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Reporting Summary

Statistics

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

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For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.		
n/a	Confirmed	
	×	The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
	x	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
×		The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
x		A description of all covariates tested
x		A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	×	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
×		For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i>) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>

Our web collection on statistics for biologists contains articles on many of the points above.

For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes

For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings

Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated

Software and code

Policy information about <u>availability of computer code</u>

Data collection

Data were collected using Labview 2018 software.

Data analysis

Data were analysed using Igor Pro 7.08, Origin 8.5, and ImageJ 1.8.0

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

The data supporting the findings of this work are deposited in Figshare, DOI: 10.6084/m9.figshare.14544495

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size

16,447 curves were measured from 129 individual RNA molecules. The number of curves measured from a single molecule before it breaks varies widely, hence sample sizes could not be pre-determined. Approximate minimum sample sizes were chosen to ensure: (i) sufficient sampling to detect intermediates present at the 1% level (i.e., a minimum of 100-200 curves); (ii) sufficient sampling to obtain at least 10 measurements of contour length changes for every intermediate detected (including very rare states), to allow for reliable average values; and (iii) a minimum of 5 molecules for baseline measurements of WT and mutant xrRNAs, or 2 molecules for measurements with anti-sense oligos.

Data exclusions

No data from measurements of single RNA molecules were excluded. All measurements that did not involve correctly assembled single RNA molecules--as reflected via incorrect persistence length, contour length, or overstretching transition force--were excluded.

Replication

Each curve represents an independent measurement of the unfolding/refolding. Measurements on different molecules represent replicates of the experiment. For each experimental condition, 162-5212 independent measurements were done on 2-69 molecules (replicates), as listed in Supplementary Tables 1-5. All the force-extension curves were reproducible across the different molecules (replicates). 3 replicates were done of gel images.

Randomization

Randomization is not relevant to this study since there were no experimental groups. The individual molecules being studied were inherently randomized in solution.

Blinding

There were no experimental groups in this study, hence blinding is not relevant. All curves were analyzed in the same way.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems	Methods	
n/a Involved in the study	n/a Involved in the study	
X Antibodies	ChIP-seq	
Eukaryotic cell lines	Flow cytometry	
Palaeontology and archaeology	MRI-based neuroimaging	
X Animals and other organisms	·	
Human research participants		
▼ Clinical data		
Dual use research of concern		