

Supplementary information S1 (table) | Major features of HUH enzymes

	Group	Protein (element)	Ys	HUH	Biochemistry	Structure
Reps	Phages	GpA (ϕ X174)	Y343 Y347	yes	Y343 and Y347 involved (Van Mansfeld <i>et al.</i> , 1986) Flip-flop mechanism (Hanai and Wang, 1993)	NO
		A (P2)	Y454 Y450	yes	Initiation (Y454) and termination (Y450) Tyr are different (Odegrip and Haggård-Ljungquist, 2001)	NO
		GpII (M13)	Y197	no	No stable covalent complex (Asano <i>et al.</i> , 1999)	NO
		SIRV1 Rep	Y108	yes	Stable covalent complex (Oke <i>et al.</i> , 2011)	2X3G (Oke <i>et al.</i> , 2011) dimer
	Viruses	Rep (AAV5)	Y156	yes	Site-specific endonuclease activity (Im and Muzyczka, 1990)	1M55 (Hickman <i>et al.</i> , 2002) 1RZ9 (Hickman <i>et al.</i> , 2004)
		Rep (TYLCV)	Y103	yes	Cleavage and joining domain (Heyraud-Nitschke <i>et al.</i> , 1995)	1L2V (Campos-Olivas <i>et al.</i> , 2002)
		Rep (FBNYV)	Y79	HUQ	In vitro endonuclease activity (Vega-Rocha <i>et al.</i> , 2007b)	2HWT (Vega-Rocha <i>et al.</i> , 2007b)
		Rep (PCV)	Y96	HUQ	In vitro endonuclease activity (Vega-Rocha <i>et al.</i> , 2007a)	2HW0 (Vega-Rocha <i>et al.</i> , 2007a)
	Plasmids	RepB pMV158	Y99	Yes	Chiral phosphorothioate (Moscoso <i>et al.</i> , 1997)	3DKY (Boer <i>et al.</i> , 2009)
		RepA pC194	Y214 E210	yes	Termination hydrolysis by Glu (Noirot-Gros <i>et al.</i> , 1994) Flip-flop with E210Y (Noirot-Gros and Ehrlich, 1996)	NO

		RepC pT181	Y191	No	Short oligonucleotide inactivation (Rasooly and Novick, 1993)	NO
Mobs	MOB _F	TrwC (R388)	Y18 Y26	yes	Initiation and termination Tyr (Grandoso <i>et al.</i> , 2000) Intramolecular reaction (Gonzalez-Perez <i>et al.</i> , 2007) TrwC transfer to the recipient (Draper <i>et al.</i> , 2005) Termination reaction in the recipient (Garcillán-Barcia <i>et al.</i> , 2007)	1OMH (Guasch <i>et al.</i> , 2003)
		Tral (F)	Y16 Y17		Two contiguous catalytic Tyr (Street <i>et al.</i> , 2003) Differential metal affinity (Larkin <i>et al.</i> , 2005)	1P4D, (Datta <i>et al.</i> , 2003)
		Tral (pCU1)	Y18-19 Y26-27	yes	Tyrosine partners (Nash <i>et al.</i> , 2011)	3L57 (Nash <i>et al.</i> , 2010)
	MOB _Q	MobA (RSF1010)	Y25	yes	In vitro SC and SS DNA cleavage (Scherzinger <i>et al.</i> , 1992)	2NS6 (Monzingo <i>et al.</i> , 2007)
	MOB _P	Tral (RP4)	Y22	Yes	Three motifs (Pansegrau <i>et al.</i> , 1994)	NO
		MbeA (ColE1)	Y19	HEN	Divergent HUH motif (Varsaki <i>et al.</i> , 2003)	NO
	MOB _V	MobM (pMV158)	?	yes	In vitro relaxase activity (Guzmán and Espinosa, 1997)	NO
	MOB _C	MobC (CloDF13)	?	No	No covalent complex (Núñez and De La Cruz, 2001)	NO
MOB _H	Tral (Neisseria chromosome)	Y93 Y201	HHH	Covalent intermediate (Salgado-Pabón <i>et al.</i> , 2007)	NO	

Tpsases	TnpA	IS608 (<i>H. pylori</i>)	Y127	HUH	Covalent intermediate (Ton-Hoang <i>et al.</i> , 2005)	2A6M, 2A6O (Ronning <i>et al.</i> , 2005) dimer 2VIH (TnpA/LE26), 2VIC (TnpA/LE26+Mn ²⁺), 2VHG (TnpA/RE31), 2VJU (TnpA/RE35) and 2VJV (TnpA/LE26/D6) (Barabas <i>et al.</i> , 2008) dimer
	TnpA	ISDra2 (<i>D. radiodurans</i>)	Y132	HUH	Covalent intermediate (Hickman <i>et al.</i> , 2010)	2XM3 (TnpA/LE27/T5), 2XMA (TnpA/RE34), 2XO6 (TnpA/Cd) and 2XQC (TnpA/Zn) (Hickman <i>et al.</i> , 2010) dimer
		IS1476 (<i>S. solfataricus</i>) Incomplete IS	?	HUH		ISfinder <i>S. solfataricus</i> genome 2F4F, 2F5G (Lee <i>et al.</i> , 2006) dimer
REP	TnpA _{REP}	E.coli		HUH	Covalent intermediate (Ton Hoang <i>et al.</i> , 2012)	(Messing <i>et al.</i> , 2012) monomer
	Tpase	IS91		HUH		NO
	Tpase	ISCR		HUH		NO

