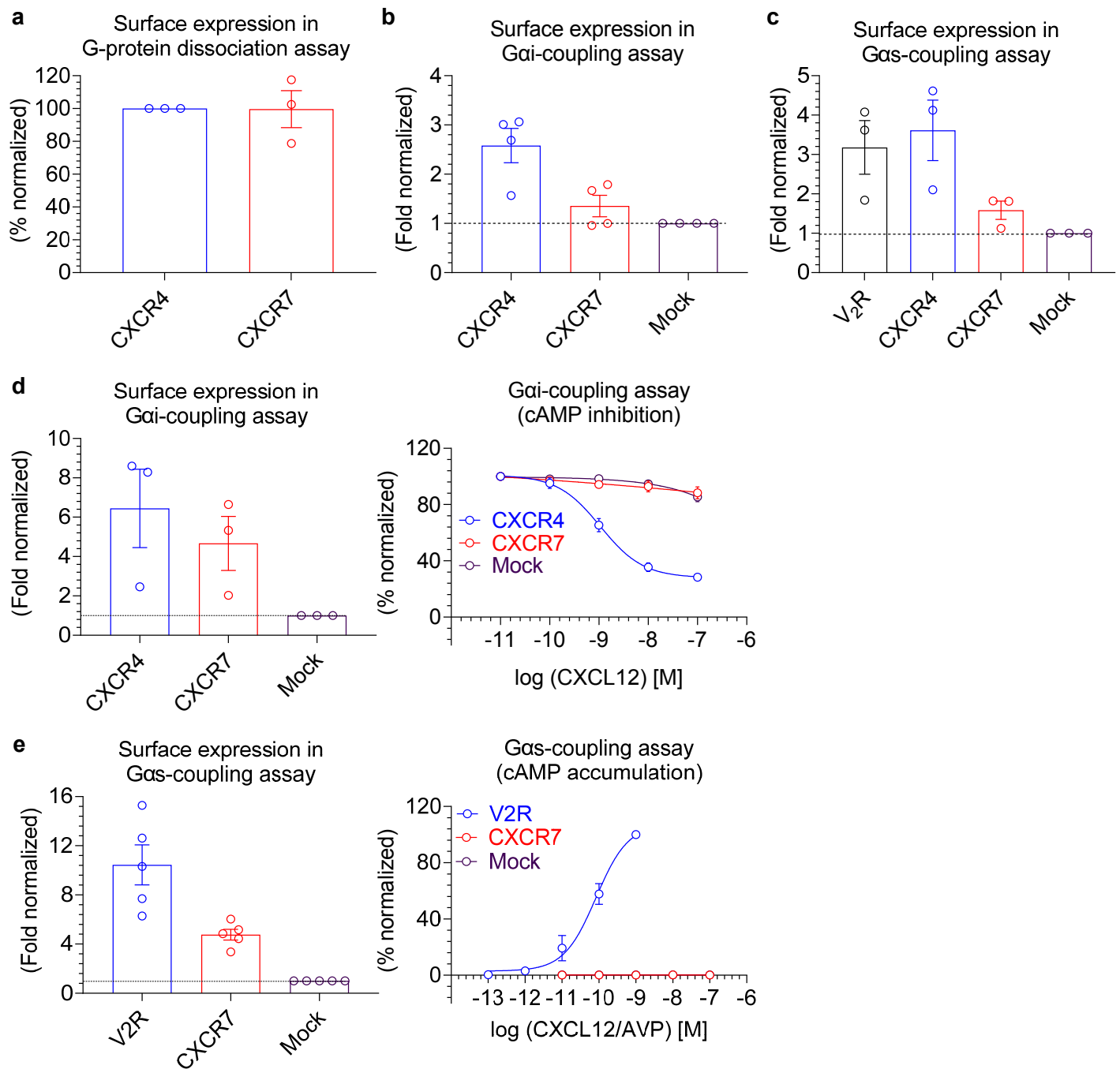


Supplementary Information

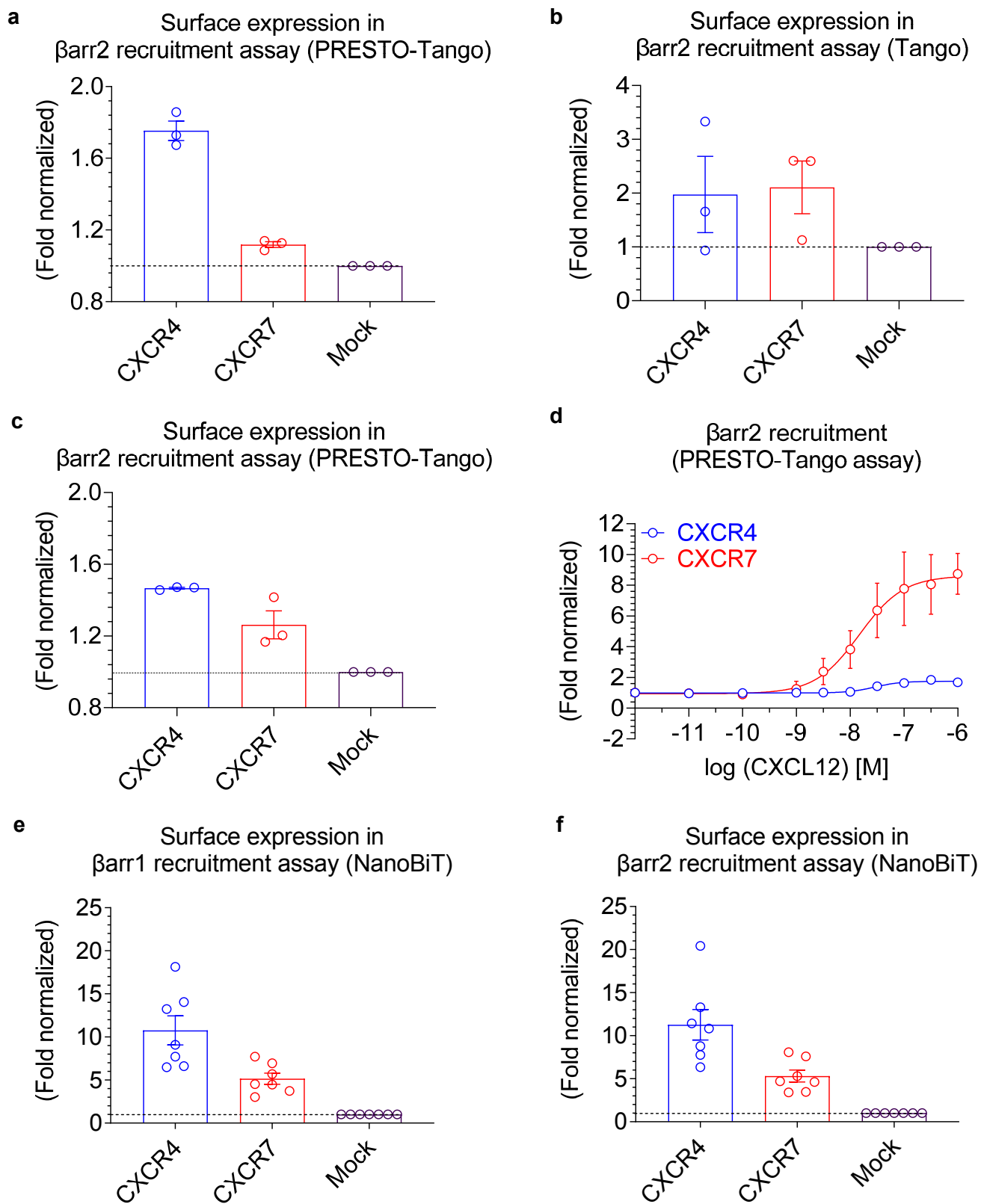
Molecular insights into intrinsic transducer-coupling bias in the CXCR4-CXCR7 system

