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Effects of grass roots on soil-water retention curve and permeability function

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The effects of Vetiver grass (*Chrysopogon zizanioides*) roots on soil-water retention curves (SWRC), permeability (k) function and saturated permeability, k_{sat} , have been investigated on clayey sand (SC), and low plasticity silt (ML). For ML soil, when the root biomass per soil volume was lower than 6.5 kg/m³, the saturated permeability increased, the air-entry suction slightly reduced and the SWRC became steeper with increasing root contents, probably due to the formation of cracks due to wetting & drying cycles during plant growing period. Nevertheless, roots appeared to reduce the saturated permeability and increase the air-entry suction of ML soil, after reaching this threshold with root content of about 6.5 kg/m³ as roots occupied the macro-pores and tended to suppress cracks and swelling. For SC soil for all root contents, only slight variation of the saturated permeability with root content can be observed for the upper bounds on saturated permeability. However, the lower bounds on saturated permeability appeared to be less significant.

Author Keywords: soil-water retention curve, permeability function, grass root, soil bioengineering

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Introduction

Vegetation has been used to prevent shallow slides and erosion in various geotechnical and geo-environmental structures, such as embankments, cover systems of landfill and mine facilities. The technique has received much attention worldwide due to its relatively low cost, aesthetic and environmental value, as well as sustainability (e.g. Coppin et al. 1990; Gray and Sotir 1996; Goldsmith et al. 2014; Bo et al. 2015). Greenway (1987) summarized various hydro-mechanical influences of vegetation on slope stability, suggesting that vegetation can exert both beneficial and adverse effects. One of the major benefits of vegetation on slope stability is through root reinforcement of soil, which is considered as additional soil strength via root cohesion (e.g. Wu et al. 1979; Nilaweera and Nutalaya 1999; Leung et al. 2015a; Jotisankasa and Taworn 2016). Vegetation can improve slope stability further through transpiration process, which in turn depletes soil moisture, increasing the soil suction and shear strength further (e.g. Indraratna et al. 2006; Ng et al. 2014).

The effects of vegetation roots on hydraulic properties of soil, namely soil-water retention curves (SWRC), and permeability, have been investigated by some researchers. Researchers from geotechnical engineering discipline (e.g. Huat et al. 2005, 2006; Rahardjo et al. 2014; Leung et al. 2015b, 2015c) have shown that actively growing plants could reduce infiltration rate and saturated permeability of the soil. They demonstrated grass roots tended to increase the air-entry suction and the size of hysteresis loop of SWRC. These studies highlighted the benefits of vegetation in reducing infiltration and improving slope stability.

Past research in the fields of soil science and ecology (Barley 1954; Li and Ghodrati 1994; HilleRisLambers 2001; Rachman et al. 2004; Scanlan and Hinz 2010) also suggested that, when actively growing, roots tended to block the macro-pores. Yet, they also showed that decayed roots would create more macro pore and preferential flow path. Rachman et al. (2004) demonstrated that soil from a grass hedge system, of more than 10 years old, had the

saturated permeability of 6 to 18 times that of soil away from the hedge. These studies from soil science and ecology discipline generally highlighted the benefits of vegetation in increasing infiltration, reducing run-off, and improving quality of ecosystem in long term (e.g. Styczen and Morgan 1995). Age of roots, together with its life span and decay stage, plays an important role. Only a snapshot study at one particular root content and age may be inadequate to predict the performance of bio-engineered slope over the whole design period. Information on hydraulic properties of the soil with different root contents is very important to better predict the performance of bio-engineered slopes using numerical analyses (e.g. Rahimi et al. 2010; Rahardjo et al. 2014; Jotisankasa et al. 2014).

The purpose of this paper is to experimentally investigate the effects of vetiver grass (*Chrysopogon zizanioides*) on soils' hydraulic properties, namely soil-water retention curves (SWRC), and the permeability function. Two major soil types (sandy and silty) with different root contents were tested. *Chrysopogon zizanioides* has been widely used for bio-engineering purposes in many tropical and sub-tropical countries in South East Asia, China, Africa, South America, as well as warmer parts of the United States, Australia and Europe. Its applications range from soil-water conservation and slope stabilization (e.g. Grimshaw 1994; Hengchaovanich 1998; Truong et al. 2008; Jotisankasa et al. 2015a), as well as pollution-control at landfills and wetlands (e.g. Roongtanakiat et al. 2003; Raharjo et al. 2015; Delis et al. 2015). Despite vetiver being one of the most widely used grasses in geotechnical and geo-environmental engineering, the study of its root effects on soil-water retention curve and permeability function is very rare.

