FASS	AT5G18580	FASS	3906					81	32	32	480						
Peptide confidence		Sequence						Rank	Peptide score	Expect	Charge	Mr(Exp)	Mr(Calc)	ΔM [Da]	m/z [Da]	# Missed Cleavages	
FASS	High	YIGSGAGLGSEALMELET				1	157.28	1.20E-14	2	1924.945	1924.945	2E-04	963.48			0	
FASS	High	QSDLLNADDLAAMWVCLR			C17(Carbamidomethyl)	1	146.63	1.40E-13	2	2203.028	2203.077	0.004	1102.547			0	
FASS	High	MNYEDFCCHIASVCTEQIPK			1	130.67	1.10E-12	2	2414.032	2414.034	-0.002	1208.023			0		
FASS	High	EMDFCLDFVLALENK			1	123	2.60E-11	2	2031.951	2031.95	7E-04	1016.983			0		
FASS	High	SQEAQTIPSFKK			1	114.05	1.90E-10	2	1326.645	1326.646	-0.001	664.3296			0		
FASS	High	OCTGTVASMLIDVR			1	107.68	1.20E-09	2	1345.702	1345.702	-2E-04	673.8584			0		
FASS	High	ICDMFLALDKDMGSSLCK	C2(Carbamidomethyl); C12(Carbamidomethyl); C17(Carbamidomethyl)		1	107.63	4.80E-10	2	2118.946	2118.946	0	1060.49			1		
FASS	High	GFLTTADHSLSR			1	100.06	6.00E-09	2	1476.772	1476.773	-2E-04	739.3934			0		
FASS	High	EYADGTLTEFIER			1	99.64	5.70E-09	2	1655.804	1655.804	-8E-04	828.909			0		
FASS	High	ENCVIDDATGAEK	C3(Carbamidomethyl); E12(Methyl)		1	89.89	2.50E-08	2	1434.628	1434.623	-0.001	718.3215			0		
FASS	High	WIEGGNYELCIEDVREIDWMVK	E8(Methyl); C10(Carbamidomethyl)		1	87.5	7.90E-08	2	1865.861	1865.862	-8E-04	933.9378			0		
FASS	High	.SNCLQELMHQSEEEVTDQEAEWNFLSLTA	C6(Carbamidomethyl); M11(Oxidation)		1	84.2	2.00E-07	2	1324.669	1324.666	0.003	663.342			0		
FASS	High	IPPAASMLWVR	C2(Carbamidomethyl)		1	79.08	8.30E-07	2	1088.59	1088.59	-2E-04	545.3022			0		
FASS	High	KPEEGSISQR	M7(Oxidation)		1	75.28	1.10E-06	3	2868.31	2868.31	4E-04	957.1107			1		
FASS	High	IDMSELDEDGGFLHSDEMESYIGGLPNAQLR	M19(Oxidation)		1	72.75	1.20E-06	3	4523.093	4523.074	0.019	1508.705			0		
FASS	High	IAIPLFYLYVMR	M11(Oxidation)		1	71.65	3.60E-06	2	1224.588	1224.588	-2E-04	613.3013			0		
FASS	High	DEIWDMVKPSDPDK	C5(Carbamidomethyl)		1	69.56	5.80E-06	2	1271.67	1271.67	1E-04	636.8421			0		
FASS	High	FFFCCDPHR			1	67.63	8.40E-06	2	1129.573	1129.573	0	565.7936			1		
FASS	High	VFDDEHVR			1	67.21	4.60E-06	3	3824.738	3824.74	-0.002	1275.92			0		
FASS	High	ILLEIFK			1	59.31	4.00E-05	2	1513.836	1513.837	-5E-04	757.9254			0		
FASS	High	DMPPPAFNQMYCR	C11(Carbamidomethyl)		1	55.9	1.40E-04	2	1671.818	1671.818	-2E-04	836.9161			1		
FASS	High	FFSPSPNFMK			1	52.39	1.20E-04	2	1271.554	1271.555	-4E-04	636.7844			0		