Parishmita Sarma¹, Carlo Marion C. Carino², Deeksha S¹, Shubhi Pandey¹, Hemlata Dwivedi-Agnihotri¹, Xue Rui³, Yubo Cao⁴, Kouki Kawakami², Poonam Kumari⁵, Yu-Chih Chen⁶, Kathryn E. Luker⁷, Prem N. Yadav⁵, Gary D. Luker^{7,8}, Stéphane A. Laporte^{5,9}, Xin Chen³, Asuka Inoue², and Arun K. Shukla^{1*}

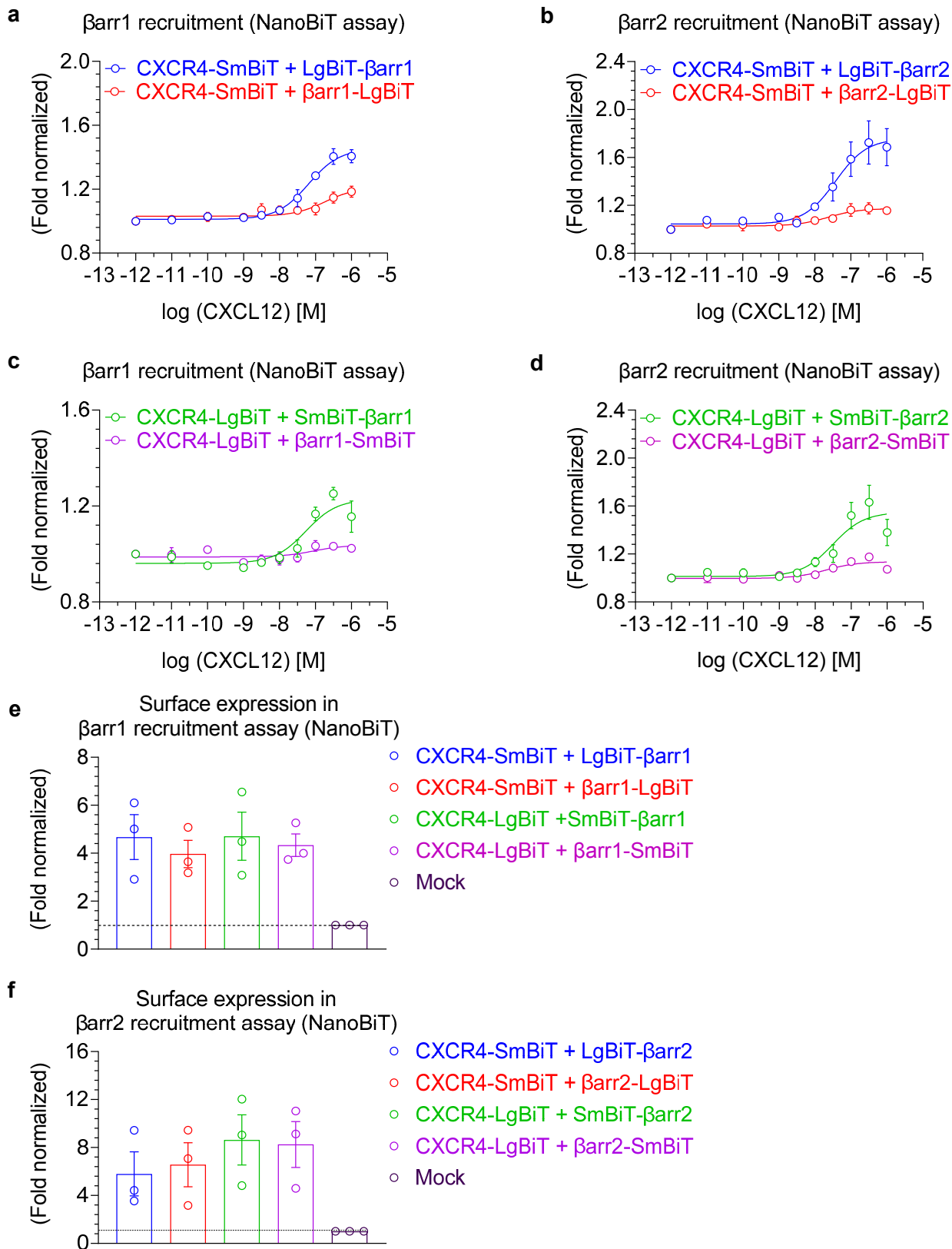
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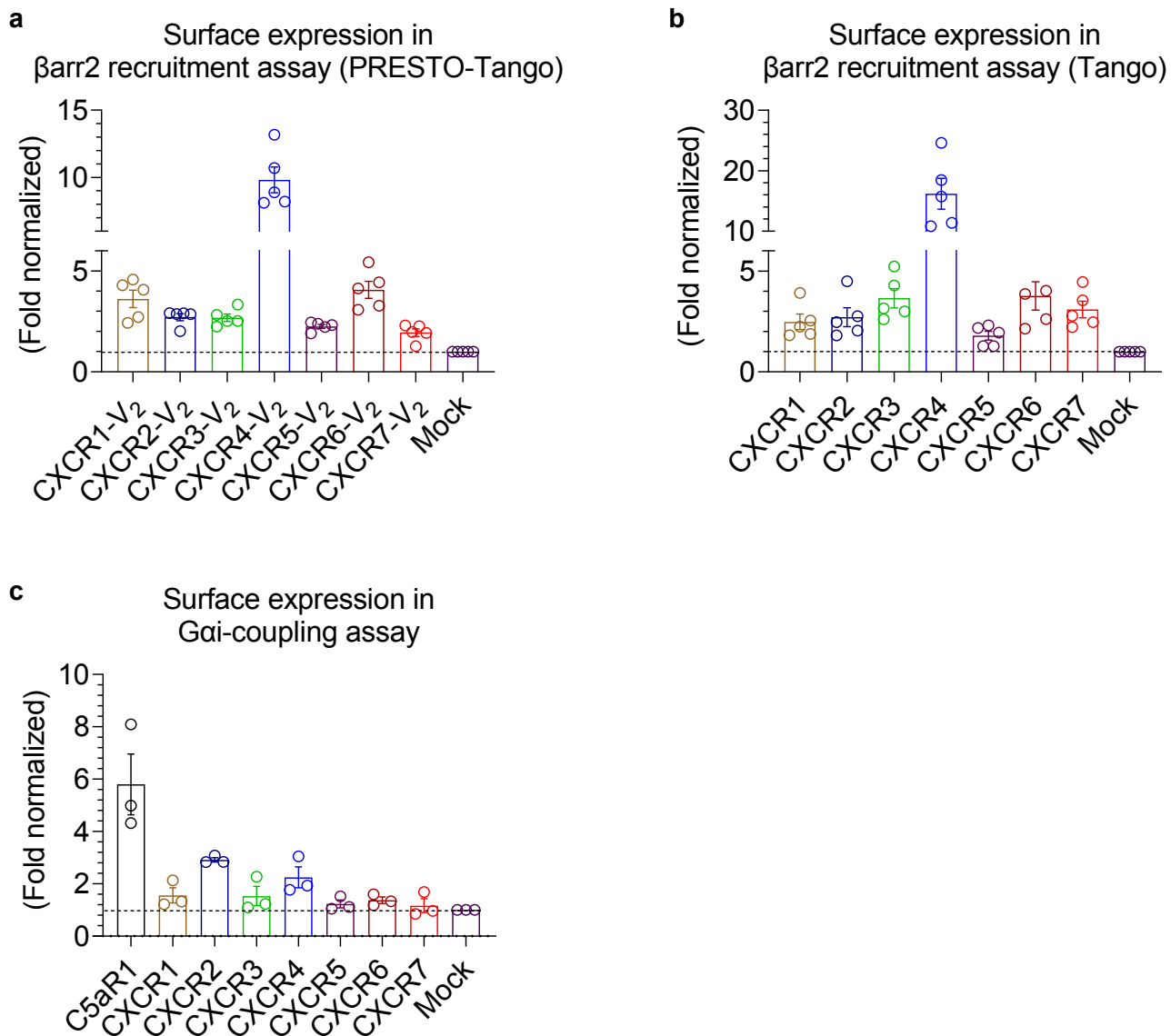
Supplementary Fig. 1. Surface expression of CXCR4 and CXCR7 in G-protein assays. **a** Surface expression of CXCR4 and CXCR7 in the G-protein dissociation assay measured using flow cytometry assay (mean±SEM; n=3 independent experiments; normalized with CXCR4 as 100%). **b-c** Surface expression of the indicated receptors in cAMP assay as measured using whole cell ELISA (mean±SEM; n=4 independent experiments for panel **b** and n=3 independent experiments for panel **c**; normalized as fold over mock (pcDNA)-transfection). **d-e** Surface expression (whole cell ELISA) and second messenger response (Gai/s-coupling assay) under higher expression levels of CXCR4 and CXCR7 (mean±SEM; n=3 independent experiments; for panel **d** and n=5-6 independent experiments for panel **e**; for receptor surface expression: n=5 independent experiments, for Gas-coupling assay; n=6) (Gai/s-coupling assay in panel **d** and **e** are normalized with minimal ligand dose for CXCR4 and maximal signal for V₂R treated as 100%, respectively; surface expression data are normalized as fold over mock (pcDNA)-transfection). Dotted line in the plots **b**, **c**, **d**, and **e** indicates 1 fold. Source data are provided as a source data file.



