## 1 Supplementary information

## 2 Type I interferon shapes brain distribution and tropism of tick-borne flavivirus

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45 Supplementary Fig. 1. IFN-I response influences host survival and distribution of LGTV infection. a Survival analysis of WT and *Ifnar<sup>-/-</sup>* mice, intracranially injected with 1000 pfu of LGTV (n = 5). Survival 46 47 differences between groups were significant (log-rank test, p = 0.0031). b OPT cross sections of a representative Ifnar<sup>/-</sup> brain showing antibody penetration in the deeper areas of the brain (arrows). c OPT-48 49 scanned immunolabeled brain reveals the distribution of LGTV infection in the adult mouse brain. 50 Volumetric 3D render of supplementary OPT scans of the brain from mock and LGTV infected mice 51 immunolabeled with antibodies against viral NS5. The signal intensity was normalized within an individual 52 brain and adjusted to identical minimum and maximum. The viral signal was overlaid onto the anatomical 53 outlines created from iso-surface rendering of the autofluorescence signal of each brain. For each image 54 pair, the top and lateral views of the same specimen are shown. Scale bar = 2000 µm. Source data are 55 provided as a Source Data file.

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50 Supplementary Fig. 2. Image and quantification of viral infection in cerebral cortex regions. 61 Quantification and statistical analysis (two-tailed unpaired t-test (a and c) with Holm-Sidak multiple 62 comparison correction (c and d) of viral OPT signal in **a** temporal association area (\*p=0.047), **b** individual 63 VOIs in the composite entorhinal cortical region, **c** composite hippocampal region, and **d** individual VOIs in 64 the hippocampal region. Source data are provided as a Source Data file.

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70 Supplementary fig. 3 Microglia staining of OB and primary microglia infected with LGTV in vitro.

a) Maximum-intensity projection of confocal z-stack. The images were taken from sagittal brain sections

72 (10  $\mu$ m) using confocal microscope, representative picture of OB is shown. Scale bars = 100  $\mu$ m. The

radiation sections were immunolabeled using anti-NS5 (green), anti-Iba1 (red) for microglia, and DAPI for nucleus

74 (blue). b) Primary microglia were isolated from WT and *Ifnar<sup>-/-</sup>* brains. The cells were infected with LGTV

75 (MOI 1) for 72 h and viral growth was measured by focus forming assay, bars show mean  $\pm$  SD (n = 6).

76 Source data are provided as a Source Data file.

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Supplementary Fig. 4. Transcriptional differences and viral load in WT and Ifnar-/- mice. a Gene 84 expression of Cd8, Cd4 and Cd19 in cortex of infected WT (n=5) and Ifnar<sup>-/-</sup> (n=5) mice. Quantified by 85 86 gPCR and normalized to housekeeping gene Gapdh, bars show mean ±SD. b Number of DEGs (log2FC>1, 87 padj < 0.05) between *lfnar*<sup>-/-</sup> and WT mice in uninfected samples. **c** Viral load in cerebral cortex at the 88 humane endpoint in WT (n=5) and Ifnar- mice (n=5) infected with 10,000 PFU intracranially. Quantified by 89 qPCR and normalized to housekeeping gene Gapdh. Bars represent mean ±SEM. d Gene expression of 90 Cd8, Cd4 and Cd19 in cortex of infected WT (n=5) and Ifnar-(n=5) mice, bars show mean  $\pm$ SEM. Quantified by gPCR and normalized to housekeeping gene Gapdh. e Heatmap showing expression of T 91 cell chemoattractants and corresponding receptors upon infection in WT and Ifnar-(If) mice. Source data 92 93 are provided as a Source Data file.

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Supplementary Fig. 5. Infiltration of immune cells in WT and Ifnar-- mice. a Representative scatterplots showing gating strategy based on forward scatter (FSC) and side scatter (SSC) for obtaining single cells used in downstream analysis of brain leucocytes. b Flow cytometry of brain leukocytes from the cerebral cortex of untreated and infected WT mice. Representative scatterplots and percentage CD8+ T cells and NK cells (CD335<sup>+</sup>). Boxes represent means of untreated (n=3) and LGTV infected (n=2). c Subclustering of 2,363 nuclei of micro/MØ cell subset belonging to all datasets, colored by gene signatures for microglia (Cx3cr1, P2ry12, Slco2b1, Tmem119) or MØ (Slfn4, Ms4a8a, Clec4e, Itga4) (PMID: 31325960). Source data are provided as a Source Data file. 

## 110 Supplementary Table 1. Key Antibody Table

Antibodies	Origin	Clonality	Clone name	Working dilution	Use in the current study	Company (Cat#)
Anti-chicken Alexa Fluor 488	Goat	Polyclonal	-	1:400	IHC	Thermo Fisher Scientific (A11039)
Anti-chicken Alexa Fluor 555	Goat	Polyclonal	-	1:500	IHC	Thermo Fisher Scientific (A21437)
Anti-chicken Alexa Fluor 680	Goat	Polyclonal	-	1:500	OPT, IHC	Abcam (ab175779)
Anti-mouse Alexa Fluor 647	Goat	Polyclonal	-	1:500	IHC	Abcam (ab150115)
Anti-rabbit Alexa Fluor 488	Donkey	Polyclonal	-	1:500	IHC	Thermo Fisher Scientific (A21206)
Anti-rabbit Alexa Fluor 594 (pre-absorbed)	Donkey	Polyclonal	-	1:500	OPT	Abcam (ab150064)
Anti-mouse CD45 Alexa 488	Rat	Monoclonal	30F-11	1:100	IHC	Thermo Fisher Scientific (53-0451- 82)
AQPI	Mouse	Monoclonal	sc-32737	1:50	IHC	Santa Cruz Biotechnology (sc- 32737)
CD8	Rat	Monoclonal	Clone 53-6.7 (RUO)	1:200	IHC	BD Pharmigen (561093)
CD335	Rat	Monoclonal	29A1.4	1:500	IHC	Thermo Fisher Scientific (AB_1724164
Doublecortin	Rabbit	Polyclonal	-	1:800	IHC	Cell Signaling (4604)
lba1	Rabbit	Polyclonal	-	1:400	IHC	Histolab (CP290)
NS5*	Chicken	Polyclonal	-	1:1000	OPT, IHC	Agrisera AB
TMEM119	Rabbit	Monoclonal	28-3	0.5µg/ml	IHC	Abcam (ab209064)
TUBB3	Rabbit	Polyclonal	-	1:3000	IHC	BioLegend (PRB-435P)

111 \*Affinity-purified NS5 antibody was produced in chicken, according to the manufacturer's protocol, using the following peptide sequence of NS5 from

112 tick-borne encephalitis virus strain Torö (GenBank: DQ401140): (carboxylated)-CMDRHDLHWELRLESS-(amidated).