FASS	High				1	52.06	1.80E-04	2	900.445	900.4454	-8E-04	451.2295			0		
FASS																	



FASS	High	YMVANOLYELCEAVGPEPTR	M2(Oxidation); C11(Carbamidomethyl)	1	138.34	6.20E-13	2	2355.087	2355.088	-4E-04	1178.551	0		
FASS	High	IINPFLAIQHILPCVK	C14(Carbamidomethyl)	1	89.37	1.40E-08	2	1857.054	1857.055	-9E-04	929.5341	0		
FASS	High	AVSLAPVGMSEITCSK	C15(Carbamidomethyl)	1	87.06	1.40E-07	2	1761.901	1761.901	3E-04	881.9576	0		
FASS	High	LDQVNQVIGIDLSQSSLPAIVELAEDR		1	85.5	3.40E-08	3	3060.67	3060.671	-4E-04	1021.231	0		
FASS	High	SALASVIMGMAPVLK	M8(Oxidation)	1	76.94	9.30E-07	2	1559.842	1559.842	3E-04	780.9282	0		
FASS	High	LLEPQDCV AHLPVIVNFSDQK	C7(Carbamidomethyl)	1	67.41	7.60E-06	3	2534.319	2534.32	-0.001	845.7803	0		
FASS	High	LSAGEWFTAR		1	63.26	2.80E-05	2	1136.561	1136.561	-3E-04	569.2879	0		
FASS	High	ELSSDSSQQHVR		1	61.6	2.20E-05	2	1243.578	1243.579	-0.001	622.7964	0		
FASS	High	ESDLVEHFTPLAK		1	56.29	1.10E-04	2	1484.751	1484.751	-4E-04	743.3827	0		
FASS	High	FAATIESAHLK		1	53.26	2.90E-04	2	1186.634	1186.635	-3E-04	594.3244	0		
FASS	High	SIYGLQLCDDMPMVR	C7(Carbamidomethyl); M11(Oxidation); M13(Oxidation)	1	48.29	2.30E-04	2	1843.788	1843.789	-0.002	922.9013	0		
FASS	High	LLPAVITASK		1	29.37	5.30E-03	2	1011.633	1011.633	-3E-04	506.8235	0		
FASS	Accession AT3G58500	Description PP2A4	Score 477	Modifications	Coverage % Unique Peptide # Peptides # AAs									
FASS		Sequence	40.9		4.0	2	10	313						
FASS	Peptide confidence	GAGYTFGODISEQFNHTNNLK			Rank	Peptide score	Expect	Charge	Mr(Exp)	Mr(Calc)	ΔM [Da]	m/z [Da]	# Missed Cleavages	
FASS		EILMDENSNVOPVK	M4(Oxidation)		1	118.69	5.40E-11	2	2340.075	2340.077	-0.002	1171.045	0	
FASS		CPDTNTYLFMGDYVDR	C1(Carbamidomethyl); M9(Oxidation)		1	98.09	9.30E-09	2	1516.743	1516.744	-0.002	759.3786	0	
FASS		QITQVYGFYDECLR	C12(Carbamidomethyl)		1	97.65	1.40E-09	2	1880.77	1880.771	-0.001	941.392	0	
FASS		VVTIFSPAPNYCYR	C11(Carbamidomethyl)		1	89.96	4.60E-08	2	1790.83	1790.83	0	896.4222	0	
FASS		YGNANVWK			1	66.82	1.20E-05	2	1588.77	1588.771	-0.001	795.3921	0	
FASS		NHTFIQFEPAPR			1	66.68	1.10E-05	2	950.4605	950.461	-5E-04	476.2375	0	
FASS		PLSEQQVR			1	64.58	1.80E-05	2	1455.726	1455.726	-4E-04	728.87	0	
FASS		RGEPDVTR			1	52.18	1.80E-04	2	955.5086	955.5087	-1E-04	478.7816	0	
FASS		AHOLVMDGFNWAHEQK	M6(Oxidation)		1	50.65	2.60E-04	2	928.4722	928.4726	-5E-04	465.2434	1	
FASS		TRM8	42.95		2.00E-03	3	1925.883	1925.884	-9E-04	642.9684	0			
