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# **Efficient evolution of human antibodies from general protein language models**

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**Supplementary Tables 1-13**



# **Supplementary Table 1: Summary of antibodies considered in this study.**

Information on the antibodies considered in each of our directed evolution campaigns. Matured indicates extensive somatic hypermutation from germline (and, in the case of MEDI8852, additional in-vitro affinity maturation). Source indicates how the antibody sequence was obtained; germline-inferred sequences were obtained from the original publications. Improved binding is defined as a 1.1-fold improvement or higher from wildtype. Preserved binding is defined as a sub-micromolar  $K_d$  for the screened antigen. ND: not determined.



# **Supplementary Table 2: MEDI8852 variants.**

Variants tested across two rounds of directed evolution, the corresponding Kabat-annotated regions, and  $K_d$  values for three HA antigens. The wildtype row is highlighted in gray; variants with improved affinity against the screened antigen (fold improvement > 1.1) are highlighted in blue; values with one-sided *t*-test *P* < 0.05 compared to wildtype are bolded. NB: no binding; ND: not determined; CV: coefficient of variation.



**Supplementary Table 3: Binding to Group 1 and Group 2 HAs.**

Binding affinity between MEDI8852 WT Fab and three variant Fabs against a panel of nine HAs. A  $K_d$  of  $\leq 0.001$  indicates an interaction with no observed dissociation when measured via BLI.



# **Supplementary Table 4: MEDI8852 UCA variants.**

Variants tested across two rounds of directed evolution, the corresponding Kabat-annotated regions, and  $K_d$  values for three HA antigens. In Round 2, all possible combinations involving K58S, V65P, P75R in the VH and G95P in the VL were made. The wildtype row is highlighted in gray; variants with improved affinity against the screened antigen (fold improvement  $> 1.1$ ) are highlighted in blue; values with one-sided *t*-test *P* < 0.05 compared to wildtype are bolded . *K*<sub>d</sub> app.: *K*<sub>d</sub> apparent; ND: not determined; CV: coefficient of variation.



# **Supplementary Table 5: mAb114 variants.**

Variants tested across two rounds of directed evolution, the corresponding Kabat-annotated regions, and  $K_d$  values for ebolavirus GP. Neutralization  $IC_{50}$  values were also determined for mAb114 WT and all Round-2 variant IgGs against GP-pseudotyped lentivirus. The wildtype row is highlighted in gray; variants with improved affinity against the screened antigen (fold

improvement > 1.1) are highlighted in blue; values with one-sided *t*-test *P* < 0.05 compared to wildtype are bolded. ND: not determined; CV: coefficient of variation.



# **Supplementary Table 6: mAb114 UCA variants.**

Variants tested across two rounds of directed evolution, the corresponding Kabat-annotated regions, and  $K_d$  values (for both IgG and Fab versions) for ebolavirus GP. The wildtype row is highlighted in gray; variants with improved affinity against the screened antigen (fold improvement > 1.1) are highlighted in blue; values with one-sided *t*-test *P* < 0.05 compared to wildtype are bolded. An asterisk (\*) indicates examples where some binding was observed but BLI data were not suitable for fitting.  $K_d$  app.:  $K_d$  apparent; ND: not determined; CV: coefficient of variation.



# **Supplementary Table 7: S309 variants.**

Variants tested across two rounds of directed evolution, the corresponding Kabat-annotated regions, and  $K_d$  values for antigens from three SARS-CoV-2 variants. The wildtype row is highlighted in gray; variants with improved affinity against the screened antigen (fold

improvement > 1.1) are highlighted in blue; values with one-sided *t*-test *P* < 0.05 compared to wildtype are bolded . W1: Wuhan-Hu-1; NE: no expression; ND: not determined; CV: coefficient of variation.



#### **Supplementary Table 8: REGN10987 variants.**

Variants tested across two rounds of directed evolution, the corresponding Kabat-annotated regions, and  $K_d$  values for antigens from two SARS-CoV-2 variants. Round-1 variants were prescreened as IgGs with a single replicate before testing the highest-avidity variants as Fabs and with multiple replicates. Neutralization  $IC_{50}$  values were also determined for REGN10987 WT and selected affinity-enhancing variant IgGs against D614G- and Beta-pseudotyped lentivirus. The wildtype row is highlighted in gray; variants with improved affinity against the screened antigen (fold improvement  $> 1.1$ ) are highlighted in blue; values with one-sided *t*-test  $P < 0.05$ 

compared to wildtype are bolded.  $K_d$  app.:  $K_d$  apparent; NA: not applicable; ND: not determined; CV: coefficient of variation.