REFERENCES

- Asano, S., Higashitani, A., and Horiuchi, K. (1999). Filamentous phage replication initiator protein gpII forms a covalent complex with the 5' end of the nick it introduced. *Nucl. Acids Res.* 27, 1882–1889.
- Barabas, O., Ronning, D.R., Guynet, C., Hickman, A.B, Ton-Hoang, B., Chandler, M. and Dyda, F. (2008). Mechanism of IS200/IS605 family DNA transposases: activation and transposon-directed target site selection. *Cell* 132, 208–220.
- Boer, D.R., Ruíz-Masó, J.A., López-Blanco, J.R., Blanco, A.G., Vives-Llàcer, M., Chacón, P., Usón, I., Gomis-Rüth, F.X., Espinosa, M., Llorca, O., *et al.* (2009). Plasmid replication initiator RepB forms a hexamer reminiscent of ring helicases and has mobile nuclease domains. *EMBO J.* 28, 1666–1678.

- Campos-Olivas, R., Louis, J.M., Clerot, D., Gronenborn, B., and Gronenborn, A.M. (2002). The structure of a replication initiator unites diverse aspects of nucleic acid metabolism. *Proc. Natl. Acad. Sci. U.S.A.* 99, 10310–10315.
- Datta, S., Larkin, C., and Schildbach, J.F. (2003). Structural insights into single-stranded DNA binding and cleavage by F factor Tral. *Structure* 11, 1369–1379.
- Draper, O., César, C.E., Machón, C., De la Cruz, F., and Llosa, M. (2005). Site-specific recombinase and integrase activities of a conjugative relaxase in recipient cells. *Proc. Natl. Acad. Sci. U.S.A.* 102, 16385–16390.
- Garcillán-Barcia, M.P., Jurado, P., González-Pérez, B., Moncalián, G., Fernández, L.A., and De la Cruz, F. (2007). Conjugative transfer can be inhibited by blocking relaxase activity within recipient cells with intrabodies. *Mol. Microbiol.* 63, 404–416.
- Gonzalez-Perez, B., Lucas, M., Cooke, L.A., Vyle, J.S., De la Cruz, F., and Moncalián, G. (2007). Analysis of DNA processing reactions in bacterial conjugation by using suicide oligonucleotides. *EMBO J.* 26, 3847–3857.
- Grandoso, G., Avila, P., Cayón, A., Hernando, M.A., Llosa, M., and De la Cruz, F. (2000). Two active-site tyrosyl residues of protein TrwC act sequentially at the origin of transfer during plasmid R388 conjugation. *J. Mol. Biol.* 295, 1163–1172.
- Guasch, A., Lucas, M., Moncalián, G., Cabezas, M., Pérez-Luque, R., Gomis-Rüth, F.X., De la Cruz, F., and Coll, M. (2003). Recognition and processing of the origin of transfer DNA by conjugative relaxase TrwC. *Nat. Struct. Biol.* 10, 1002–1010.
- Guzmán, L.M., and Espinosa, M. (1997). The mobilization protein, MobM, of the streptococcal plasmid pMV158 specifically cleaves supercoiled DNA at the plasmid oriT. *J. Mol. Biol.* 266, 688–702.
- Hanai, R., and Wang, J.C. (1993). The mechanism of sequence-specific DNA cleavage and strand transfer by phi X174 gene A* protein. *J. Biol. Chem.* 268, 23830–23836.
- Heyraud-Nitschke, F., Schumacher, S., Laufs, J., Schaefer, S., Schell, J., and Gronenborn, B. (1995). Determination of the origin cleavage and joining domain of geminivirus Rep proteins. *Nucleic Acids Res.* 23, 910–916.
- Hickman, A.B., Ronning, D.R., Kotin, R.M., and Dyda, F. (2002). Structural unity among viral origin binding proteins: crystal structure of the nuclease domain of adeno-associated virus Rep. *Mol. Cell* 10, 327–337.
- Hickman, A.B., Ronning, D.R., Perez, Z.N., Kotin, R.M., and Dyda, F. (2004). The nuclease domain of adeno-associated virus rep coordinates replication initiation using two distinct DNA recognition interfaces. *Mol. Cell* 13, 403–414.
- Im, D.S., and Muzyczka, N. (1990). The AAV origin binding protein Rep68 is an ATP-dependent site-specific endonuclease with DNA helicase activity. *Cell* 61, 447–457.
- Larkin, C., Datta, S., Harley, M.J., Anderson, B.J., Ebie, A., Hargreaves, V., and Schildbach, J.F. (2005). Inter- and Intramolecular Determinants of the Specificity of Single-Stranded DNA Binding and Cleavage by the F Factor Relaxase. *Structure* 13, 1533–1544.
- Lee, H.H., Yoon, J.Y., Kim, H.S., Kang, J.Y., Kim, K.H., Kim, D.J., Ha, J.Y., Mikami, B., Yoon, H.J. and Suh, S.W. Crystal Structure of a Metal Ion-bound IS200 Transposase. *J. Biol. Chem.* 281, 4261–4266.
- Van Mansfeld, A.D., Van Teeffelen, H.A., Baas, P.D., and Jansz, H.S. (1986). Two juxtaposed tyrosyl-OH groups participate in phi X174 gene A protein catalysed cleavage and ligation of DNA. *Nucleic Acids Res.* 14, 4229–4238.
- Monzingo, A.F., Ozburn, A., Xia, S., Meyer, R.J., and Robertus, J.D. (2007). The Structure of the Minimal Relaxase Domain of MobA at 2.1 Å Resolution. *J Mol Biol* 366, 165–178.
- Moscoso, M., Eritja, R., and Espinosa, M. (1997). Initiation of replication of plasmid pMV158: mechanisms of DNA strand-transfer reactions mediated by the initiator RepB protein. *J. Mol. Biol.* 268, 840–856.