Materials

Two major soil types were selected for investigation, namely low plasticity Silt (ML), and clayey Sand (SC). The ML soil was a residual soil derived from weathered argillaceous rock (Shale and Mudstone). It was collected from a 26° slope area of fruit plantation in Uttaradit province, Northern Thailand. The vicinity of this area suffered from wide-spread shallow landslides in 2006 (ADPC 2006; Jotisankasa and Tapparnich 2010; Jotisankasa et al. 2015b). The SC soil was a completely weathered rock taken from upper part of a landslide that took place in 2011 in 1:1 cut slope of sandstone interbedded with siltstone in Ban-Natum, Suratthani, Southern Thailand (Tansamrit and Suanburi 2012; Jotisankasa et al. 2015b). These two soils represent typical materials encountered in most of bio-engineered slopes in the tropics. Their basic properties are summarized in Table 1. Tests on soil nutrients revealed that the SC soil, a completely weathered rock taken from a cut slope, was less fertile than the ML soil.

The soil specimens (10cm in diameter and 15 cm in height) were formed by static compaction in one layer at 80% standard Proctor maximum dry density and the optimum moisture content as summarized in Table 1. This choice of density was selected following Goldsmith et al. (2001) and Gray (2002). They suggested that an 80-85% Standard Proctor compaction provided a compromise between requirements of shear strength and plant growth. As-compacted suction of each soil sample was also measured using miniature tensiometer and found to be above 90 kPa for all three soils. The soil specimen's diameter of 10cm was chosen such that it represented the approximate spacing between Vetiver grass clumps initially grown in hedgerows in practice. Transparent acrylic molds were used to hold the samples so root patterns could be observed directly on the mold's side. The observation of side root content can be linked to the field measurement of root using mini-rhizotron technique (e.g. Rasse et al. 2000; Rewald and Ephrath 2013; Jotisankasa et al. 2015a). Each

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of these samples was planted with three clumps of *Chrysopogon zizanioides* of *Surathanni* variety. No fertilizer was added to the soil. A small hole of less than 5 centimeter depth was dug in the middle of the soil specimen for growing the grass. The soil dug from this hole was backfilled around the grass clumps and on top of soil specimen after planting. The grasses were allowed to grow until reaching a certain age, varying from four to ten months before testing. Slight variation in quality of each grass tiller may still exist, and this may result in samples of the same growing period, yet having different root contents. Therefore, the aim of this study was to relate the hydraulic properties with root biomass content and not with the grass age.

The initial water content during planting was in the optimum condition as identified from Standard Proctor compaction test as summarized in Table 1 (i.e. 13.1% for ML and 14% for SC). Irrigation procedure following Truong et al. (2008) was adopted. Daily irrigation was provided during the first 15 days of growing, after which watering was done once in several days. Aluminum foil was wrapped around the transparent acrylic tube to prevent direct sunlight onto the root zone. Each sample's base was underlain by a sponge layer during plant growing period, thus allowed to drain freely. On several separate specimens, suction measurement using miniature tensiometers showed that the suction in specimen during planting varied from 2 to 80kPa.

Testing methods

Once the root area, as appeared on the mold's side, reached a desired amount ranging from about 5 to 15% of total area, the specimen was considered ready for hydraulic tests. All the grass's leaves were then trimmed to minimize plant transpiration during test. The test program involved first soaking the sample under water for about 4 days before the drying Instantaneous Profile (IP) test, whereby soil suction increased from zero to about 100 kPa at

the end of test. After the completion of IP test, the sample was re-soaked for 4 days prior to the saturated permeability test. This testing sequence was necessary since the saturated permeability test involved additional trimming all the grass stump on top of the sample to ensure better flush surface and sealing all the gaps on sample mold with silicone sealant. Such manipulation during saturated permeability test made it more difficult to perform the IP test afterward. As a result, the drying IP test was carried out prior to the saturated permeability test.

