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Reporting Summary

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For	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	\square The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
\boxtimes	A description of all covariates tested
	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
\boxtimes	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
\boxtimes	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
\boxtimes	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated
	. Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection

Nikon Elements BR software (version 4.30.01), BD FACSDiva software (version 6.1.3), ChemiDoc MP Image Lab software (version 6.0.1, BioRad), QuantStudio3 Real-Time PCR System software (version 1.4.3, ThermoFisher)

Data analysis

FlowJo software (version 10), ChemiDoc MP Image Lab software (version 6.0.1, BioRad), QuantStudio3 (ThermoFisher), FASTQC (version 0.11.1), STAR (version 2.5.2), Salmon (version 0.7.2), GSEA (version 2.2.3), Go Analysis (Go Panther 11.1), Sciex Analyst software (version 1.6.2), Geneious software (Biomatters), Discovery Workbench software (version 4.0, Meso Scale Delivery), R (version 3.5.0), GraphPad Prism (versions 6 and 8), Applied Biosystems 7500 Real-Time PCR software (version 2.0.6), Simplified Presentation of Incredibly Complex Evaluations (SPICE) software (version 6.0), Partek Genomics Suite software (version 6.6)

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about <u>availability of data</u>

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

Data availability

The data generated are available from corresponding authors on reasonable request.

Fiel	d-sp	ecific	rep	orti	ng
Please	select the	one below	that is th	e best fit	t for

your research. If you are not sure, read the appropriate sections before making your selection. X Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences For a reference copy of the document with all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u> Life sciences study design All studies must disclose on these points even when the disclosure is negative. For BLT mouse studies, no statistical methods were used to predetermine sample size. At least 3 animals were used for each experimental Sample size group, the minimum to achieve statistical significance. Based on our previous data on SIV-infected ART-treated rhesus macaques, with a sample size of at least 7, we would be able to detect a significant difference between pre- and post- AZD5582 treatment samples in the levels of plasma RNA at the 0.05 significance level with a power of 0.90. Data exclusion was applied only to one RNAseq analysis represented on the heatmap of extended data Figure 7. One RM without on-ART Data exclusions viremia was excluded from this analysis for technical issues (higher than expected unmapped and multi-mapped reads, and lower than expected unique identified reads compared to the means). Replication In Figure 1b, symbols represent technical replicates of DMSO-normalized reporter signal induced by a dose titration of a panel of mono- and bivalent SMACm in a Jurkat luciferase reporter model of HIV-1 latency with 48 h exposure. In Extended Data Fig. 1f, fold induction of ncNF-kB target gene expression was measured by quantitative RT-PCR. Points represent two technical replicates. The data presented are representative of three independent experiments. In Extended Data Fig. 1g, for DMSO-normalized induction of luciferase activity from the Jurkat reporter model after exposure to AZD5582, points represent three replicates in one assay run, representative of several independent experiments. All attempts at replication were successful. In two independent experiments, plasma viremia was observed following AZD5582 administration to HIV-infected, ART-suppressed BLT mice. The use of non-human primates precludes our ability to replicate experiments. Sample sizes were chosen to maximize the likelihood of detecting statistical differences. For the study that examined the impact of AZD5582 administration on plasma and tissue viremia in BLT mice during ART suppression, mice Randomization were randomized for assignment to either experimental or control groups using randomization software available at random.org. For RM studies, peak plasma viral load (measured by standard assay) and plasma viral load before LRA intervention (as measured by ultrasensitive assay) were controlled for when allocated animals into experimental groups.

Blinding

Investigators were not blinded to group allocations or when assessing outcomes. In some instances, cells were pooled from individual humanized mice for each tissue and experimental group for the isolation of resting CD4+ T cells (Fig. 2).