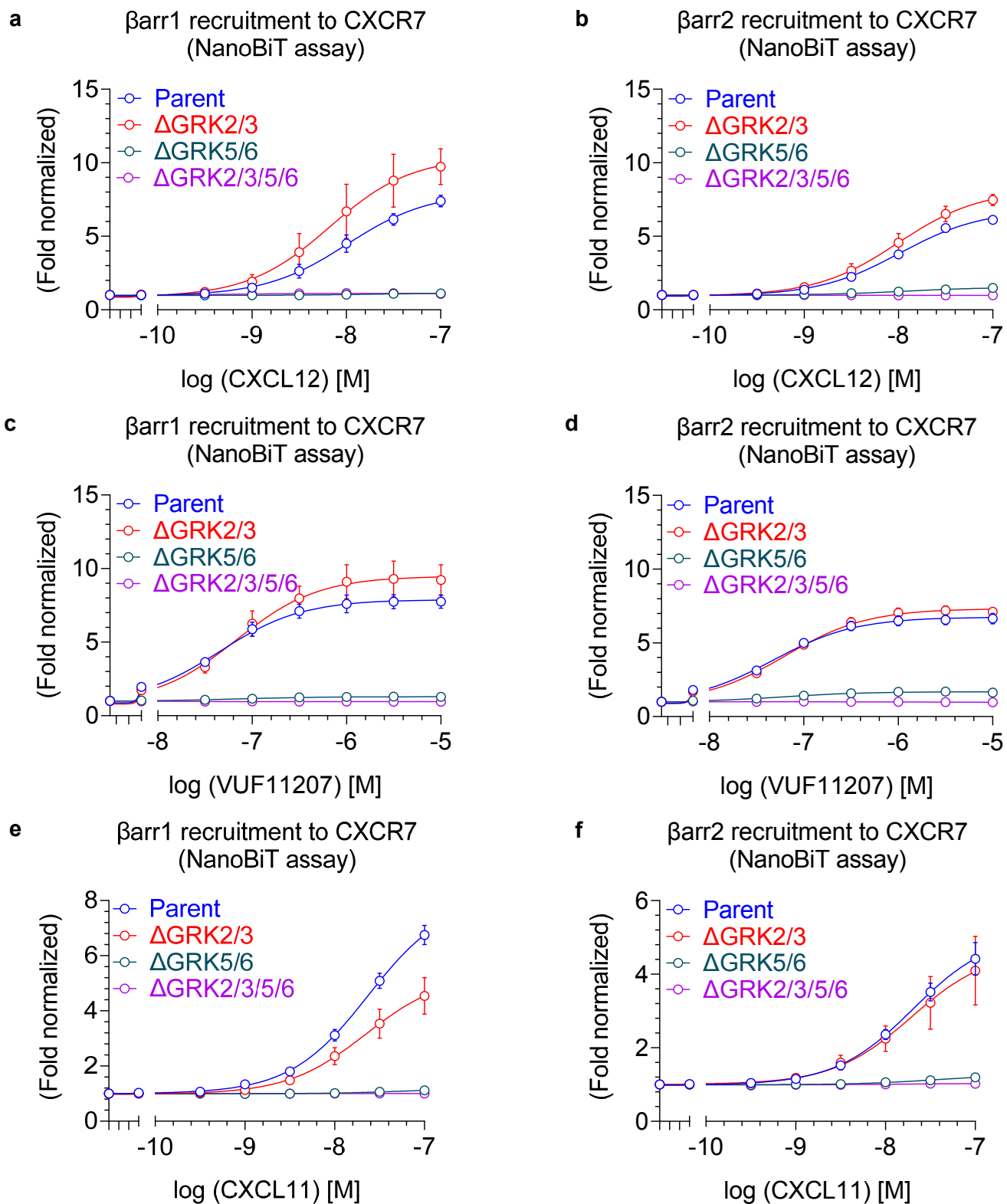
Supplementary Fig. 2. Surface expression of CXCR4 and CXCR7 in β arr recruitment assays. a-c, e-f Surface expression of CXCR4 and CXCR7 in β arr recruitment assays measured using whole cell ELISA (mean \pm SEM; n=3 independent experiments for panel a-b; n=7 independent experiments for panel e-f; n=3 independent experiments for panel c; normalized as fold over mock (pcDNA)-transfection). d CXCL12-induced β arr2 recruitment to CXCR4 and CXCR7 in PRESTO-Tango assay (mean \pm SEM; n=3 independent experiments; normalized with the luminescence signal at minimal ligand dose treated as 1). Dotted line in the plots a, b, c, e, and f indicates 1 fold. Source data are provided as a source data file.