The drying IP test was used for determining the SWRC and unsaturated permeability function. The IP method is a transient testing technique whereby flow rate through a soil specimen (or a soil profile) varies with time during test (e.g. Hamilton et al. 1981; Krisdani et al. 2009; Ng and Leung 2012). Profiles of water content and suction at several points along the specimen are normally monitored at fixed time intervals in order to compute the flow rate, hydraulic gradient and permeability at different suctions.

The experimental set up of the drying IP test in this study is shown in Figure 1. This set up was similar to the simplified evaporation IP method proposed by Peters and Durner (2008). The top surface of soil sample was left exposed to ambient air to induce upward onedimensional flow due to evaporation. The sample's base was underlain by an impermeable solid plate. Since all the grasses had been trimmed prior to test, no transpiration due to grass would take place and it was expected to be no localized increase in suction around the root zone. Typical results of a drying IP test are shown in Figure 2. Profiles of soil suction were monitored continuously using miniature tensiometers (Jotisankasa et al. 2007) at three locations on sample's side, as shown in Figure 2a and Figure 2e. The accuracy of tensiometer obtained during calibration was ± 0.15 kPa. The sample's weight was continuously measured using a data-logging balance (measuring to 0.01g precision), as shown in Figure 2c, in order to calculate the total flow rate across the sample.

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The values of suction at three locations were used to calculate the hydraulic gradient, *i*, as follows;

$$i = \frac{d(h_e - \frac{s}{\gamma_w})}{dz} \tag{1}$$

where h_e is the elevation head at each tensiometer location, *s* is matric suction, γ_w the unit weight of water and *z* the elevation as shown in Figure 1. The value of hydraulic gradient, *i*, presented in Figure 2b, was computed using linear regression over the three suction measurements as demonstrated in Figures 2e and 2f. All the values of R² appeared to be greater than 0.85 suggesting reasonable goodness of fit and linearity. It is noted that during evaporation, the relation between total head and elevation can be non-linear as the suction at the top increased beyond 100kPa. It was therefore considered the test results after top suction reached 100kPa unreliable and filter out those results from interpretation. As pointed out by Peters and Durner (2008) and later confirmed by Mairaing et al. (2012), linearization of the hydraulic gradient only lead to minor errors in hydraulic conductivity measurement, even in the late stage of evaporation where pressure head profiles are strongly non-linear.

The discharge velocity, *v*, at any particular time as presented in Figure 2d, was computed as follows;

$$v = \frac{dV_w}{A \cdot dt} \tag{2}$$

where dV_w is the change of volume of water in soil sample calculated from change in soil mass during test as shown in Figure 2c, A is the cross section area of sample, and t is the elapse time. Linear regression is used to calculate the slope between water volume change with time. The value of permeability at any suction and volumetric water content can then be calculated using Darcy's law,

$$k = \frac{v}{i} \tag{3}$$

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It is noted that drying IP test was carried out in this study as opposed to wetting IP test. Based on the current testing technique, results of wetting test have been shown by Mairaing et al. (2012) to be less accurate than drying test. This was due to larger non-linearity of the pore water pressure distribution with depth in wetting test.

The SWRC was determined by the drying IP test by plotting the calculated volumetric water content with suctions measured from three locations. It was also compared with the SWRC determined using the point-wise method (Mairaing et al. 2012). The point-wise method involved incremental measurement of soil suction at different moisture contents. After each suction measurement, the sample is either dried or wetted, depending on the test type, by about 1% water content in order to obtain suctions at different moisture contents. Drying is realized by exposing the sample in the air, while wetting is achieved by directly adding water into the sample using a very fine spray. After changing the moisture content to a desired amount, the sample was carefully wrapped and cured for equilibration of the suction throughout the sample. A minimum curing period of about 2-3 days between each increment was allowed for. It is noted that point-wise SWRC tests were carried out on compacted sample with 63mm diameter and 20mm height, which was smaller than the drying IP specimens. The smaller sample size was chosen to speed up the suction equilibration time and shorten the test duration.

Figure 3 compares the SWRCs obtained from point-wise tests with those from drying IP tests. The volumetric water contents were plotted against suctions from all three tensiometers in the drying IP tests. For ML sample, SWRCs from the three tensiometers appeared to be similar. For SC sample, the SWRC based on the middle tensiometer indicated a better agreement with the SWRC from point wise test. The SWRC based on the middle tensiometer was considered a reasonable representation of the SWRC of the whole sample

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and will be presented hereafter. In general, there is a good agreement between SWRCs from the drying IP test and point-wise test.