FASS	Accession AT5G26910	Description	Score 366	Modifications	Coverage % Unique Peptide # Peptides # AAs									
FASS		Sequence	22.3		13	13	853							
FASS		ETYEEGIDIEGIEVISTLVDDLVNDLVSQTQR			Rank	Peptide score	Expect	Charge	Mr(Exp)	Mr(Calc)	ΔM [Da]	m/z [Da]	# Missed Cleavages	
FASS		SDSSCCASSVTSDDGGOTR	C5(Carbamidomethyl); C6(Carbamidomethyl)		1	105.63	1.20E-09	3	3408.656	3407.647	1.009	1137.226	0	
FASS		EYALGMATDVPLASLDEMEGR	M6(Oxidation); M19(Oxidation)		1	83.18	7.70E-09	2	1975.749	1975.748	9E-04	988.8819	0	
FASS		DNOPSMTSVLNQK	M6(Oxidation)		1	77.96	5.30E-07	2	2446.108	2446.103	0.004	1224.061	0	
FASS		EGVSVSALVEPHLSSAF			1	76.26	1.20E-06	2	1476.688	1476.688	4E-04	739.3514	0	
FASS		OQLAAELSAEK			1	63.41	1.60E-05	2	1666.894	1666.893	8E-04	834.4542	0	
FASS		DWLAEEELNR			1	64.88	2.60E-04	2	1103.582	1103.582	-8E-04	562.709	0	
FASS		VSLIEVDEIGK			1	47.62	7.20E-04	2	1144.551	1144.551	1E-04	573.283	0	
FASS		GGFLFEQR			1	46.62	1.30E-03	2	1200.66	1200.66	-6E-04	601.3371	0	
FASS		CEQMFMGSCR	: carbamidomethyl); M4(Oxidation); M6(Oxidation); C9(Carbamidomethyl)		1	41.95	2.90E-03	2	952.4771	952.4767	5E-04	477.2458	0	
FASS		ITSLTLPSTSK			1	40.77	3.90E-03	2	1336.466	1336.467	-4E-04	669.2405	0	
FASS		VPVESGSISK			1	40.69	5.40E-03	2	1001.539	1001.533	-6E-04	501.7766	0	
FASS		QNSNDTFLNK			1	38.21	7.60E-03	2	1179.551	1179.551	-7E-04	590.7829	0	
FASS	Accession AT2G45240	Description	Score 270	Modifications	Coverage % Unique Peptide # Peptides # AAs									
FASS		MAP1A	26.4		8	8	398							
FASS		Sequence	Rank		Peptide score	Expect	Charge	Mr(Exp)	Mr(Calc)	ΔM [Da]	m/z [Da]	# Missed Cleavages		
FASS		VEPNSDLQHVEIK	1	83.72	2.70E-07	2	1605.834	1605.836	-0.002	803.9243	0			
FASS		LPSSCPDPVWLTK		1	79.16	7.90E-07	2	1501.781	1501.782	-4E-04	751.8979	0		
FASS		VVPAEIIPDWIAIDGTPK		1	76.85	9.50E-07	2	1964.024	1964.026	-0.001	983.0194	1		
FASS		SAQFHTLIVTETGVETLAR		1	74.98	1.20E-06	3	2300.204	2300.201	0.003	767.7419	0		
FASS		VIHPGVTTDEIDR		1	54.41	2.40E-04	2	1450.741	1450.742	-9E-04	726.3776	0		
FASS		EVLDAAAR		1	52.36	2.00E-04	2	843.4437	843.445	-0.001	422.7292	0		
FASS		EIGEIVNR		1	44.47	1.20E-03	2	928.4976	928.4978	-1E-04	465.2561	0		
FASS		AIAIVKPGVR		1	36.16	3.40E-04	2	1022.66	1022.66	-3E-04	512.3372	1		
FASS	Accession AT3G05750	Description	Score 190	Modifications	Coverage % Unique Peptide # Peptides # AAs									