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems	Methods		
n/a Involved in the study	n/a Involved in the study		
Antibodies	ChIP-seq		
Eukaryotic cell lines	Flow cytometry		
Palaeontology	MRI-based neuroimaging		
Animals and other organisms	·		
Human research participants			
Clinical data			

Antibodies

Antibodies used

rat anti-mouse CD24-biotin (clone M1/69), BD Biosciences (Cat. # 557436), 10ug biotin-labeled rat anti-mouse CD24 antibody was adsorbed to 1mg streptavidin-labeled magnetic Dynabeads ,https://www.bdbiosciences.com/us/applications/research/ stem-cell-research/cancer-research/mouse/purified-rat-anti-mouse-cd24-m169/p/557436 anti-cIAP1-unconjugated (clone EPR4673), Abcam (Cat. # 108361), 1:1,000 dilution, https://www.abcam.com/ciap1-antibodyepr4673-ab108361.html

anti-p100/p52-unconjugated (clone 18D10), Cell Signaling Technology (Cat. # 3017), 1:1,000 dilution, https:// www.cellsignal.com/products/primary-antibodies/nf-kb2-p100-p52-18d10-rabbit-mab/3017 anti-IkBa-unconjugated (clone 44D4), Cell Signaling Technology (Cat. # 4812), 1:1,000 dilution, https://www.cellsignal.com/ products/primary-antibodies/ikba-44d4-rabbit-mab/4812

anti-clAP2-unconjugated (clone E40), Abcam (Cat. # ab32059), 1:1,000 dilution, https://www.abcam.com/ciap2-antibody-e40-ab32059.html

anti-beta-actin-HRP (clone AC-15), Abcam (Cat. # ab49900), 1:30,000 dilution, https://www.abcam.com/beta-actin-antibody-ac-15-hrp-ab49900.html

 $anti-clAP1-unconjugated (goat polyclonal IgG), R\&D Systems (Cat. \# AF8181, Lot \# KH50516111), 10 ug/ml dilution, https://www.rndsystems.com/products/human-ciap-1-hiap-2-antibody_af8181$

anti-CD3-BV421 (clone SP34-2), BD Biosciences (Cat. # 562877), 1:250 dilution, https://www.bdbiosciences.com/us/reagents/research/antibodies-buffers/immunology-reagents/anti-human-antibodies/cell-surface-antigens/bv421-mouse-anti-human-cd3-sp34-2/p/562877

anti-CD16-BV605 (clone 3G8), BD Biosciences (Cat. # 563172), 1:50 dilution, https://www.bdbiosciences.com/us/applications/research/stem-cell-research/cancer-research/human/bv605-mouse-anti-human-cd16-3g8/p/563172

anti-CD4-BV711 (clone L200), BD Biosciences (Cat. # 563913), 1:50 dilution, https://www.bdbiosciences.com/us/reagents/research/antibodies-buffers/immunology-reagents/anti-non-human-primate-antibodies/cell-surface-antigens/bv711-mouse-anti-human-cd4-l200/p/563913

anti-CD14-BV786 (clone M5E2), BD Biosciences (Cat. # 563698), 1:50 dilution, https://www.bdbiosciences.com/us/applications/research/stem-cell-research/hematopoietic-stem-cell-markers/human/negative-markers/bv786-mouse-anti-human-cd14-m5e2/p/563698

anti-CD123-PerCP-Cy5.5 (clone 7G3), BD Biosciences (Cat. #558714), 1:25 dilution, https://www.bdbiosciences.com/us/applications/research/b-cell-research/surface-markers/human/percp-cy55-mouse-anti-human-cd123-7g3/p/558714 anti-CD20-PE-CF594 (clone 2H7), BD Biosciences (Cat. # 562295), 1:500 dilution, https://www.bdbiosciences.com/us/applications/research/stem-cell-research/hematopoietic-stem-cell-markers/human/negative-markers/pe-cf594-mouse-anti-human-cd20-2h7/p/562295

anti-CD8-PE-Cy7 (clone SK1), BD Biosciences (Cat. # 335787), 1:500 dilution, https://www.bdbiosciences.com/us/reagents/research/clinical-research---ruo-gmp/single-color-antibodies/pe-cytrade7-mouse-anti-human-cd8-sk1/p/335787 anti-CD11c-Alexa700 (clone 3.9), Ebioscience (Cat. # 50-112-9413), 1:50 dilution, https://www.thermofisher.com/antibody/product/CD11c-Antibody-clone-3-9-Monoclonal/56-0116-42