After the drying IP test was completed, the specimen was soaked under water again for 4 days, prior to the saturated permeability test. Upper part of the sample, about 5 mm in thickness, was then trimmed in order to have a flush top surface. The test procedure similar to the ASTM D2434 was followed to determine the saturated permeability. Prior to each test, the soil sample was flushed by flowing water of about 1.2meter water head, until no air bubble could be observed from the base outlet. During the test, the sample was subjected to pressure head of 1.4 meter. This choice of pressure head corresponds to the likely field condition where depth of the root zone may be limited to 1 to 1.5 meter with similar pore water pressure when fully submerged or infiltrated with rain water.

It is noted that the flexible wall method (ASTM D5084-10) could not be adopted in this study due to the difficulty in handling sample during rubber membrane sheathing without disturbing the roots. The constant head procedure was adopted for testing the sandy SC specimens, while the falling head technique was employed for the more fine-grained ML sample. Three orifices on the mold side, previously used for tensiometer installation, were plugged with plastic bungs and all the joints were sealed with silicone adhesive. Water tightness was carefully checked during saturated permeability testing.

Measurement of root content

In this study, root contents were directly measured in terms of "root biomass per soil volume", ρ_R , kg/m³. After completion of the saturated permeability test, the specimen was dismantled and the final moisture content of soil was determined. The remaining soil was washed with water and passed through Sieve #18 (1mm openings) to expose roots. Roots that were washed away and passed through Sieve #18 were considered small and not included in

root measurement. Figure 5 shows a specimen and its exposed roots. These roots were then dried in the oven at 105 ° C overnight to determine their dry biomass of roots, used for calculating the "root biomass per soil volume", ρ_R , kg/m³. This oven-drying temperature was in accordance with recommendation by Böhm (1979). Table 2 summarizes the root contents for all samples.

Phase relationships considering roots

Soil containing roots consists of materials in four phases, including solid grains, water, air, and roots. Since roots are of different specific gravity to the solid grains, the new phase relationships considering roots are defined as follows;

$$n = \frac{V_v}{V} = \frac{V - V_s - V_r}{V} = \frac{V - \frac{W_s}{G_s \cdot \gamma_W} - \frac{W_r}{G_R \cdot \gamma_W}}{V}$$
(4)

$$\theta = \frac{V_w}{V} = \frac{(W - W_r - W_s)}{V} \cdot \frac{1}{\gamma_w}$$
(5)

,where *n* is porosity, *V* is total volume, V_v is void volume, V_s is volume of soil particles, V_r is volume of roots, V_w is water volume, , *W* is total weight, W_s is weight of soil particles, W_r is weight of roots, γ_w is water unit weight, θ is volumetric water content, G_s and G_R are the specific gravity of the soil grain and root respectively. Designations of all phase variables are illustrated in Figure 5. The value of root density was assumed to be 641 kg/m³, according to Gray and Sotir (1996). These formulations were derived based on the fundamental phase diagram of multi-component materials (e.g. Landva and Clark 1990; Oweis and Khera 1998; Jotisankasa 2001). They give a better representation of those voids which are unoccupied by the roots, the parts which directly influences hydraulic properties of the soils. A similar formulation has been presented in Ng et al. (2016).

Effects of roots on saturated permeability

Figure 6 and Table 2 show variation of saturated permeability, k_{sat} , with root contents for the two soils. Trend lines and error bars for the permeability values have been drawn in Figure 6 corresponding to the 80% confidence interval. In other words, about 80% of the test result population is expected to lie within these two trendlines. This confidence interval was calculated based on parameters in Table 3 following the statistical method by Clewer and Scarisbrick (2001). Due to the limited population size (sample number, n =2-3), only 80% confidence interval could be proposed. It is noted a higher confidence level (e.g. 95%) could be achieved if more specimens were available. Those samples of similar root biomass content were considered as replicates belonging to the same group of treatment. Since permeability of most soils has been reported to have log-Normal statistic distribution (e.g. Benson 1993; Baecher and Christian 2003), the permeability data had been transformed to logarithmic values, and subsequently used for determining the standard error and the 80% confidence interval using the t-distribution, based on each group's sample size, n, as shown in Table 3.