FASS		TRM6	7.9		6	6	801							
FASS		Sequence	Rank		Peptide score	Expect	Charge	Mr(Exp)	Mr(Calc)	ΔM [Da]	m/z [Da]	# Missed Cleavages		
FASS		QNQFAETSVSNQR	1	64.48	1.80E-05	2	1507.702	1507.702	3E-04	754.8582	0			
FASS		FSSSSCASSPLPMK	M12(Oxidation)	1	58.48	2.40E-05	2	1374.596	1374.597	-9E-04	688.3055	0		
FASS		QDFIFER		1	53.34	3.70E-04	2	919.4763	919.4763	0	460.7455	0		
FASS		SPGFVQSR		1	46.03	2.00E-03	2	876.4449	876.4454	-4E-04	439.2298	0		
FASS		CEQMFMGTC	: carbamidomethyl); M4(Oxidation); M6(Oxidation); C9(Carbamidomethyl)	1	40.14	9.70E-05	2	1322.476	1322.476	-3E-04	662.2453	0		
FASS	Accession AT4G28760	Description	Score 186	Modifications	Coverage % Unique Peptide # Peptides # AAs									
FASS		TRM20	6		4	4	924							
FASS		LLQELEVAVIDLTR			Rank	Peptide score	Expect	Charge	Mr(Exp)	Mr(Calc)	ΔM [Da]	m/z [Da]	# Missed Cleavages	
FASS		INSIVSETTTR	1	112.2	1.20E-10	2	1739.968	1739.967	0.001	870.9912	0			
FASS		VSNLFFFK		1	90.69	5.40E-08	2	1320.687	1320.689	-0.001	661.3551	0		
FASS		GTEEHTVQPTR		1	52.73	2.70E-04	2	1000.538	1000.538	-1E-04	501.2763	0		
FASS			1	38.82	6.30E-03	2	1253.599	1253.6	-0.001	627.8068	0			
FASS	Accession AT1G63670	Description	Score 131	Modifications	Coverage % Unique Peptide # Peptides # AAs									
FASS		TRM12	7.3		4	4	689							
FASS		Sequence	Rank		Peptide score	Expect	Charge	Mr(Exp)	Mr(Calc)	ΔM [Da]	m/z [Da]	# Missed Cleavages		
FASS		IGLSFDDPPNIDLIEK	1	72.17	3.80E-06	2	1687.865	1687.867	-0.002	844.9397	0			
FASS		TSVDSTVGHVR		1	66.85	9.50E-06	2	1245.595	1245.595	-1E-04	623.8047	0		
FASS		VLNLESSLR		1	53.7	2.30E-04	2	1029.584	1029.582	0.002	515.7993	0		
FASS		QEQQPSPVSLER		1	43.53	2.70E-03	2	1624.803	1624.806	-0.003	813.4088	0		
FASS	Accession AT1G74160	Description	Score 85	Modifications	Coverage % Unique Peptide # Peptides # AAs									
FASS		TRM4	2.3		2	2	1025							
FASS		Sequence	Rank		Peptide score	Expect	Charge	Mr(Exp)	Mr(Calc)	ΔM [Da]	m/z [Da]	# Missed Cleavages		
FASS		SENFLLEEEEDDFLK	1	81.76	2.80E-07	2	1726.795	1726.794	0.002	864.4049	0			
FASS		SILAEDVTIR		1	39.09	6.50E-03	2	1115.62	1115.619	0.002	558.8174	0		

Protein identification details were obtained with the LTQ Orbitrap Velos (Thermo Fisher Scientific) and Mascot Distiller software (version 2.3.2.0, Matrix Science) combined with the Mascot search engine (Matrix Science) using the Mascot Daemon interface (version 2.4, Matrix Science).