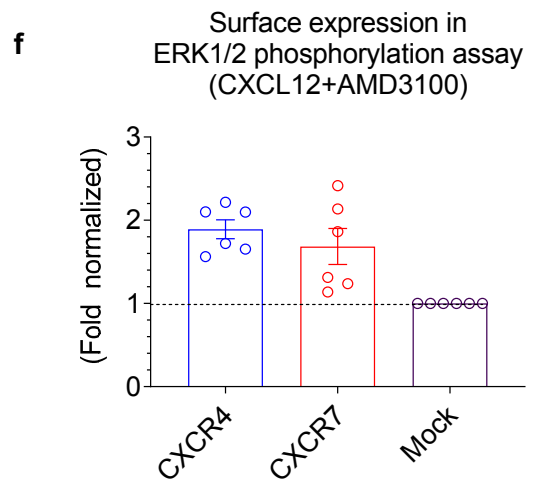
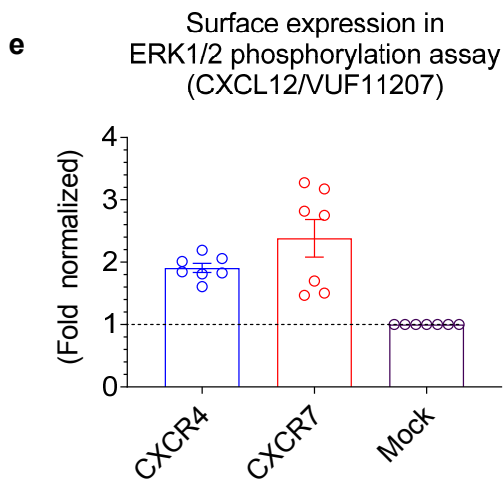
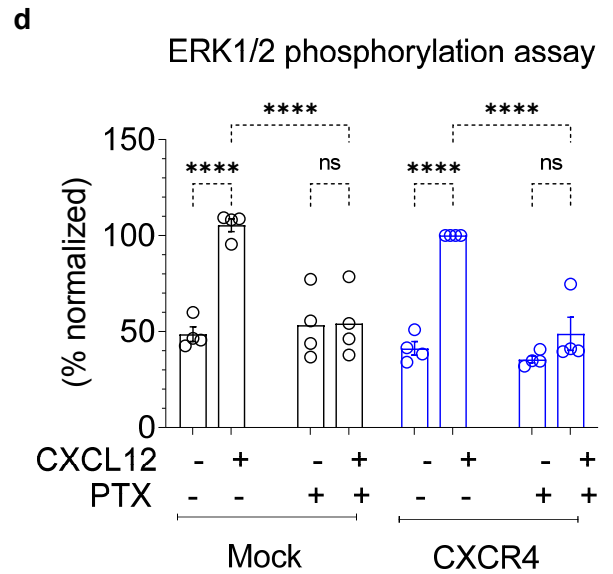
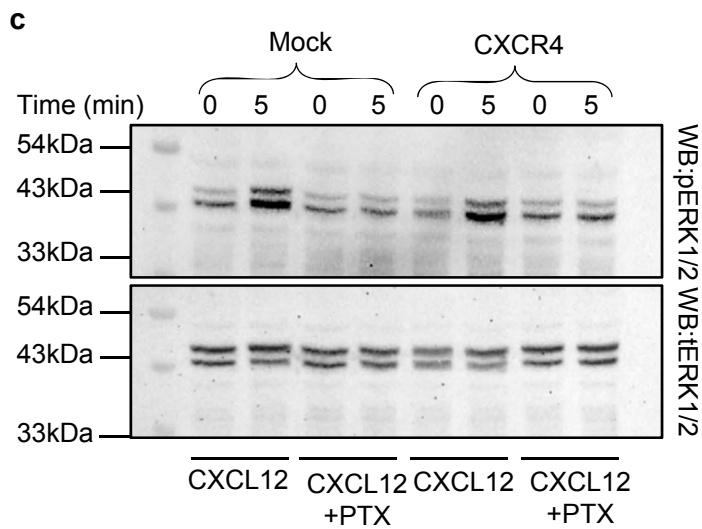
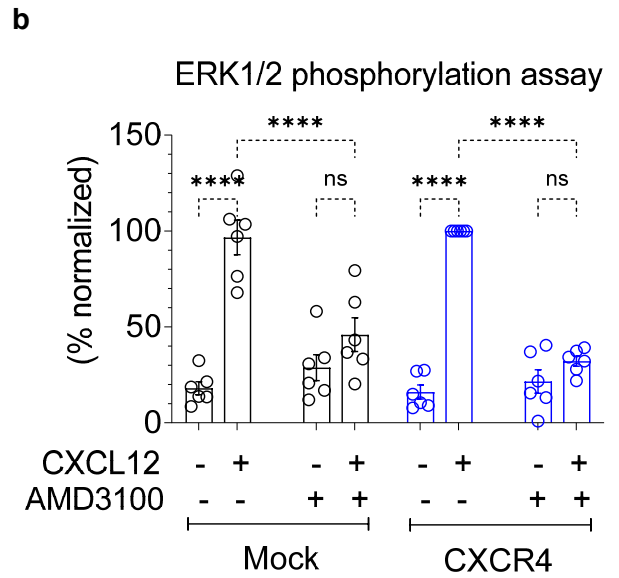
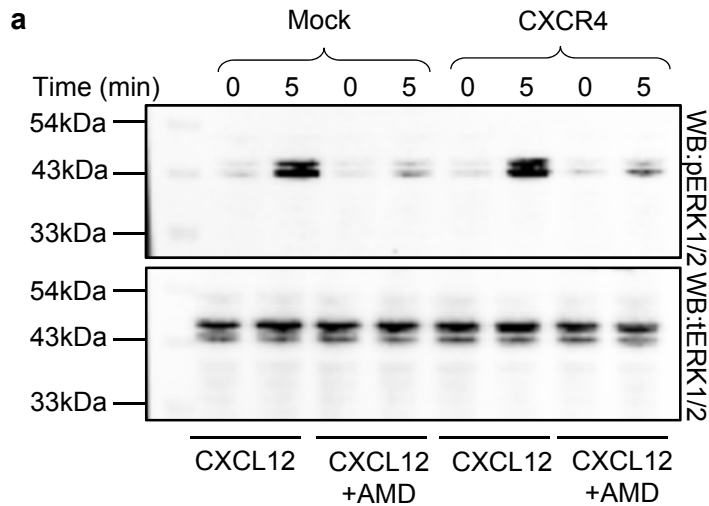
Supplementary Fig. 3. Optimization of NanoBiT-based β arr recruitment assay for CXCR4. a-d CXCL12-induced β arr1/2 recruitment to CXCR4 in NanoBiT assay with indicated combination of the constructs (mean \pm SEM; n=3 independent experiments; normalized with the luminescence signal at minimal ligand dose treated as 1). e-f Surface expression of CXCR4 constructs under indicated transfection conditions in the β arr recruitment assay as measured using whole cell ELISA (mean \pm SEM; n=3 independent experiments; normalized as fold over mock (pcDNA)-transfection). Dotted line in the plots e and f indicates 1 fold. Source data are provided as a source data file.



