1 Supplementary Information

A point mutation in AgrC determines cytotoxic or colonizing properties associated with phenotypic variants of ST22 MRSA strains.

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21

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26 Supplementary Methods

27 Gel Filtration Chromatography for AgrC_{Y223C}

Analytical size exclusion experiments were conducted using Superdex S200 column to determine the oligomeric status of $AgrC_{Y223C}$. Also, $AgrC_{Y223C}$ was incubated with DTT to examine their effect on the Cysteine 223 and therefore on the oligomeric status of $AgrC_{Y223C}$.

31 Toxin ELISAs

Levels of α-toxin and PVL were determined by ELISA using toxin-specific antibodies
 provided by GSK Vaccines (Belgium) and bioMérieux R&D Immunodiagnostic (France),
 respectively^{1,2}.

36 SupplementaryTable 1. Strains and plasmids used in this study

Strain/plasmid	Clinical diagnosis	MRSA/MSSA	Response	α-toxin	PVL	Reference/Source
			Profile*	(µg/ml) [#]	(µg/ml) [#]	
(A) S. aureus strains						
PUNE08 (Sa08)	SSTI	MRSA-IV	С	75	19.6	3
PUNE08 agrC-Y223C	; -	MRSA-IV	Р	N.D	N.D	This study
M37 (Sa37)	SSTI	MRSA-IV	Р	9	1.5	3
M37 agrC-C223Y	-	MRSA-IV	С	N.D	N.D	This study
Sa6454	SSTI	MRSA-IV	С	18	1.6	This study
SaN08	Meningitis	MRSA-IV	С	32	8	3
Sa113	Brain abscess	MRSA-IV	С	20	3.2	3
Sa114	Cerebral abscess	MRSA-IV	С	23	4	3
Sa754	Invasive infection	MRSA-IV	С	20	1	3
Sa165	colonizing strain	MRSA-IV	С	19.3	2.3	3
Sa115	colonizing strain	MRSA-IV	Р	0.2	0.08	3
(B) EMRSA-15 refere	ence S. aureus strains	6				
HAR22	SSTI	MRSA-IV	С	N.D	N.D	Provided by Dr. H. Lencastre ⁴
NCTC 13142	SSTI	MRSA-IV	С	N.D	N.D	Provided by A Kearns ⁵
(C) <i>E. coli</i> strains						
BL21 (DE3)	-	-				NEB
IM08B	-	-				6
(D) Plasmids						
pIMAY-Z	-	-				6
pET22b						Novagen Inc.
pET28a						7

MRSA, methicillin-resistant *S. aureus*; MSSA, methicillin-susceptible *S. aureus*; P, proliferative; C, cytotoxic; SSTI, skin and soft tissue infection. N.D, Not
 determined; Names in parenthesis for strains are nomenclature based on previous publication

39 * The response profile is determined by the pattern of proliferative responses elicited by different dilutions of bacterial supernatants against PBMC. A
40 proliferative profile was denoted if all dilutions elicited a proliferative response. A cytotoxic pattern was denoted if a) proliferation was only noted at the highest
41 dilution of the supernatants, and b) the supernatants (1:50 dilution) resulted in inhibition of PHA-induced responses in co-stimulation experiments.

42 [#]Amounts of toxins present in supernatants cultures grown to stationary phase (17hours)

57 SupplementaryTable 2. Toxin gene profile of the clinical ST22 S. aureus strains.

Strains	ST/SPA type	<i>agr</i> type	se/tsst-1 /egc	pvl	<i>luk</i> D/E	Staphylokinase	Hemolysins	Proteases	SSL genes	MSCRAMM/ Adhesin	Capsule/ Biofilm
PUNE08	ST22/t852	I	-/-/+	+	-	+	hla,hlb,hld,	aur, sspA,B,P	ssl 5,7,9	clf A,B, cna, eno, fnbA, sdrC,	5/ ica A,C,D
M37	ST22/t852	Ι	-/-/+	+	-	+	hla,hlb,hld,	aur, sspA,B,P	ssl 5,7,9	clf A,B, cna, eno, fnbA, sdrC	5/ ica A,C,D

Based on gene microarray data: *se,* staphylococcal enterotoxin; *pvl*, Panton-Valentine leucocidin; *hla*, alpha-hemolysin; *hlb*, beta-hemolysin; *hld*, deltahemolysin; *spl*, serine protease like; *sspA*, serine protease; *sspB*, cysteine protease staphopain B; *sspP*, staphopain A precursor; *ssl*, staphylococcal superantigen-like; *clf*, clumping factor; *cna*, collagen adhesins; *eno*, enolase; *fib*, fibrinogen binding-protein; *fnbA*, fibronectin A; *sdrC*, serine-aspartate repeatcontaining protein C