Proteins and peptides headers used in the table are listed below.

**Score:** Displays the protein score. **Coverage %:** Displays by default the percentage of the protein sequence covered by identified peptides. # **Unique Peptides:** Displays the number of peptide sequences unique to a protein group. # **Peptides:** Displays the number of distinct peptide sequences in the protein group. # **AAs:** Shows by default the sequence length of the protein. **Peptide Confidence:** Indicates a confidence level associated with the peptide sequence: high confidence (>99%), medium confidence (>95%, <99%), low confidence (<95%). **Sequence:** Displays the sequence of amino acids that compose the peptide. **Modifications:** Displays the static and dynamic modifications identified in the peptide. **Rank:** Displays the ordering of peptides by rank. **Peptide score:** Displays the peptide ions score. **Expect:** Expectation value for the peptide match. (The number of times we would expect to obtain an equal or higher score, purely by chance. The lower this value, the more significant the result.) **Charge:** Displays the charge state of the peptide. **Mr(Exp):** experimental relative molecular mass, in daltons. **Mr(Calc):** calculated relative molecular mass, in daltons. **ΔM [Da]:** difference (error) between the experimental and calculated masses, in daltons. **m/z [Da]:** Displays the mass-to-charge ratio of the precursor ion, in daltons. # **Missed Cleavages:** Displays the number of cleavage sites in a peptide sequence that a cleavage reagent (enzyme) did not cleave. This number excludes cases where an amino acid such as proline inhibits the cleaving enzyme. For example, if proline resides next to lysine or arginine, trypsin does not cleave the lysine or arginine. The # of Missed Cleavages value is 0 for a complete digest of all peptides.

**Supplementary Table S4: List of primers used in this study**

Primer name	Construct	Sequence
TON1a_fwd		published Van Leene et. al. 2007
TON1a_nostop_rev		published Van Leene et. al. 2007
TON1a_stop_rev		AGAAAGCTGGGTCTCAATCTCCTCTTCTTC
TON1b_fwd		AAAAAGCAGGCTCCACCATGGACATTATAAGAGAG
TON1b_stop_rev		AGAAAGCTGGGTCTCAACTTTCCCTTCTCTTC
TON1b_nostop_rev		AGAAAGCTGGGTCTCATCTTCCCTTCTCTTC
FASS_fwd	Entry vectors of TON1a, TON1b, FASS and TRM19 to generate TAP constructs	AAAAGCAGGCTCCACCATGATAGCGGATCTAGCGATGG
FASS_stop_rev		AGAAAGCTGGGTCTCACTGAGACTCTCCCTCAGG
FASS_nostop_rev		AGAAAGCTGGGTCTCTAGAAGATGACCTACCAAGC
AT3G53540_fwd		AGAAAGCTGGGTCTCGAAGATTGCACCTACCAAGC
AT3G53540_stop_rev		GGGGACAATTGTACAAAAAAAGCAGGCTCACC
AT3G53540_nostop_rev		GGGGACCACTTGTACAAAGAACGCTGGGT
attB1-F		GGGGACAATTGTACAAAAAAAGCAGGCTACACAGA
attB2-R		GGGGACAATTGTACAAAAAAAGCAGGCTACACAGA
FASSGW (+1)		GGGGACAATTGTACAAAAAAAGCAGGCTACAAATGTATAGCGGATCTAG
FASSGW (-480)		GGGGGACCACTTTGTACAAAGAAAGCTGGGTACTGAGACTCTCCCTCAG
FASSGW (-185)		GGGGGACCACTTTGTACAAAGAAAGCTGGGTCTCGGCGATCACATAAAG