# **Supplementary Table 9: C143 variants.**

Variants tested across two rounds of directed evolution, the corresponding Kabat-annotated regions, and  $K_d$  values for antigens from three SARS-CoV-2 variants. Neutralization IC<sub>50</sub> values were also determined for C143 WT and selected affinity-enhancing variant IgGs against D614G- and Beta-pseudotyped lentivirus. The wildtype row is highlighted in gray; variants with improved affinity against the screened antigen (fold improvement > 1.1) are highlighted in blue; values with one-sided *t*-test  $P < 0.05$  compared to wildtype are bolded. An asterisk (\*) indicates examples where some binding was observed but BLI data were not suitable for fitting. W1: Wuhan-Hu-1;  $K_d$  app.:  $K_d$  apparent; NB: no binding; ND: not determined; CV: coefficient of variation.





#### **Supplementary Table 10: Originality of affinity-enhancing substitutions.**

Each row corresponds to an amino acid substitution that enhances the binding affinity of its corresponding variant antibody, and some of which also enhance affinity in combination with other substitutions. We computed frequencies of amino acid substitutions among natural sequences using two datasets, UniRef90 and abYsis (**Methods**); UniRef90 was the sequence database used to train the language models in our algorithm and abYsis is a separate, curated database of natural antibody sequences. The "wildtype residue frequency" indicates the percentage of sequences in a multiple sequence alignment with the same residue as wildtype at the given position; the "mutant residue frequency" is the same statistic except for the mutant residue. The "top residue" indicates the amino acid with the highest frequency observed at the given site, the "top residue frequency" indicates the percentage of sequences that contain the top residue at the given site, and dashes indicate settings in which the mutant residue is also the top residue. Substitutions with frequencies up to 5% are considered "rare," those with frequencies above 5% and up to 10% are considered "uncommon," and those above 10% are considered

"common." Blue shading indicates substitutions to rare or uncommon residues according to frequency information from either UniRef90 or abYsis. The "site-independent rank" indicates the rank of the corresponding rare or uncommon substitution when ordering all possible singleresidue substitutions according to the likelihood ratio to wildtype under a UniRef90-based siteindependent model (**Supplementary Table 12**), where a rank of 1 indicates the highest likelihood ratio.





**Supplementary Table 11: Germline frequencies of affinity-enhancing substitutions.**

Each row corresponds to an amino acid substitution that enhances the binding affinity of its corresponding variant antibody, and some of which also enhance affinity in combination with other substitutions. We computed frequencies of amino acid substitutions based on IMGT germline sequences, where for each substitution, we restrict the multiple sequence alignment to only include sequences from its corresponding V or J gene, which are listed in the "gene" column (**Methods**). The "number of database sequences" indicates the number of germline sequences found for that gene in IMGT (the denominator of frequency values). The "wildtype residue frequency" indicates the percentage of sequences in a multiple sequence alignment with the same residue as wildtype at the given position; the "mutant residue frequency" is the same statistic except for the mutant residue. The "top residue" indicates the amino acid with the highest frequency observed at the given site, the "top residue frequency" indicates the percentage of sequences that contain the top residue at the given site, and dashes indicate settings in which the mutant residue is also the top residue. Substitutions with frequencies up to 5% are considered "rare," those with frequencies above 5% and up to 10% are considered "uncommon," and those above 10% are considered "common." Blue shading indicates substitutions to rare or uncommon residues according to frequency information from IMGT germline sequences.



# **Supplementary Table 12: Benchmarking results comparing sequence-based methods for proposing substitutions.**

We compared the changes in avidity of substitutions recommended by four sequence-based baseline methods (abYsis, UniRef90, AbLang, and Sapiens) to those of the substitutions used in the first round of our evolutionary campaigns (this study). Summary statistics are: (1) the fraction with improved binding, defined as a change in IgG apparent  $K_d$  with nominal one-sided *t*-test  $P < 0.05$ ; (2) the median fold improvement across all substitutions; and (3) the maximum fold improvement across all substitutions. The best values in each category are shaded light blue. The full set of benchmark results is provided in **Supplementary Data 3**.



# **Supplementary Table 13: Enrichment of high-fitness variants based on language-model-recommended substitutions.**

Each row corresponds to a protein tested via a high-throughput scanning mutagenesis assay that measures various notions of protein fitness, which are summarized in the "Fitness setting" column. All assays involve deep mutational scans that profile variants that

represent 90% or more coverage of all single-residue substitutions except for that of PafA, which changes every residue to either a glycine or a valine. The cutoff indicates the study-specific criterion for determining a high-fitness variant. The "Sample size" indicates the number of acquired variants ( $|\mathcal{A}|$ ) and "Sample successes" indicates the number of those variants with high fitness according to the cutoff. The "Population size" indicates the number of variants profiled in the scanning mutagenesis assay, where "Population successes" indicates the number of those variants with high fitness according to the cutoff. "Hit rate" indicates the percentage fraction of high-fitness variants among the language-model-recommended variants (sample successes divided by sample size) whereas "Background" indicates the percentage fraction of high-fitness variants among all single-residue variants (population successes divided by population size). The hypergeometric *P* value computes enrichment of high-fitness variants among the acquired variants by assuming that the number of sample successes has a hypergeometric null distribution with parameters given by the other values (sample size, population successes, and population size); blue shading indicates a one-sided, hypergeometric *P*-value of less than 0.05.