anti-HLA-DR-APC-Cy7 (clone L243), BD Biosciences (Cat. # 335796), 1:50 dilution, https://www.bdbiosciences.com/us/applications/research/stem-cell-research/mesenchymal-stem-cell-markers-bone-marrow/human/negative-markers/apc-cytrade7-mouse-anti-human-hla-dr-l243/p/335796

anti-p100-unconjugated (clone EPR18756), Abcam (Cat. # ab191594), 1:25 dilution, https://www.abcam.com/nfkb-p100nfkb2-antibody-epr18756-ab191594.html

anti-CD45-APC (clone HI30), BD Biosciences (Cat. # 555485), 3 ul/test, https://www.bdbiosciences.com/us/applications/research/stem-cell-research/cancer-research/human/apc-mouse-anti-human-cd45-hi30/p/555485

anti-CD3-FITC (clone HIT3a), BD Biosciences (Cat. # 555339), 3 ul/test, https://www.bdbiosciences.com/us/applications/research/t-cell-immunology/th-1-cells/surface-markers/human/fitc-mouse-anti-human-cd3-hit3a/p/555339

anti-CD19-PE (clone HIB19), BD Biosciences (Cat. # 555413), 3 ul/test, https://www.bdbiosciences.com/us/applications/research/clinical-research/oncology-research/blood-cell-disorders/surface-markers/human/pe-mouse-anti-human-cd19-hib19/p/555413

anti-CD4-PerCP (clone SK3), BD Biosciences (Cat. # 347324), 3 ul/test, https://www.bdbiosciences.com/us/applications/research/t-cell-immunology/th-1-cells/surface-markers/human/percp-mouse-anti-human-cd4-sk3-also-known-as-leu3a/p/347324 anti-CD4-PE (clone RPA-T4), BD Biosciences (Cat. # 555347), 3 ul/test, https://www.bdbiosciences.com/us/applications/research/t-cell-immunology/th-1-cells/surface-markers/human/pe-mouse-anti-human-cd4-rpa-t4/p/555347

anti-CD8-PerCP (clone SK1), BD Biosciences (Cat. # 347314), 3 ul/test, https://www.bdbiosciences.com/us/reagents/research/clinical-research---ruo-gmp/single-color-antibodies/percp-mouse-anti-human-cd8-sk1/p/347314

anti-CD45-APC-Cy7 (clone 2D1), BD Biosciences (Cat. # 557833), 3 ul/test, https://www.bdbiosciences.com/us/applications/research/stem-cell-research/cancer-research/human/apc-cy7-mouse-anti-human-cd45-2d1/p/557833

anti-CD3-PE-Cy7 (clone SK7), BD Biosciences (Cat. # 557851), 3 ul/test, https://www.bdbiosciences.com/us/applications/research/t-cell-immunology/th-1-cells/surface-markers/human/pe-cy7-mouse-anti-human-cd3-sk7-also-known-as-leu-4/p/557851

anti-CD8-FITC (clone SK1), BD Biosciences (Cat. # 340692), 3 ul/test, https://www.bdbiosciences.com/us/applications/clinical/blood-cell-disorders/asr-reagents/cd8-fitc-sk1/p/340692

anti-CD38-APC (clone HB7), BD Biosciences (Cat. # 340439), 3 ul/test, https://www.bdbiosciences.com/us/applications/research/t-cell-immunology/regulatory-t-cells/surface-markers/human/apc-mouse-anti-human-cd38-hb7/p/340439

anti-HLA-DR-PE (clone TU36), BD Biosciences (Cat. # 555561), 3 ul/test, https://www.bdbiosciences.com/us/applications/research/stem-cell-research/mesenchymal-stem-cell-markers-bone-marrow/human/negative-markers/pe-mouse-anti-human-hla-dr-tu36-also-known-as-t36-t36/p/555561

anti-CD3-BV421 (clone UCHT1), BD Biosciences (Cat. # 562426), 3 ul/test, https://www.bdbiosciences.com/us/applications/research/t-cell-immunology/th-1-cells/surface-markers/human/bv421-mouse-anti-human-cd3-ucht1-also-known-as-ucht-1-ucht-1/p/562426