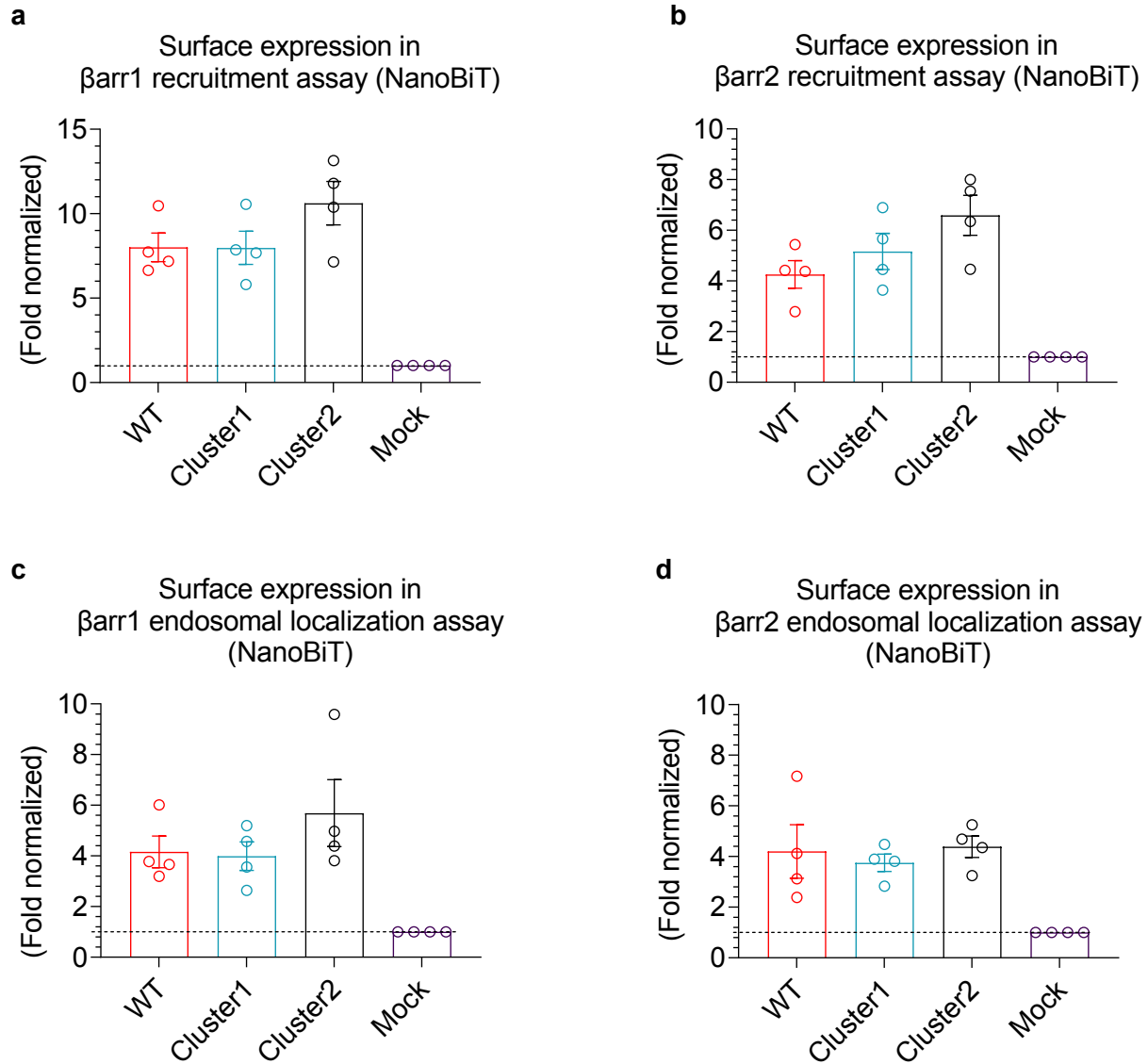
Supplementary Fig. 4. Surface expression of CXCRs. **a-b** Surface expression of indicated receptors in β arr recruitment assays measured using whole cell ELISA (mean \pm SEM; n=5 independent experiments for panel a-b; normalized as fold over mock(pcDNA)-transfection). **c** Surface expression of the indicated receptors in cAMP assay as measured using whole cell ELISA (mean \pm SEM; n=3 independent experiments; normalized as fold over mock (pcDNA)-transfection). Dotted line in plots a, b, and c indicates 1 fold. Source data are provided as a source data file.