# **Supplementary Information**

## *Antibody sequences*

Below are the antibody protein sequences defined as wildtype in this study:

• MEDI8852 VH:

QVQLQQSGPGLVKPSQTLSLTCAISGDSVSSYNAVWNWIRQSPSRGLEWLGRTY YRSGWYNDYAESVKSRITINPDTSKNQFSLQLNSVTPEDTAVYYCARSGHITVFG VNVDAFDMWGQGTMVTVSS

• MEDI8852 VL:

DIQMTQSPSSLSASVGDRVTITCRTSQSLSSYTHWYQQKPGKAPKLLIYAASSRGS GVPSRFSGSGSGTDFTLTISSLQPEDFATYYCQQSRTFGQGTKVEIK

• MEDI8852 UCA VH:

QVQLQQSGPGLVKPSQTLSLTCAISGDSVSSNSAAWNWIRQSPSRGLEWLGRTY YRSKWYNDYAVSVKSRITINPDTSKNQFSLQLNSVTPEDTAVYYCARGGHITIFG VNIDAFDIWGQGTMVTVSS

• MEDI8852 UCA VL:

DIQMTQSPSSLSASVGDRVTITCRASQSISSYLNWYQQKPGKAPKLLIYAASSLQS GVPSRFSGSGSGTDFTLTISSLQPEDFATYYCQQSRTFGQGTKVEIK

 $\bullet$  mAb114 VH:

EVQLVESGGGLIQPGGSLRLSCAASGFALRMYDMHWVRQTIDKRLEWVSAVGP SGDTYYADSVKGRFAVSRENAKNSLSLQMNSLTAGDTAIYYCVRSDRGVAGLF DSWGQGILVTVSS

• mAb114 VL:

DIQMTQSPSSLSASVGDRITITCRASQAFDNYVAWYQQRPGKVPKLLISAASALH AGVPSRFSGSGSGTHFTLTISSLQPEDVATYYCQNYNSAPLTFGGGTKVEIK

• mAb114 UCA VH:

EVQLVESGGGLVQPGGSLRLSCAASGFTFSSYDMHWVRQATGKGLEWVSAIGT AGDTYYPGSVKGRFTISRENAKNSLYLQMNSLRAGDTAVYYCVRSDRGVAGLF DSWGQGTLVTVSS

• mAb114 UCA VL:

DIQMTQSPSSLSASVGDRVTITCRASQGISNYLAWYQQKPGKVPKLLIYAASTLQ SGVPSRFSGSGSGTDFTLTISSLQPEDVATYYCQKYNSAPLTFGGGTKVEIK

• S309 VH:

QVQLVQSGAEVKKPGASVKVSCKASGYPFTSYGISWVRQAPGQGLEWMGWIST YNGNTNYAQKFQGRVTMTTDTSTTTGYMELRRLRSDDTAVYYCARDYTRGAW FGESLIGGFDNWGQGTLVTVSS

• S309 VL:

EIVLTQSPGTLSLSPGERATLSCRASQTVSSTSLAWYQQKPGQAPRLLIYGASSRA TGIPDRFSGSGSGTDFTLTISRLEPEDFAVYYCQQHDTSLTFGGGTKVEIK

• REGN10987 VH:

QVQLVESGGGVVQPGRSLRLSCAASGFTFSNYAMYWVRQAPGKGLEWVAVISY DGSNKYYADSVKGRFTISRDNSKNTLYLQMNSLRTEDTAVYYCASGSDYGDYL LVYWGQGTLVTVSS

• REGN10987 VL:

QSALTQPASVSGSPGQSITISCTGTSSDVGGYNYVSWYQQHPGKAPKLMIYDVSK RPSGVSNRFSGSKSGNTASLTISGLQSEDEADYYCNSLTSISTWVFGGGTKLTVL

• C143 VH:

EVQLVESGGGLVQPGGSLRLSCAASGFSVSTKYMTWVRQAPGKGLEWVSVLYS GGSDYYADSVKGRFTISRDNSKNALYLQMNSLRVEDTGVYYCARDSSEVRDHP GHPGRSVGAFDIWGQGTMVTVSS

• C143 VL:

QSALTQPASVSGSPGQSITISCTGTSNDVGSYTLVSWYQQYPGKAPKLLIFEGTKR SSGISNRFSGSKSGNTASLTISGLQGEDEADYYCCSYAGASTFVFGGGTKLTVL