anti-HLA-DR-PerCP (clone L243), BD Biosciences (Cat. # 347364), 3 ul/test, https://www.bdbiosciences.com/us/applications/research/stem-cell-research/mesenchymal-stem-cell-markers-bone-marrow/human/negative-markers/percp-mouse-anti-human-hla-dr-l243/p/347364

anti-CD4-BV605 (clone RPA-T4), BD Biosciences (Cat. # 562658), 3 ul/test, https://www.bdbiosciences.com/us/applications/research/t-cell-immunology/th-1-cells/surface-markers/human/bv605-mouse-anti-human-cd4-rpa-t4/p/562658 anti-CD8-APC-Cy7 (clone SK1), BD Biosciences (Cat. # 557834), 3 ul/test, https://www.bdbiosciences.com/us/reagents/research/antibodies-buffers/immunology-reagents/anti-human-antibodies/cell-surface-antigens/apc-cy7-mouse-anti-human-cd8-sk1/p/557834

anti-CD25-APC (clone 2A3), BD Biosciences (Cat. # 340938), 3 ul/test, https://www.bdbiosciences.com/us/applications/clinical/blood-cell-disorders/asr-reagents/cd25-apc-2a3/p/340938

 $anti-CD45-V500 \ (clone \ H130), \ BD \ Biosciences \ \ (Cat. \# 560777), \ 3 \ ul/test, \ https://www.bdbiosciences.com/us/applications/research/stem-cell-research/cancer-research/human/v500-mouse-anti-human-cd45-hi30/p/560777$

anti-mouse IgG1k-APC (clone MOPC-21), BD Biosciences (Cat. # 555751), 3 ul/test, https://www.bdbiosciences.com/us/reagents/research/antibodies-buffers/immunology-reagents/anti-human-antibodies/cell-surface-antigens/apc-mouse-igg1-isotype-control-mopc-21/p/555751

anti-mouse IgG2ak-PerCP (clone X39), BD Biosciences (Cat. # 340765), 3 ul/test, https://www.bdbiosciences.com/us/

applications/clinical/blood-cell-disorders/asr-reagents/mouse-iggsub2asub-percp-x39/p/340765

anti-mouse IgG1k-PE (clone MOPC-21), BD Biosciences (Cat. #559320), 3 ul/test, https://www.bdbiosciences.com/us/applications/research/intracellular-flow/intracellular-antibodies-and-isotype-controls/anti-human-antibodies/pe-mouse-igg1-isotype-control-mopc-21/p/559320

anti-mouse IgG1k-PE-Cy7 (clone MOPC-21), BD Biosciences (Cat. # 557872), 3 ul/test, https://www.bdbiosciences.com/us/reagents/research/antibodies-buffers/immunology-reagents/anti-human-antibodies/cell-surface-antigens/pe-cy7-mouse-igg1-isotype-control-mopc-21/p/557872

anti-mouse IgG2ak-FITC (clone G155-178), BD Biosciences (Cat. # 553456), 3 ul/test, https://www.bdbiosciences.com/us/reagents/research/antibodies-buffers/immunology-reagents/anti-mouse-antibodies/cell-surface-antigens/fitc-mouse-igg2a-isotype-control-g155-178/p/553456

anti-CD3-APC-Cy7 (clone SP34-2), BD Biosciences (Cat. # 557757),

https://www.bdbiosciences.com/us/reagents/research/antibodies-buffers/immunology-reagents/anti-non-human-primate-antibodies/cell-surface-antigens/apc-cv7-mouse-anti-human-cd3-sp34-2/p/557757

anti-Ki-67-AF700 (clone B56), BD Biosciences (Cat. # 561277), https://www.bdbiosciences.com/us/applications/research/intracellular-flow/intracellular-antibodies-and-isotype-controls/anti-human-antibodies/alexa-fluor-700-mouse-anti-ki-67-b56/p/561277