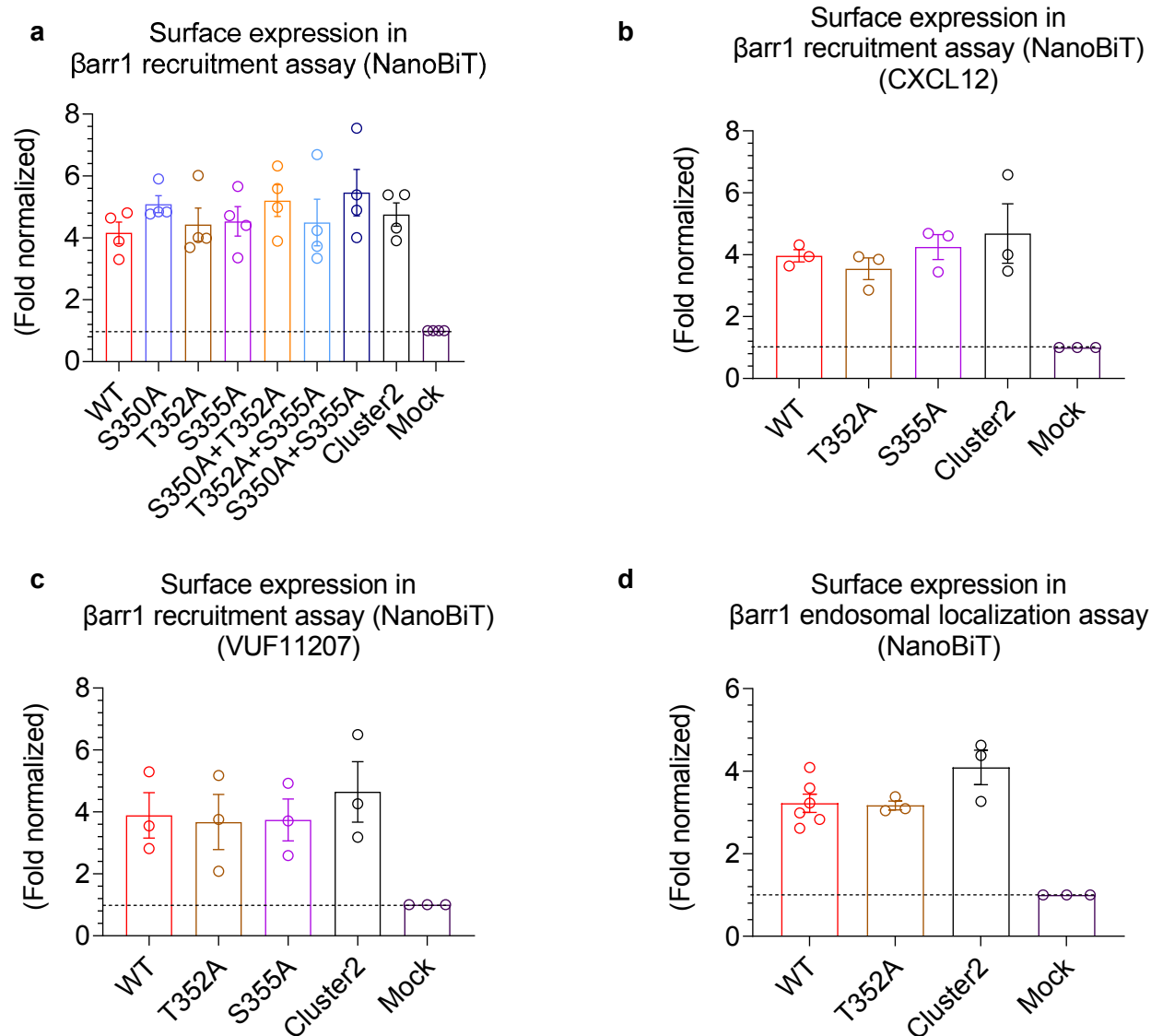
Supplementary Fig. 5. Effect of PTX treatment on β arr recruitment. a-f CXCL12-, VUF11207-, and CXCL11-induced β arr1 and 2 recruitment to CXCR7 in GRK knock-out cells using the NanoBiT assay with co-expression of the catalytic subunit of pertussis toxin (PTX) (Receptor-SmBiT+LgBiT- β arr1/2), respectively (mean \pm SEM; n=4-5 independent experiments; i.e., for CXCL12- and VUF11207-induced β arr1 recruitment: Parent, n=4; Δ GRK2/3, n=4; Δ GRK5/6, n=5; Δ GRK2/3/5/6, n=4; for CXCL12-induced β arr2 recruitment: Parent, n=5; Δ GRK2/3, n=4; Δ GRK5/6, n=4; Δ GRK2/3/5/6, n=4; for VUF11207-induced β arr2 recruitment: Parent, n=5; Δ GRK2/3, n=4; Δ GRK5/6, n=5; Δ GRK2/3/5/6, n=4; for CXCL11-induced β arr1 and β arr2 recruitment: Parent, Δ GRK2/3, Δ GRK5/6, Δ GRK2/3/5/6, n=4; normalized as fold response with respect to the unstimulated condition treated as 1). Source data are provided as a source data file.



