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NCOMMS-19-07159B Corresponding author(s): Dr. Glenn Merlino

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Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see<u>Authors & Referees</u> and the<u>Editorial Policy Checklist</u>.

Statistics

For	all st	atistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Cor	firmed
	×	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	×	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	×	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
	×	A description of all covariates tested
	×	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	×	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	×	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted Give <i>P</i> values as exact values whenever suitable.
	×	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
	×	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
	×	Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated
		Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

Software and code

Policy information about <u>availability of computer code</u>			
Data collection	See page 43		
Data analysis	See methods and page 43		

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a <u>data availability statement</u>. This statement should provide the following information, where applicable: - Accession codes, unique identifiers, or web links for publicly available datasets

- A list of figures that have associated raw data
- A description of any restrictions on data availability

See pages 42-43

× Life sciences

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	See page 24
Data exclusions	See pages 23-24: Of the TCGA data patient samples were collected between 0-14 days after disease classification only to ensure accurate staging alongside gene expression
Replication	All experiments were reproducible or validated in multiple data sources.
Randomization	See page 24
Blinding	See page 24

Reporting for specific materials, systems and methods

Methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involved in the study	n/a	Involved in the study
	X Antibodies	×	ChIP-seq
	x Eukaryotic cell lines		Flow cytometry
×	Palaeontology	x	MRI-based neuroimaging
	× Animals and other organisms		
×	Human research participants		
×	Clinical data		

Antibodies

Antibodies used	See methods 16-43
Validation	Antibody registry code provided. Antibodies pre-published and have been validated by the manufacturers. KDELR3 antibody was further validated by Immunofluorescence confirmation of co-localization with DDK-tagged KDELR3

Eukaryotic cell lines

Policy information about <u>cell lines</u>	
Cell line source(s)	See methods 16-43
Authentication	See methods 24-27
Mycoplasma contamination	See methods 24-25
Commonly misidentified lines (See <u>ICLAC</u> register)	See methods 24-25 - No commonly identified lines used in this study

Animals and other organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research

Laboratory animals	See methods 16-43
Wild animals	N/A
Field-collected samples	N/A
Ethics oversight	see methods page 24

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Flow Cytometry

Plots

Confirm that:

X The axis labels state the marker and fluorochrome used (e.g. CD4-FITC).

The axis scales are clearly visible. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers).

X All plots are contour plots with outliers or pseudocolor plots.

🗶 A numerical value for number of cells or percentage (with statistics) is provided.

Methodology

Sample preparation	See methods 16-43
Instrument	See methods 16-43
Software	see methods pages 32-33
Cell population abundance	See methods 16-43
Gating strategy	See methods 16-43

X Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.