anti-HLA-DR-PerCP-Cy5.5 (clone G46-6), BD Biosciences (Cat. # 560652), https://www.bdbiosciences.com/us/applications/research/stem-cell-research/mesenchymal-stem-cell-markers-bone-marrow/human/negative-markers/percp-cy55-mouse-anti-human-hla-dr-g46-6/p/560652

anti-CCR5-APC (clone 3A9), BD Biosciences (Cat. # 550856), https://www.bdbiosciences.com/us/applications/research/t-cell-immunology/th-1-cells/surface-markers/human/apc-mouse-anti-human-cd195-3a9/p/550856

anti-CD8-BV711 (clone RPA-T8), BD Biosciences (Cat. # 563677), https://www.bdbiosciences.com/us/reagents/research/antibodies-buffers/immunology-reagents/anti-human-antibodies/cell-surface-antigens/bv711-mouse-anti-human-cd8-rpa-t8/p/563677

anti-CD4-BV650 (clone OKT4), Biolegend (Cat. # 317436), https://www.biolegend.com/en-us/products/brilliant-violet-650-anti-human-cd4-antibody-7786

anti-PD-1-BV421 (clone EH12.2H7), Biolegend (Cat. # 329920), https://www.biolegend.com/en-us/products/brilliant-violet-421-anti-human-cd279-pd-1-antibody-7191

anti-CD3-AF700 (clone SP34-2), BD Bioscience (Cat. # 557917), https://www.bdbiosciences.com/us/reagents/research/antibodies-buffers/immunology-reagents/anti-non-human-primate-antibodies/cell-surface-antigens/alexa-fluor-700-mouse-anti-human-cd3-sp34-2/p/557917

anti-CD69-PE-CF594 (clone FN50), BD Bioscience (Cat. # 562617), https://www.bdbiosciences.com/us/applications/research/t-cell-immunology/regulatory-t-cells/surface-markers/human/pe-cf594-mouse-anti-human-cd69-fn50-also-known-as-fn-50/p/562617

anti-CD25-PE-Cy7 (clone BC96), Biolegend (Cat. # 302612), https://www.biolegend.com/en-us/products/pe-cy7-anti-human-cd25-antibody-1909

anti-CD45RA-PE-Cy7 (clone5H9), BD Biosciences (Cat. # 561216), https://www.bdbiosciences.com/us/applications/research/b-cell-research/surface-markers/non-human-primates/pe-cy7-mouse-anti-human-cd45ra-5h9/p/561216

anti-CD62L-PE (clone SKII), BD Biosciences (Cat. # 654666), https://www.bdbiosciences.com/us/applications/clinical/blood-cell-disorders/asr-reagents/cd62l-pe-sk11-also-known-as-anti-leu-8/p/654666

anti-CD95-BV605 (clone DX2), Biolegend (Cat. # 305627), https://www.biolegend.com/en-us/products/brilliant-violet-605-anti-human-cd95-fas-antibody-8778

anti-CD28-PE-Cy5.5 (clone CD28.2), Beckman Coulter (Cat. # B24027), https://www.beckman.com/reagents/coulter-flow-cytometry/antibodies-and-kits/single-color-antibodies/cd28/b24027

Validation

The specificity of the antibodies purchased from commercial sources (BD Biosciences, Abcam, Cell Signaling Technology, R&D Systems, Ebioscience, Biolegend, and Beckman Coulter) were validated by the manufacturer as noted on their website (links provided above for each antibody).

Eukaryotic cell lines

Policy information about cell lines

Cell line source(s)

Jurkat Clone E6-1 cells (American Type Culture Collection TIB-152), TZM-bl cells (NIH AIDS Reagent Repository) and HEK 293T cells (European Collection of Authenticated Cell Cultures)

Authentication

Cell lines were authenticated by morphological identification and virus susceptibility profiles.