As shown in Figure 6a, when the root biomass per volume, ρ_R , increased to about 6.5 kg/m³, the permeability of the ML soil appeared to increase up to 2 to 5 times the original permeability of soils without roots prior to grassing. This was expected to be due to desiccation cracks and preferential flow induced by drying & wetting cycles (Albrecht and Benson 2001; Li et al. 2016), as experienced during plant growth. As the root amount exceeded the threshold of 6.5 kg/m³, the permeability of ML soil decreased and became nearly one order of magnitude smaller when ρ_R was about 8 kg/m³. Roots would have significantly occupied macro voids at this stage.

For the SC soil (Fig. 6b), despite the scatter in data, the lower bound of k_{sat} value tended to decrease by 20-90% once the root biomass per volume, ρ_R , increased to about 6 kg/m³, probably due to root occupying the pores. The upper bound of k_{sat} value nevertheless

appeared to be relatively constant. No significant increase in permeability in this range of root contents was observed for the SC soil, probably due to its more coarse-grained nature and less desiccation shrinkage tendency. Interestingly, once the root content became more than 8 kg/m³ (Sample SC-1) the k_{sat} value of SC soil rose back to the same range of the soil without root. It was expected to be due to root decay, as no fertilizer was added it the specimen and its nutrients would have been depleted over the growing period. Depletion of the nutrients in the specimen would have induced the roots to be more active in searching for food, resulting in greater root content. As root decayed, preferential flow path along the impoverished roots may have happened. Since this observation was only based on one sample (SC-1), no confidence level can be drawn for this root content and the root decay explanation is not yet conclusive. Still, it is noteworthy that the SC soil had lower nutrients than the ML soil as shown in Table 1 and thus root decay would have started earlier for the SC soil.

Four main phenomena during plant growth that would have affected soils' permeability can be inferred. Firstly, cycles of wetting and drying due to plant watering and evapotranspiration would tend to increase the permeability. This effect is more significant for fine-grained material such as ML at early stage of plant growth. The second is the occupying of macro-pore by plant roots which tend to decrease the permeability. The third is the decay of roots which would generate preferential flow paths along the impoverished roots, thus increasing the permeability. The fourth would be related to organic carbon released by plant roots in the form of polymeric gel (the so-called root exudation, mucilage, or rhizodeposition) which were reported to decrease the mobility of clay colloids and enhance water retention around the roots (e.g. Grayston et al. 1997; Traoré et al. 2000; Kroener et al. 2014; Tian et al. 2015; Ghezzehei and Albalasmeh 2015). Recent work by Bengough (2012) shows that some of these root exudations exhibit hydrophobic or hydrophilic behavior, depending on the soil water content, causing the hysteretic-like behavior. It is difficult to quantify variation of each

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of these effects with time. Yet it can be said that there are both effects of increasing and decreasing saturated permeability due to the presence of roots during the course of plant life.

Effects of roots on soil-water retention curves

Figure 7 shows the SWRCs of ML specimens with various root contents. As explained previously, these curves were obtained using suction reading from the middle tensiometer which represented the average suction in the sample. The air-entry suction was determined from the intersection point of the horizontal line through soaked volumetric water content and the tangent to SWRC after desaturation point. These air-entry values corresponding to each SWRC are summarized in Table 2 and plotted in Figure 8 against root biomass. As root biomass increased up to 6.5 kg/m³, the air-entry suction of ML soils appeared to decrease slightly (from 2.3 to 0.7 kPa), probably due to micro-cracks induced by wetting & drying cycles. Specimens with micro-cracks would desaturate at a lower suction than those without. The micro-crack explanation is also supported by the corresponding initial increase in permeability as shown in Figure 6, as explained earlier. This observation is similar to previous studies (e.g. Peng et al. 2007; Bodner et al. 2013; Ma et al. 2015), which revealed the increase in large pore induced by wetting & drying cycles as a consequence of crack and micro-crack formation. After the threshold of root biomass, ρ_R , of about 6.5 kg/m³, was exceeded (Sample ML-1, ML-2), the air-entry suction appeared to increase slightly (from 0.7 to 1.8 kPa) perhaps due to the occupying of pores by the roots. The soaked volumetric water content, θ_s , is defined as the water content of the specimen after soaking for 4 days prior to the IP drying test. It is noted that soaking specimens for 4 days may not necessarily bring the sample to full saturation ($S_r = 100\%$) due to the presence of occluded air that requires positive pressure to dissolve (Hillel 1998; Lu et al. 2014). Therefore, the soaked volumetric water content, θ_s , is slightly less than the porosity, n. Nevertheless, it is very certain that there was no suction in soil specimens after 4 days of soaking, as confirmed by the tensiometer reading at the start of drying IP tests. Expectedly, as positive pore water pressure increases, the volumetric water content will increase from the soaked condition towards the fully saturated condition or the value of porosity. This behavior is of great practical significance for prediction of rainfall-induced landslide or embankment failure due to infiltration or groundwater rise.