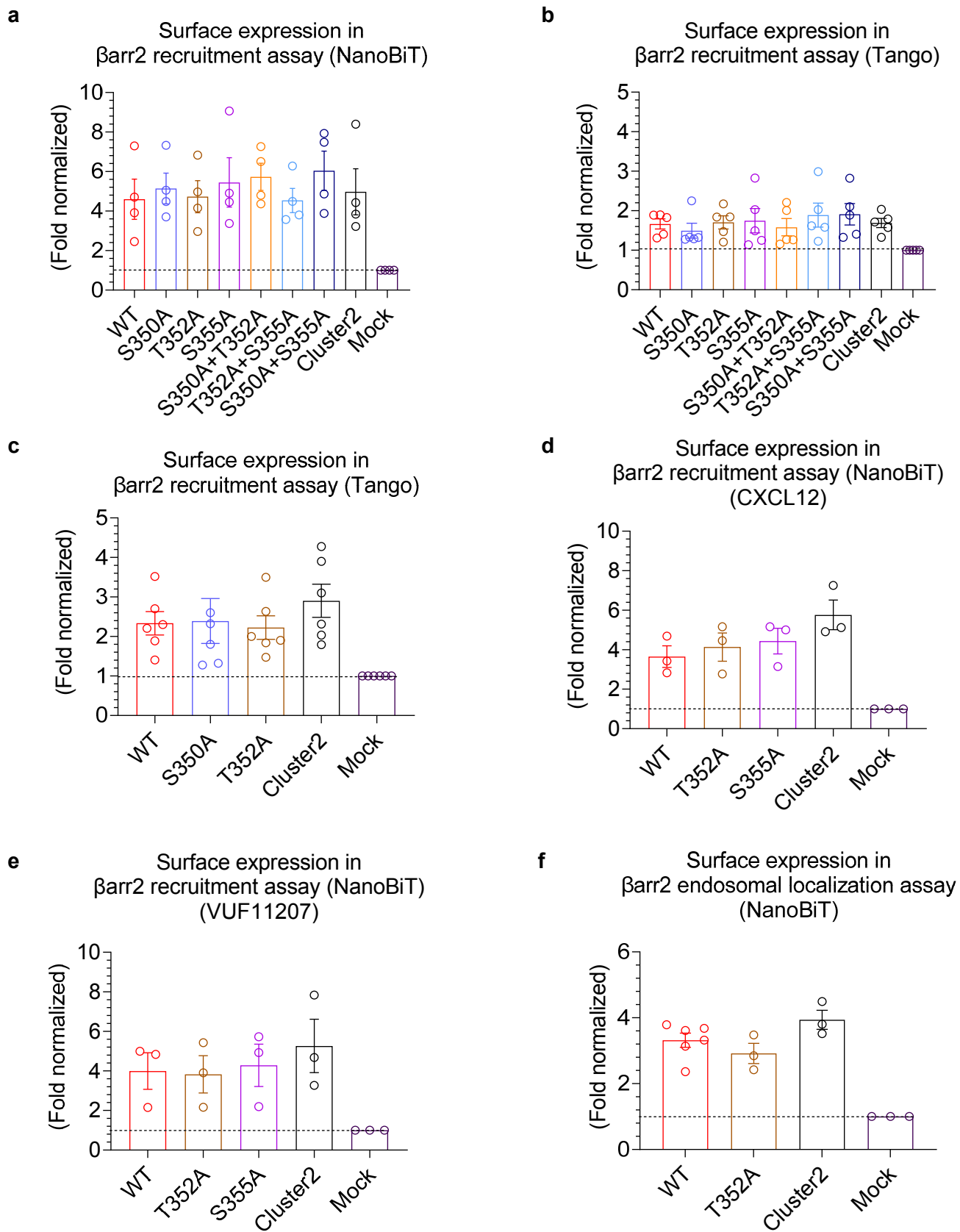
Supplementary Fig. 6. Effect of AMD3100 and PTX treatment on ERK1/2 phosphorylation. a-d CXCR4-induced ERK1/2 phosphorylation in HEK-239 cells transfected with either pcDNA (mock) or CXCR4 plasmid with and without pre-treatment of AMD3100 and PTX as indicated. Panels b and d present densitometry-based quantification of data presented in panels a and c (mean±SEM; n=6 and n=4 independent experiments, respectively; normalized with 5 min stimulation of CXCR4 condition without AMD3100/PTX; Two-way ANOVA, Tukey's multiple comparison test. The exact *p* values for panel b are as follows: mock-AMD3100: 0 min vs. mock-AMD3100: 5 min (*p* < 0.0001), mock-AMD3100: 5 min vs. mock+AMD3100: 5 min (*p* < 0.0001), mock+AMD3100: 0 min vs. mock+AMD3100: 5 min (*p* = 0.4441), CXCR4-AMD3100: 0 min vs. CXCR4-AMD3100: 5 min (*p* < 0.0001), CXCR4-AMD3100: 5 min vs. CXCR4+AMD3100: 5 min (*p* < 0.0001), CXCR4+AMD3100: 0 min vs. CXCR4+AMD3100: 5 min (*p* = 0.8975). The exact *p* values for panel d are as follows: mock-PTX: 0 min vs. mock-PTX: 5 min (*p* < 0.0001), mock-PTX: 5 min vs. mock+PTX: 5 min (*p* < 0.0001), mock+PTX: 0 min vs. mock+PTX: 5 min (*p* > 0.9999), CXCR4-PTX: 0 min vs. CXCR4-PTX: 5 min (*p* < 0.0001), CXCR4-PTX: 5 min vs. CXCR4+PTX: 5 min (*p* < 0.0001), CXCR4+PTX: 0 min vs. CXCR4+PTX: 5 min (*p* = 0.7397) (*****p* < 0.0001, ns= non-significant). **e-f** Surface expression of CXCR4 and CXCR7 in ERK1/2 phosphorylation assay was measured using whole cell ELISA (mean±SEM; n=7 independent experiments for panel e and n=6 independent experiments for panel f, normalized as fold-over mock (pcDNA)-transfection. Dotted line in the plots e and f indicates 1 fold. Source data are provided as a source data file.



Supplementary Fig. 7. Surface expression of CXCR7 mutants in different assays. a-e Surface expression of indicated receptors was measured in different assays using whole cell ELISA (mean \pm SEM; n=4 independent experiments; normalized as fold over mock (pcDNA)-transfection). Dotted line in the plots a, b, c, and d indicates 1 fold. Source data are provided as a source data file.



Supplementary Fig. 8. Surface expression of CXCR7 mutants in different assays. a-d Surface expression of indicated receptors were measured in different assays using whole cell ELISA (mean±SEM; n=4 independent experiments for panel a, n=3 independent experiments for panel b and c, n=3-6 independent experiments for panel d; for β arr1 endosomal localization: WT, n=6; T352A, n=3; and Cluster2, n=3 normalized as fold over mock (pcDNA)-transfection). Dotted line in the plots a, b, c, and d indicates 1 fold. Source data are provided as a source data file.