- Nash, R.P., Habibi, S., Cheng, Y., Lujan, S.A., and Redinbo, M.R. (2010). The mechanism and control of DNA transfer by the conjugative relaxase of resistance plasmid pCU1. *Nucleic Acids Res.* 38, 5929–5943.
- Nash, R.P., Niblock, F.C., and Redinbo, M.R. (2011). Tyrosine partners coordinate DNA nicking by the *Salmonella typhimurium* plasmid pCU1 relaxase enzyme. *FEBS Lett.* 585, 1216–1222.
- Noirot-Gros, M.F., Bidnenko, V., and Ehrlich, S.D. (1994). Active site of the replication protein of the rolling circle plasmid pC194. *EMBO J.* 13, 4412–4420.
- Noirot-Gros, M.F., and Ehrlich, S.D. (1996). Change of a catalytic reaction carried out by a DNA replication protein. *Science* 274, 777–780.
- Núñez, B., and De La Cruz, F. (2001). Two atypical mobilization proteins are involved in plasmid CloDF13 relaxation. *Molecular Microbiology* 39, 1088–1099.
- Odegrip, R., and Haggård-Ljungquist, E. (2001). The two active-site tyrosine residues of the α protein play non-equivalent roles during initiation of rolling circle replication of bacteriophage ϕ 2. *J. Mol. Biol.* 308, 147–163.
- Pansegrau, W., Schröder, W., and Lanka, E. (1994). Concerted action of three distinct domains in the DNA cleaving-joining reaction catalyzed by relaxase (Tral) of conjugative plasmid RP4. *J. Biol. Chem.* 269, 2782–2789.
- Rasooly, A., and Novick, R.P. (1993). Replication-specific inactivation of the pT181 plasmid initiator protein. *Science* 262, 1048–1050.
- Salgado-Pabón, W., Jain, S., Turner, N., Van der Does, C., and Dillard, J.P. (2007). A novel relaxase homologue is involved in chromosomal DNA processing for type IV secretion in *Neisseria gonorrhoeae*. *Mol Microbiol* 66, 930–947.
- Scherzinger, E., Lurz, R., Otto, S., and Dobrinski, B. (1992). In vitro cleavage of double- and single-stranded DNA by plasmid RSF1010-encoded mobilization proteins. *Nucleic Acids Res.* 20, 41–48.
- Street, L.M., Harley, M.J., Stern, J.C., Larkin, C., Williams, S.L., Miller, D.L., Dohm, J.A., Rodgers, M.E., and Schildbach, J.F. (2003). Subdomain organization and catalytic residues of the F factor Tral relaxase domain. *Biochimica Et Biophysica Acta (BBA) - Proteins and Proteomics* 1646, 86–99.
- Varsaki, A., Lucas, M., Afendra, A.S., Drainas, C., and De la Cruz, F. (2003). Genetic and biochemical characterization of MbeA, the relaxase involved in plasmid ColE1 conjugative mobilization. *Mol. Microbiol.* 48, 481–493.
- Vega-Rocha, S., Byeon, I.-J.L., Gronenborn, B., Gronenborn, A.M., and Campos-Olivas, R. (2007a). Solution Structure, Divalent Metal and DNA Binding of the Endonuclease Domain from the Replication Initiation Protein from Porcine Circovirus 2. *Journal of Molecular Biology* 367, 473–487.
- Vega-Rocha, S., Gronenborn, B., Gronenborn, A.M., and Campos-Olivas, R. (2007b). Solution Structure of the Endonuclease Domain from the Master Replication Initiator Protein of the Nanovirus Faba Bean Necrotic Yellows Virus and Comparison with the corresponding Geminivirus and Circovirus Structures. *Biochemistry* 46, 6201–6212.