FASSGW (+186)		GGGGGACAATTGTACAAAAAAAGCAGGCTACAAATGGTGTGCGCTTACACAGC
TRM1 (+1)	Entry vectors of FASS and truncated versions of FASS	GGGGACAATTGTACAAAAAAAGCAGGCTACAAATGTATAGCGGATCTAG
TRM1 (+587)		GGGGACAATTGTACAAAAAAAGCAGGCTACAAATGAGCTGGGATCGAATGTT
TRM1 (-698)		GGGGGACCACTTTGTACAAAGAAAGCTGGGTATTCCCCTGACTCTGAGG
TRM1 (-827)		GGGGGACCACTTTGTACAAAGAAAGCTGGGTATTAAAGTGGGATTTC
TRM1 (-905)		GGGGGACCACTTTGTACAAAGAAAGCTGGGTAGCAGAAGCAAACCTCAT
TON1a (+1)	Entry vectors of TRM1 and truncated versions of TRM1	GGGGACAATTGTACAAAAAAAGCAGGCTTGTGAGGATTACAAAG
TON1a (-107)		GGGGGACCACTTTGTACAAAGAAAGCTGGGTACGAATCTTGGCAA
TON1a (+108)		GGGGGACCAATTGTACAAAAGCAGGCTACAAATGTGAAGCTGAGTTACG
TON1a (-260)		GGGGGACCACTTTGTACAAAGAAAGCTGGGTATTCCCCTCTTC
RCN1GW FWD		GGGGGACAATTGTACAAAAAAAGCAGGCTACAAATGGTATGGTAGATGAA
RCN1GW REV	PP2AA1 entry vector	GGGGGACCAATTGTACAAAGAAAGCTGGTAGGATTGTGCTGCTGTTG
TRM8 (+1)		GGGGGACAATTGTACAAAGAAAGCTGGTAGAATGGAGGTTTGAGG
TRM8 (-854)		GGGGGACCACTTTGTACAAAGAAAGCTGGTAGAATGCCGAGAGAAGAT
R1		AGCACATCTCCCTTGTGTGAAAGG
R2		AACTTGCTTATGATGTTAAGGC
T (T-DNA)	Genotyping of <i>pp2aa1</i> mutant	TGTCGGCGGTATCGGGGGGGTC
oA2RAV		GCGTGCCTGTCCTCTTGAC
oA2new3'		CACATTAGTAGCAAGAACATGGACAAACCCG
oA3leader		GATCGCTCGGAACTTGGAAGCAGC
oA3DDEV		GCCAAAGCACCTCATCGTCATCGC
SALK_069250 RP	Genotyping of <i>pp2aa2</i> mutant	TGTTCTCTGATCTGTTCTCG
SALK_069250 LP		TAATTGGTATCAGGGCACTGC
SALK_035009 RP		GTGGATTATCACCATCATCG
SALK_035009 LP		GCTTGAAGAACAGCATTTCG
LB-Salk2		GCTTCTCTCTCTCTCTCT
ACT2_2_FWD	RT-PCR primers for actin, eif4A, PP2Ac1-c5 and PP2AA1-A3	TTGACTACGAGCAGGAGATGG
ACT2_2_REV		ACAAACGAGGGCTGAAACAG
eif4A-FWD		ACGGAGACATGGACAGAAC
eif4A-REV		GCTGAGTTGGGAGATCGAAG
at3g58500_LP1		TCTCGAAAGTATGCGAATG
at3g58500_RP1		CCACCGTGAAGGCAGAATA
at2g42500_LP2		GAACCAACCTTCATCCAGIT
at2g42500_RP2		TGAGATGAGCTGTGTCAGTGA
at3g25800_fwd		CGATGCTGCTGTAACAAATC
at3g25800_rev		ATTGCCCATTCAGGACCA
at1g10430_fwd		CCAGAGAACGAGCTAGGAGGATTGATC
at1g10430_rev		TTGAAAAGCACCACCGATCC
at1g59830_fwd		TATGAGAGAAAACAAAACCAAGACG
at1g59830_rev		TCTCGTCTGTCCTTAAAC
at1g69960_fwd		ATGGACCAAGAATTCTCTCAGTTG
at1g69960_rev		TAGTGTGGAATTGCACTTTAG
at1g13320_fwd		GACCGGAGCCAACTAGGAC
at1g13320_rev		AAAACCTGGTAACCTTCCAGCA
at1g25490-fwd		ATCCITGTTACCGGCCATT
at1g25490-rev		TGCAAGTCGGACTCCAG

## **Supplementary Methods for the LC-MS/MS VELOS1 analysis (related to Supplementary Table S2)**

Tuning of equipment and sample analyses were performed using a standard operation procedure. Quality performance of the LC MS analyses was assessed using control standards. Nanoflow LC-MS/MS analyses were performed on a dual channel NanoLC Ultra 2D system (Eksigent, Dublin, California, USA) and connected to an LTQ Orbitrap Velos mass spectrometer (Thermo, Bremen, Germany) using a chip based nano electrospray source (Triversa Advion Biosystems, Ithaca, NY, USA) operated at 1.8 kV. Mobile phase A ( $\text{CH}_3\text{CN}$  : CHOOH :  $\text{H}_2\text{O}$  in a 2:0.1:97.9 (v/v) ratio) and mobile phase B ( $\text{CH}_3\text{CN}$  : CHOOH :  $\text{H}_2\text{O}$  in a 90:0.1:9.9 (v/v) ratio) were reconstituted from ULC/MS-grade  $\text{CH}_3\text{CN}$ , CHOOH and  $\text{H}_2\text{O}$  (Biosolve-chemicals, Valkenswaard, the Netherlands). All standards and samples were dissolved in mobile phase A. A volume of 10  $\mu\text{L}$  of each sample or control standard was loaded on a  $\text{C}_{18}$  precolumn (PepMap 100, 5  $\mu\text{m}$  particles, 20 mm x 200  $\mu\text{m}$  ID, Dionex, Sunnyvale, California, USA) using a flow rate of 6  $\mu\text{L}/\text{min}$  (100% mobile phase A). Five minutes after injection the trapping column was switched in line with an analytical  $\text{C}_{18}$  column (Acclaim Pepmap 100, 3  $\mu\text{m}$  particles, 150 mm x 75  $\mu\text{m}$  ID, Dionex). Effective elution of peptides was accomplished within a 45 minutes gradient flow (95% A to 50% A) at a flow rate of 350 nL/min.

MS spectra were recorded in the orbitrap FT analyzer with a resolution of 60000 (at m/z 400) and an AGC target setting of 500000. The maximum injection time was set to 500 msec and lock mass was enabled (polysiloxane ion at m/z 445.12024). Collision induced dissociation MS/MS spectra were acquired using the ion trap in data dependent mode selecting up to the 20 most abundant multiply charged precursor ions from the MS spectrum. The maximum injection time was set to 50 msec and an AGC setting of 7500. Fragmentation was accomplished by CID wideband activation at normalized collision energy of 35 eV and a 30msec activation time. After MS/MS the precursor m/z's were excluded for 30 seconds. Peak lists were generated and submitted for protein identification with Proteome Discoverer 1.3.0.339 (Thermo, Bremen, Germany). Spectrum grouping was allowed with a maximum retention time of 1 minute and a precursor mass tolerance of 2 ppm. Peak lists were generated only for MS/MS spectra containing more than 5 peaks, with a total intensity above 50. The relative signal-to-noise limit was set to 5. Peak lists were submitted for protein identification against TAIR10 containing 35386 sequence entries with search engine SEQUEST. Enzyme was set to trypsin, allowing for maximum 1 missed cleavage site. Precursor mass tolerance was set to 3 ppm and fragment mass tolerance at 0.8 Da. Fixed modifications were set to carbamidomethylation of cysteines. Variable modifications were set to methionine oxidation and methylation of aspartic acid and glutamic acid, with a maximum of 4 modifications per peptide. Peptides were validated using a decoy database search. The strict target False Discovery Rate (FDR) was set to 0.01 (or 1%), the relaxed FDR was set to 0.05 (or 5%). Only high confident (>99%) peptides were withheld. Only proteins with at least two matched high confident peptides were retained.

