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Reporting Summary

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For	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	\square The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
\boxtimes	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
\boxtimes	A description of all covariates tested
	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
\boxtimes	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
\boxtimes	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
\boxtimes	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i>), indicating how they were calculated

Our web collection on $\underline{statistics\ for\ biologists}$ contains articles on many of the points above.

Software and code

Policy information about availability of computer code

Data collection No software was used for data collection.

Data analysis

(More detailed software information can be found in Supplementary Table 23)

Bedtools 2.25.0 ClinVar 01-09-2017 OMIM 01-10-2017 HapMap Phase III

BWA Software Package 0.7.10,0.7.17

PICARD 1.119,2.18.11 Samtools 0.1.19,1.9 GATK Tools 3.3, 3.5, 4.0.8.0 dbSNP Build 150 VerifyBamID 1.1.1, 1.1.3

Eagle 2.0 SHAPEIT 2.17 PLINK 1.90 ADMIXTURE 1.3.0 GENESIS 0.2.6 CLUMPP 1.1.2 ADMIXTOOLS 1.0 TREEMIX 1.1.3 VCFTools 0.1.15

Selscan 1.1.0b Ensembl 01-10-2017 GeneCards 01-10-2017

gnomAD 2.0.2 exomeAD 2.0.3 BioMart 2.34.2 eDGAR 10-11-2016 HGVD 2.3 GWAS Catalogue v.2019-10-14 VFP 90 GTEx 7 MarViN r1 https://github.com/Illumina/MarViN FIGENSOFT v 7 2 1 MALDER v.1.0 Beagle 4.1. RFMix v2 SNPEff 4.3-3 FxAC r2.01 Haplogrep2 2.1.1. AMY-tree 2.0 ClueGO v.2.5.1 ALOFT 1.0 DisGenet 6.0 eDGAR database release 10/11/2016 CTDbase update 2019 SWEED 3.3.1

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors/reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets

dbPHSP https://jjwanglab.org/dbphsp

- A list of figures that have associated raw data
- A description of any restrictions on data availability

WGS data from the H3Africa projects used in this study have been deposited in the European Genome-phenome Archive (EGA) with accession numbers:

H3AfricaChipDesign: Study EGAS00001002976
TrypanoGEN1: Data Set EGAD00001004393
TrypanoGEN2: Data Set EGAD00001004220
MALSIC: Data Set EGAD00001004557
AWI-GEN: Data Set EGAD00001004448
CAfGEN: Data Set EGAD00001004533
ELSI: Data Set EGAD00001004316
NEEDI: Data Set EGAD00001004334
ACCME: Data Set EGAD00001004505

DNA samples are archived in H3Africa biorepositories as part of the H3Africa Consortium agreement. The Data and biospecimens are available on request through the Data and Biospecimen Access Committee of the H3Africa consortium. Novel SNVs identified and reported here will be deposited into the database of Single Nucleotide Polymorphisms (dbSNP).

Field-specific reporting

Please select the one belo	w that is the best fit for your research	n. If you are not sure, read the appropriate sections before making your selection.	
Life sciences	Behavioural & social sciences	Ecological, evolutionary & environmental sciences	
For a reference copy of the document with all sections, see nature.com/documents/nr-reporting-summary-flat.pdf			

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size

No statistical method was used to determine sample size. Sample size was maximised to include as wide a diversity of African populations and ethnolinguistic groups as possible. For specific analysis methods, subsets of the data (e.g. high depth of coverage data) were used to ensure robust conclusions.

Data exclusions

Exclusion criteria were pre-established on the basis of relatedness and sequencing quality. Samples were excluded based on sequencing-quality control metrics generated prior to any downstream analysis; this included poor mapping of reads to the reference genome, missingness thresholds, and familial relatedness. Single nucleotide variants were excluded based on excessive missingness, deviations from

		berg equilibrium, call ambiguity and low minor allele frequencies. In addition, where a genetically homogeneous set of individuals ed for an analysis, possible outliers were identified and removed.		
Replication		esults were not externally replicated.		
Randomization	The experir	ments were not randomized.		
Blinding	Investigators were not blinded to the allocation during analyses, since this it not an association study.			
Reportin	g for	specific materials, systems and methods		
/e require information	on from auth	nors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, it to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.		
Materials & exp	perimenta	al systems Methods		
/a Involved in th		n/a Involved in the study		
Antibodies		ChIP-seq		
Antibodies Antibodies Eukaryotic Palaeontol Animals an	cell lines	Flow cytometry		
Palaeontol	ogy	MRI-based neuroimaging		
Animals an	d other orga	nisms		
Human res	earch partici	pants		
Clinical dat	a			
luman rese	arch pa	rticipants		
olicy information a	about <u>studi</u>	es involving human research participants		
Population chara	cteristics	There are no covariate-relevant population characteristics. Individuals were chosen to represent a broad geographical and ethnolinguistic diversity and were sampled from three ongoing genomics consortium projects on the continent - the H3Africa Consortium, the Southern African Human Genome Programme (SAHGP), and the Trypanosomiasis Genomics Network of the H3Africa Consortium (TryopanoGEN).		
Recruitment		The participants in the study were previously recruited to individual cohorts as part of ongoing genomics projects on the continent:		
		(i) Samples from 519 individuals from 8 projects in the H3Africa consortium were submitted for consideration. Individuals were predominantly recruited as control individuals from the respective studies, with the exception of those from case-only studies in Cameroon (Sickle cell), Botswana (HIV), Mali (families with neurological disease), and Benin (sickle cell). Individuals with high relatedness (PIHAT> 0.18) were removed from the analysis. Individuals comprising the Berom from Nigeria were recruited as controls for a cervical cancer study, and thus are all female. (ii) A total of 300 (control) individuals were recruited from five countries (Uganda, Zambia, Democratic Republic of the Congo, Cameroon and Ivory Coast) participating in the TrypanoGEN project to study the host and parasite genetics and genomics of trypanosomiasis infection across Africa.		
		(iii) 16 (unaffected) individuals were recruited by the Southern African Human Genome Programme. These individuals met inclusion criteria: male, over the age of 18 years, four grandparents who speak the same language as the participant, not known to be related to the other participants in the study, and willing to provide broad informed consent (including consent to share data and DNA for future studies approved by the HREC (Medical)). Two main Bantu-speaking ethnolinguistic groups were included: The Sotho (Sotho-Tswana speakers; n=8) were recruited from in and around the town of Ventersburg in the Free State Province and the Xhosa speakers (Nguni language: n=7) were recruited from the Fastern Cape Province. One individual was a		

Ethics oversight

Ethics approval (country and institutional) was obtained by each PI of the individual studies. We attach further details of each study's approval, which will be included in the online methods and supplementary materials section of the paper (detailed information was not included in initial submission).

Note that full information on the approval of the study protocol must also be provided in the manuscript.

Zulu speaker (Nguni language) from Johannesburg.