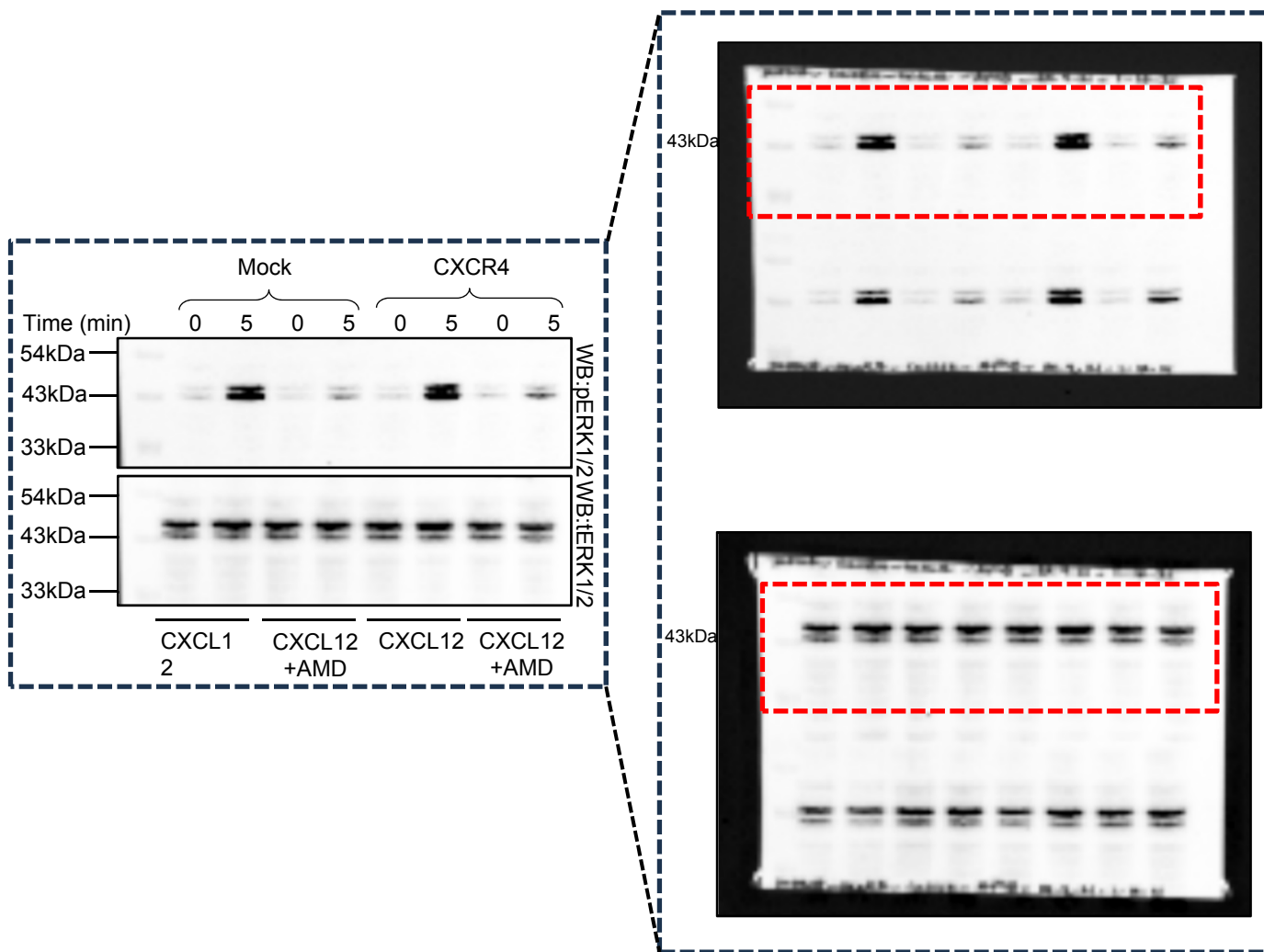


Supplementary Fig. 9. Surface expression of CXCR7 mutants in different assays. a-f Surface expression of indicated receptors were measured in different assays using whole cell ELISA (mean \pm SEM; n=4 independent experiments for panel a, n=5 for panel b, n=6 for panel c, n=3 for panel d and e, n=3-6 for panel f; β arr2 endosomal localization: WT, n=6; T352A, n=3; and Cluster2, n=3; normalized as fold over mock (pcDNA)-transfection). Dotted line in the plots a,b,c,d, and e indicates 1 fold. Source data are provided as a source data file.

Supplementary Table 1. List of the primers used in the study.

Construct	Primer	Sequence
CXCR1 Tango	CXCR1_Fw	CGCGGATCC ATGTCCAATATCACCG
	CXCR1_Rv	CCGGAATTC CAAATTTGAAGAGACATTAAC
CXCR2 Tango	CXCR2_Fw	CGCGGATCCATGGAGGACTTCAACATGGAG
	CXCR2_Rv	CCGGAATTCAGGGTCTGGAGGTG
CXCR3 Tango	CXCR3_Fw	CGCGGATCCATGGTTCTGGAAGTTTCC
	CXCR3_Rv	CCGGAATTCGAGGCCAGAATAACTAGC
CXCR4 Tango	CXCR4_Fw	CGCGGATCCATGGAGGGGATATCAATC
	CXCR4_Rv	CCGGAATTCAGAGCTATGAAATGAGCTG
CXCR5 Tango	CXCR5_Fw	CGCGGATCCATGAACTATCCCCTGACC
	CXCR5_Rv	CCGGAATTCAAAGGTGGTCAGGCTAG
CXCR6 Tango	CXCR6_Fw	CGCGGATCCATGGCCGAACACGAC
	CXCR6_Rv	CCGGAATTCAGCTGGAACATGGAGG
CXCR7(S350A)	CXCR7_S350A_Fw	CTCCAGAGTCGCCGAGACGGAGTAC
	CXCR7_S350A_Rv	GCATCGATGAGCTTGGTG
CXCR7(T352A)	CXCR7_T352A_Fw	AGTCTCAGAGGCCGAGTACTCTGCC
	CXCR7_T352A_Rv	CTGGAGGCATCGATGAGC
CXCR7(S355A)	CXCR7_T355A_Fw	GACGGAGTACGCCGCTTGGAGC
	CXCR7_T355A_Rv	TCTGAGACTCTGGAGGCAT
CXCR7(S350A+T352A)	CXCR7_S350A+T352A_Fw	AGTCGCCGAGGCCGAGTACTCTG
	CXCR7_S350A+T352A_Rv	CTGGAGGCATCGATGAGC
CXCR7(T352A+S355A)	CXCR7_T352A+S355A_Fw	GGCCGAGTACGCCGCTTGGAGC
	CXCR7_T352A+S355A_Rv	TCTGAGACTCTGGAGGCATC
CXCR7(S350A+S355A)	CXCR7_S350A+S355A_Fw	GACGGAGTACGCCGCTTGGAGCAG
	CXCR7_S350A+S355A_Rv	TCGGCGACTCTGGAGGCA
CXCR7_Cluster1	CXCR7_Cluster1_Fw	AGGGCTCGCCAAGCTCATCGATGCCTCC
	CXCR7_Cluster1_Rv	GCTTTGGCCGCTACTTGAAGATGAAGGCC
CXCR7_Cluster2	CXCR7_Cluster2_Fw	GAGTACGCTGCCTTGGAGCAGAGCACC
	CXCR7_Cluster2_Rv	CGCCTCTGCGACTCTGGAGGCATCGATG
CXCR7_cSmBiT	CXCR7_cSmBiT_Fw	CGGGGTACCGAGGAGATCTGCCACCATGGGC
	CXCR7_cSmBiT_Rv	TCCCCGGGTTTGGTGCTCTGCTCCAAG
CXCR4_cSmBiT	CXCR4_cSmBiT_Fw	CGGGGTACCGAGGAGATCTGCCACCATGGGC
	CXCR4_cSmBiT_Rv	TCCCCGGGAGAGCTATGAAATGAGCTGG
CXCR4_LgBiT	CXCR4_LgBiT_Fw	CGGGGTACCGAGGAGATCTGCCACCATGGGC
	CXCR7_LgBiT_Rv	TCCCCGGGAGAGCTATGAAATGAGCTGG

Source Data file :Supplementary Fig. 6a



Source Data file :Supplementary Fig. 6c