## **Supplementary Methods for the LC-MS/MS VELOS2 analysis (related to Supplementary Table S3)**

The obtained peptide mixtures were introduced into an LC-MS/MS system, the Ultimate 3000 RSLC nano (Dionex, Amsterdam, The Netherlands) in-line connected to an LTQ Orbitrap Velos (Thermo Fisher Scientific, Bremen, Germany). The sample mixture was loaded on a trapping column (made in-house, 100  $\mu\text{m}$  internal diameter (I.D.) x 20 mm (length), 5  $\mu\text{m}$

C18 Reprosil-HD beads, Dr. Maisch GmbH, Ammerbuch-Entringen, Germany). After back-flushing from the trapping column, the sample was loaded on a reverse-phase column (made in-house, 75 µm I.D. x 150 mm, 5 µm C18 Reprosil-HD beads, Dr. Maisch). Peptides were loaded with solvent A (0.1% trifluoroacetic acid, 2% acetonitrile), and separated with a linear gradient from 2% solvent A' (0.1% formic acid) to 50% solvent B' (0.1% formic acid and 80% acetonitrile) at a flow rate of 300 nL/min, followed by a wash step reaching 100% solvent B'.

The mass spectrometer was operated in data-dependent mode, automatically switching between MS and MS/MS acquisition for the ten most abundant peaks in a given MS spectrum. In the LTQ-Orbitrap Velos, full scan MS spectra were acquired in the Orbitrap at a target value of 1E6 with a resolution of 60,000. The ten most intense ions were then isolated for fragmentation in the linear ion trap, with a dynamic exclusion of 20 seconds. Peptides were fragmented after filling the ion trap at a target value of 1E4 ion counts.

From the MS/MS data in each LC run, Mascot Generic Files were created using the Mascot Distiller software (version 2.3.2.0, Matrix Science, [www.matrixscience.com/Distiller.html](http://www.matrixscience.com/Distiller.html)). When generating these peak lists, grouping of spectra was allowed with a maximum intermediate retention time of 30 seconds and a maximum intermediate scan count of 5 was used where possible. Grouping was done with 0.005 Da precursor tolerance. A peak list was only generated when the MS/MS spectrum contained more than 10 peaks. There was no de-isotoping and the relative signal-to-noise limit was set to 2. These peak lists were then searched with the Mascot search engine (MatrixScience, [www.matrixscience.com](http://www.matrixscience.com), PMID 10612281) using the Mascot Daemon interface (version 2.4, Matrix Science). Spectra were searched against the TAIR10 database containing 35386 sequence entries. Variable modifications were set to methionine oxidation and methylation of aspartic acid and glutamic acid. Fixed modifications were set to carbamidomethylation of cysteines. Mass tolerance on MS was set to 10 ppm (with Mascot's C13 option set to 1) and the MS/MS tolerance at 0.5 Da. The peptide charge was set to 1+, 2+ and 3+ and the instrument setting was set to ESI-TRAP. Trypsin was set as the protease used, allowing for 1 missed cleavage, and also cleavage was allowed when arginine or lysine is followed by proline. Only high confident peptides, ranked one and with scores above the threshold score, set at 99% confidence, were withheld. Only proteins with at least two matched high confident peptides were retained.