Gene	Primer	Sequence (5´-3´)
agrC	AgrC_F	TTATGACTTCGCAGATTATTCTGTACT
	AgrC_R	TAGAAAAACTTTCTTGGAACAATTCAT
hla	hla_F	CGGCACATTTGCACCAATAAGGC
	hla_R	GGTTTAGCCTGGCCTTCAGC
RNAIII	RNAIII_F	GTGATGGAAAATAGTTGATGAGTTGTTT
	RNAIII_R	GAATTTGTTCACTGTGTCGATAATCC
psma	psmα_F	TATCAAAAGCTTAATCGAACAATTC
	psmα_R	CCCCTTCAAATAAGATGTTCATATC
lukSPV	lukSPV_F	GTGTTGTTCTTCTAGTAGCATGAGT
	lukSPV_R	ATCACATCATTAGGTAAAATGTCTG
spa	spa_F	CAACGGAGTACATGTCGTTA
	spa_R	GCTTGAGCTTTGTTAGCATC
clfA	clfA_F	CAAAGGTGATTTAGCTTTACGTTCAACTT
	clfA_R	TGCTACTTCGTTGTCCCATGAC
clfB	clfB_F	AATGTGTTACCACTTTGATTAGGGTCAA
	clfB_R	GCTGCTGATGCTAAAGGTACAAATG
fnbA	fnbA_F	AACCATTATCCCAAGTTAAGGTATATCCTC
	fnbA_R	TGGTACTGATGAAGTTGATTTTAGAACACA
IL-6	hIL-6_F	AATTCGGTACATCCTCGACGG
	hIL-6_R	GGTTGTTTTCTGCCAGTGCCT
TNF	hTNF_F	TGGCCCAGGCAGTCAGA
	hTNF_R	GGTTTGCTACAACATGGGCTACA
IL-1β	hlL-1β_F	GCCCTAAACAGATGAAGTGCTC
	hIL-1β_R	GAACCAGCATCTTCCTCAG
CXCL8	hIL-8_F	GAGCACTCCATAAGGCACAAA
	hIL-8_R	ATGGTTCCTTCCGGTGGT
LL37	hLL37-F	TCGGATGCTAACCTCTACCG
	hLL37-R	GTCTGGGTCCCCATCCAT
beta-Actin	hbetaAct_F	CTCTTCCAGCCTTCCTTCCT
	hbetaAct_R	AGCACTGTGTTGGCGTACAG

63 Supplementary Table 3. qRT-PCR and *agr*-sequencing primers used in this study



Supplementary Figure 1. Amino acid sequence variation in AgrC results in differential 66 phenotypic response profiles of clinical ST22 S. aureus strains. (a) Amino acid sequence 67 analysis of AgrC from indicated strains at specified positions. (b) Size exclusion profile of 68 AgrC_{Y223C} and wild-type AgrC. The chromatograms reveal that the AgrC_{Y223C} protein is prone 69 to aggregation and elutes in the void volume of the experiment. (c) DLS profile for wild-type 70 AgrC_t with increasing H_2O_2 . (d) DLS profile for AgrC_{Y223C} with increasing H_2O_2 concentration. 71 Addition of H_2O_2 seemed to drastically increase the aggregation of AgrC_{Y223C} protein. (e) 72 Representative images of hemolysis on blood agar plates induced by indicated clinical and 73 respective mutant S. aureus strains. 74



Supplementary Figure 2. Representative images of Western Blot analyses of (**a**) TNF-R1 and (**b**) Caspase 1 subunits expression after 2 h (left panel) and 4 h (right panel) of keratinocytes infection. (**c**) β -Actin was used as a loading control (n=4). Uncropped full-size blots are shown.



81

Supplementary Figure 3. Cytotoxic strains induce caspase 1 activation in infected keratinocytes. Representative single channel images of Caspase 1 (green; left panel) secretion and DAPI (blue; right panel) after 2 h of keratinocytes monolayer infection with indicated strains. The scale bars in images equal 60 µm.



Supplementary Figure 4. *S. aureus* strains exhibiting colonizing phenotype reside within
phagolysosome in infected Keratinocytes. Representative single channel images of LAPM1
(red; center panel), DAPI (blue; right panel) and *S. aureus* (green; left panel) co-localization
in infected keratinocytes monolayers after 2 h of infection. The scale bars in images equal
60 μm.



Supplementary Figure 5. *S. aureus* strains exhibiting colonizing phenotype reside within phagolysosome in infected skin tissue models. Representative single channel images of LAPM1 (red; center panel), DAPI (blue; right panel) and *S. aureus* (green; left panel) colocalization in infected skin tissue models after 24 h of infection. The scale bars in images equal 60 µm.



Supplementary Figure 6. S. aureus strains exhibiting colonizing phenotype induce
autophagy in infected Keratinocytes. Representative single channel images of LC3AB (red;
center panel), DAPI (blue; right panel) and S. aureus (green; left panel) co-localization in
infected keratinocytes monolayers after 2 h of infection. The scale bars in images equal
60 μm.



Supplementary Figure 7. S. aureus strains exhibiting colonizing phenotype induce
autophagy in infected skin tissue models. Representative single channel images of LC3AB
(red; center panel), DAPI (blue; right panel) and S. aureus (green; left panel) co-localization
in infected skin tissue models after 24 h of infection. The scale bars in images equal 60 µm.

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- 136