Figure 8 shows the variation between porosity and soaked volumetric water content with root biomass, as calculated from measured volume and mass of soil and roots using Equations 4 and 5. As root biomass increased and occupied more void space, the porosity decreased accordingly. The soaked volumetric water content, θ_s , also decreased but remained close to the porosity, (i.e. degree of saturation in soaked condition being nearly 100%) up to root biomass, ρ_R , of about 8 kg/m³. At this value of ρ_R , the soaked volumetric water content, θ_s , became lower than the porosity, indicating more occluded air in the sample with high root content.

Figure 9 shows the SWRCs of clayey Sand (SC) with various root contents, obtained from the middle tensiometer. The variation between porosity and soaked volumetric water content, θ_s , with root content, as calculated from measured volume and mass of soil and roots using Equations 4 and 5, are shown in Figure 10. Again, porosity appeared to decrease with root content. Some increase in the soaked volumetric water content, θ_s , with roots could be observed for Samples SC-3 and SC-4. The observed increase in θ_s is similar to findings of previous SWRC tests on sandy materials with roots by Rahardjo et al. (2014) and Leung et al. (2015). This increase in θ_s of these samples with roots may be attributed to root exudation. As explained earlier, the compound exuded around the roots tends to enhance water retention (i.e. increase in volumetric water content near saturation point in Figure 9) and decrease the mobility of clay colloids (i.e. decrease of permeability as shown in Figure 6). Once the root biomass exceeded 8 kg/m³ (i.e. Sample SC-1), the value of θ_s however reduced, suggesting more presence of occluded air, possibly due to the more hydrophobic behavior of the exudation. At desaturation, SWRC of Sample SC-1 became noticeably steeper than the bare sample. This may be explained using the root decay hypothesis, previously put forward to explain the corresponding increase in saturated permeability. As roots became impoverished, and spaces were left around them, the pore size should become more uniformly distributed and the SWRC would be steeper. The air entry values, as presented in Figure 12 and Table 2, nevertheless appeared to be relatively unchanged as root biomass increased.

Effects of roots on permeability functions

Figures 11 and 12 shows permeability functions of the ML and SC specimens with different root contents respectively. Again, these curves were obtained from the middle tensiometer which represented the average suction in the sample. The saturated permeability, k_{sat} , obtained from the conventional test (as shown previously in Figure 6) was also plotted in the same graph as the k-value at suction 0.1 kPa for convenience. Interestingly, there is significant difference between the k-value at near saturated condition determined by IP test (referred herein as k_{1kPa}) and k_{sat} , determined by the conventional test. The values of k_{1kPa} , are defined as the permeability at suction range between 0.1 and 1 kPa in Figures 11 and 12, which correspond to degree of saturation between 90 and 100%, hence being close to soaked condition. The difference between the values of k_{1kPa} and k_{sat} could be due to a combination of factors, i.e. the permeability dependency on positive pore pressure and effective stress (e.g. Leroueil and Hight 2013); the less accuracy of the permeability determined by IP test at small suction (Peters and Durner 2008); as well as the presence of air-bubbles and their dissolution with testing time (Snehota et al. 2015). Figure 12 shows that biomodality of permeability function, or the double s-curve, is evident in Sample SC-2, where k_{sat} was similar to k_{1kPa} . This trend was not observed for Samples SC-3 and SC-4 even though they were of similar root content. The reason for this is unclear but expected to be due to plant variation which could also cause significant change in permeability at small range of suction. It is speculated that the root content of SC-2, SC-3, and SC-4 was around the threshold at which root effect started to impede water flow. The effect of root may be abrupt, i.e. with a minor increase of root content, the water flow was blocked. This range of suction (0.1 to 1 kPa) corresponds to macro pore size range from 1.5 to 0.15 mm which was about the size of grass roots. Distribution of these macro pores would be affected by roots to a greater extent than other smaller size pores. For the smaller pore size, corresponding to higher suction of 30 kPa, the k-function appeared to converge indicating less influence of roots on these void classes.