Mycoplasma contamination

Cell lines were tested negative for mycoplasma by the supplier

Commonly misidentified lines (See ICLAC register)

No commonly misidentified cell lines were used

Animals and other organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research

Laboratory animals

BLT mice were constructed using 12-15 week old female NOD.Cg-Prkdcscid ll2rgtm1Wjl/SzJ mice (NSG; The Jackson Laboratory, Bar Harbor, ME) mice. Female 20 week old BALB/cJ (The Jackson Laboratory, Bar Harbor, ME) were used for the serum chemistry analysis. Three healthy male rhesus macaques (Macaca mulatta) of Indian origin, age 6-7 years, were utilized for the AZD5582 pharmacokinetic study. Twenty-one male and female Mamu-B*08 and -B*17 negative rhesus macaques, age 3-6 years, were infected with SIVmac239 and treated with ART (Supplementary Table 7).

Wild animals The study did not involve wild animals.

Field-collected samples The study did not involve samples collected from the field.

Ethics oversight

Mice were maintained under specific pathogen-free conditions by the Division of Comparative Medicine at the University of North Carolina, Chapel Hill. Mouse experiments were conducted in accordance with NIH guidelines for the housing and care of laboratory animals and in accordance with protocols reviewed and approved by the Institutional Animal Care and Use Committee at the University of North Carolina, Chapel Hill. Healthy Rhesus macaques for pharmacokinetic studies were housed at GlaxoSmithKline and all procedures were conducted in accordance with the GlaxoSmithKline Policy on the Care, Welfare, and Treatment of Laboratory Animals and were reviewed by the IACUC at GlaxoSmithKline. Rhesus macaques infected with SIV were housed at the Yerkes National Primate Research Center (Atlanta, GA) and treated in accordance with Emory University and Yerkes National Primate Research Center Institutional Animal Care and Use Committee regulations.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Human research participants

Policy information about studies involving human research participants

Population characteristics Cells used in

Cells used in the QVOA and RNAseq experiments were obtained from participants stably suppressed on ART. At the time of sample donation, participants had a mean age of 43 years [range, 26-61 years], a mean duration on ART of 7 years [range, >6 months -22 years] and a mean CD4 count of 634 [range, 372-1364 cells/µl]. All participants were male, and 88% were Caucasians and 12% African American. Twenty-five percent of the participants were treated during acute infection and 75% during chronic infection.

Recruitment

Cells used in the QVOA assays were selected randomly across participants enrolled in a longitudinal reservoir measurement study and thus should not be subjected to self-selection bias. For the RNA seq experiments, cells from participants with a demonstrated increase in cell associated HIV RNA following ex-vivo exposure to AZD5582 were selected, potentially introducing a self-selection bias. However, given that these were global human gene expression measurements, we do not believe our results were impacted by this bias.

Ethics oversight

All human subjects samples were obtained under a specimen procurement protocol reviewed and approved by the University of North Carolina Biomedical Institutional Review Board and the McGill University Health Centre Ethical Review Board. Informed consent was obtained from all participants.

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Flow Cytometry

Plots

Confirm that:

The axis labels state the marker and fluorochrome used (e.g. CD4-FITC).

The axis scales are clearly visible. Include numbers along axes only for bottom left plot of group (a 'group' is an analysis of identical markers).

All plots are contour plots with outliers or pseudocolor plots.

X A numerical value for number of cells or percentage (with statistics) is provided.

Methodology

Sample preparation Sample preparation for the flow cytometric analysis of peripheral blood and tissues from humanized mice and rhesus macaques is detailed in the Methods section.

Instrument Flow cytometry data was collected on BD LSRII, BD LSRFortessa, BD FACSAria LSR II, or BD FACSCanto instruments using BD FACSDiva software.

Software Flow cytometry data was analyzed with FlowJo software.

Cell population abundance Resting CD4+ T cells represented 0.12-9.48% (mean: 3.42%) of the total cell population. Prior to sorting of macaque resting CD4+ T cells by FACS, CD4+ T cells were enriched by magnetic bead selection. Post-sort purity was 97.8%.

Gating strategy

For the analysis of the frequency and phenotype of different human immune cell populations in the peripheral blood and tissues of BLT mice, an antibody specific for human CD45, a pan leukocyte marker, was used first to gate human leukocytes. Gates to define positive and negative populations were defined by isotype controls when appropriate.

Tick this box to confirm that a figure exemplifying the gating strategy is provided in the Supplementary Information.