Conclusion

Grass roots can affect soil's hydraulic properties in a number of ways. Previous research showed that actively growing roots could reduce saturated permeability while decayed root would cause preferential flow path, increasing permeability. In this study, the effects of Vetiver grass (*Chrysopogon zizanioides*) on soil-water retention curves (SWRC), permeability (k) function and saturated permeability, k_{sat} , have been investigated on two soil types; low plasticity silt (ML) and clayey sand (SC), compacted at 80% standard Proctor density.

Grass roots have been shown to affect the hydraulic behavior of soils more significantly in the range of macro pore size greater than about 1.5mm, corresponding to suction less than 1 kPa. For ML soil, when the root biomass per soil volume were less than 6 kg/m³, the saturated permeability increased, the air-entry suction reduced and the SWRC

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became steeper with increasing root contents, probably due to the formation of cracks due to wetting & drying cycles during plant growing period. Nevertheless, roots appeared to reduce the saturated permeability and slightly increase the air-entry suction of ML soil, after reaching this threshold with root content of about 6.5 kg/m³ as roots occupied the macropores and tended to suppress cracks and swelling. For SC soil for all root contents, only slight variation of the saturated permeability with root content can be observed for the upper bound on saturated permeability. However, the lower bound on saturated permeability appeared to decrease as root biomass increased. It has also been shown that for suction range beyond 30 kPa, the influence of roots on permeability appeared to be less significant.

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Figure Captions

Figure 1 Instantaneous Profile (IP) drying test

Figure 2 Typical test result of drying IP test (Sample ML-3, root biomass per soil volume = 5.59 kg/m^3)

Figure 3 Drying SWRCs from IP drying tests and point-wise SWRC tests, on samples with

no root (a) low plasticity Silty soil, ML, and (b) clayey Sand, SC

Figure 4 Example of a specimen and exposed roots

Figure 5 Phase diagrams for soil containing roots

Figure 6 Variation of saturated permeability with root content of (a) Low plasticity Silt, and

(b) clayey Sand (Trendlines and error bar correspond to 80% confidence interval)

Figure 7 Soil-water retention curves of silt samples (ML) with and without roots (numbers in

brackets indicate root biomass per soil volume)

Figure 8 Variation of porosity soaked volumetric water content and air-entry suction with root biomass for low plasticity Silt (ML)

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Figure 9 Soil-water retention curves of clayey sand (SC) with and without roots (numbers in brackets indicate root biomass per soil volume)

Figure 10 Variation of porosity, soaked volumetric water content and air-entry suction with root biomass for clayey sand (SC)

Figure 11 Permeability functions of low plasticity Silt (ML) with and without roots (numbers in brackets indicate root biomass per soil volume)

Figure 12 Permeability functions of clayey Sand (SC) with and without roots (numbers in

brackets indicate root biomass per soil volume)





Figure 1 Instantaneous Profile (IP) drying test

197x138mm (300 x 300 DPI)



Figure 2 Typical test result of drying IP test (Sample ML-3, root biomass per soil volume = 5.59 kg/m3) 405x395mm (300 x 300 DPI)



Figure 3 Drying SWRCs from IP drying tests and point-wise SWRC tests, on samples with no root (a) low plasticity Silty soil, ML, and (b) clayey Sand, SC

296x378mm (300 x 300 DPI)



Figure 4 Example of a specimen and exposed roots

182x120mm (300 x 300 DPI)



Figure 5 Phase diagrams for soil containing roots

89x60mm (300 x 300 DPI)



Figure 6 Variation of saturated permeability with root content of (a) Low plasticity Silt, and (b) clayey Sand (Trendlines and error bar correspond to 80% confidence interval)

255x355mm (300 x 300 DPI)



Figure 7 Soil-water retention curves of silt samples (ML) with and without roots (numbers in brackets indicate root biomass per soil volume)

254x178mm (300 x 300 DPI)



Figure 8 Variation of porosity soaked volumetric water content and air-entry suction with root biomass for low plasticity Silt (ML)

287x186mm (300 x 300 DPI)



Figure 9 Soil-water retention curves of clayey sand (SC) with and without roots (numbers in brackets indicate root biomass per soil volume)

259x182mm (300 x 300 DPI)



Figure 10 Variation of porosity, soaked volumetric water content and air-entry suction with root biomass for clayey sand (SC)

277x173mm (300 x 300 DPI)



Figure 11 Permeability functions of low plasticity Silt (ML) with and without roots (numbers in brackets indicate root biomass per soil volume)

259x171mm (300 x 300 DPI)



Figure 12 Permeability functions of clayey Sand (SC) with and without roots (numbers in brackets indicate root biomass per soil volume)

253x180mm (300 x 300 DPI)

Test	Test standard/Method	Material		
1031	i est standard/ivietnou	ML	SC	
Grain size distribution				
Gravel \leq 75 mm, %		0	31.4	
Sand \leq 4.75 mm, %	ASTM D422-63	20.2	45.7	
Silt \leq 75 μ m, %	ASIM DI140	51.8	15.4	
Clay \leq 2 μ m, %		28.0	7.5	
Specific gravity, G _s	ASTM D854-02	2.65	2.69	
Atterberg's Limits				
Liquid limit, LL, %		39.3	31.2	
Plastic limit, PL, %	AS1M D4318-10	29.4	19.4	
Plasticity index, PI, %		9.9	11.8	
Soil nutrients	Wet oxidation	1.95 (medium)	0.69 (very low)	
Organic content, %		, , ,		
рН	pH meter (Soil:Water, 1:1)	6 (highly acidic)	5 (highly acidic)	
P, phosphorous, mg/kg	Bray II (modified)	17 (medium)	3 (low)	
K, potassium, mg/kg		107 (high)	64 (low)	
Ca, calcium, mg/kg	Atomic Absorption	1509 (high)	427 (medium)	
Mg, magnesium, mg/kg	Spetrophotometer	216 (high)	69 (medium)	
Standard Proctor compaction test				
Maximum dry density, kN/m ³	ASTM D698 - 12e2	16.4	17.6	
Optimum water content, %		18.9	15.4	
As-compacted dry density,		13.1	14.0	
kN/m ³	ASTM D698 - 12e2			

Table 1 Basic Soil Properties

Soil type	Test no.	Age of plant, days	Root biomass per unit volume of soil, kg/m ³	Saturated permeability, m/s	Air-entry suction, kPa	IP test duration, days
	ML-a	NA	0	2.02E-06	2.3	12
	ML-b	NA	0	1.32E-06	-	-
Low- Plasticity Silt, ML	ML-c	NA	0	1.12E-06	-	-
	ML-1	143	7.14	3.25E-07	1.6	19
	ML-2	301	7.99	3.46E-07	1.8	10
	ML-3	302	5.59	4.43E-06	2.0	11
	ML-4	317	6.50	7.75E-06	0.7	15
Clayey Sand, SC	SC-a	NA	0	7.39E-06	0.7	9
	SC-b	NA	0	6.14E-06	-	-
	SC-c	NA	0	3.77E-06	-	-
	SC-1	177	8.69	9.47E-06	0.8	21
	SC-2	222	5.85	5.80E-06	0.9	24
	SC-3	303	5.49	3.68E-06	0.9	13
	SC-4	60	5.71	4.02E-07	0.8	14

 Table 2 Description of tests and their results

Test group	Root biomass	Standard	Sample number,	Standard error,
	per unit			
	volume of soil,	deviation of	n	S
	kg/m ³			\sqrt{n}
		$\log-k(m/s), s$		
ML-a, ML-b, ML-c	0	0.132	3	0.0762
ML-1, ML-2	7.14 to 7.99	0.019	2	0.0136
ML-3, ML-4	5.59 to 6.5	0.172	2	0.1214
SC-a, SC-b, SC-c	0	0.151	3	0.0872
SC-2, SC-3, SC-4	5.49 to 5.85	0.620	3	0.3580

Table 3 Statistical parameters used for confidence interval calculation

